PARASITIC INFECTIONS
OF MAN AND ANIMALS
IN HAWAII

Joseph E. Alicata
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FOREWORD

Parasites probably were introduced into Hawaii with the first colonization by man perhaps fifteen hundred or more years ago. However, parasitism appears not to have been important or at least not recognized until about 1800 when European and American ships began to call frequently. Since that time, parasites have been found in many species; for instance, in birds, including chickens, turkeys, pigeons, pheasants, doves, ducks, sparrows, herons, coots, and quails, and in mammals, including mice, rats, mongooses, rabbits, cats, dogs, pigs, sheep, cattle, horses, and man. There is a certain uniqueness in the compressed history of the infestations paralleling the sweeping spread of virus diseases when introduced into new territories.

The reports of these parasitic diseases have heretofore been widely scattered in the literature, and Professor Alicata's publication now provides an orderly and systematic presentation of the entire field. He considers in sequence the considerable number of diseases reported to be caused in Hawaii by protozoa, the very large number caused by nemathelminthes, and the smaller group caused by platyhelminthes. This publication will furnish basic information for future parasitologists who in turn will be immensely grateful.

WINDSOR C. CUTTING, M.D.
Director

University of Hawaii
Pacific Biomedical Research Center
Honolulu, Hawaii, U.S.A.

November 1964
CONTENTS

INTRODUCTION .......................................................... 5

CLASSIFICATION OF INTERNAL PARASITES OF MAN AND ANIMALS IN
HAWAII ................................................................. 7
   Phylum: Protozoa .................................................. 8
   Phylum: Nemathelminthes (Roundworms) ..................... 9
   Phylum: Platyhelminthes (Tapeworms and Flukes) ........ 13

THE LIFE CYCLES OF PARASITES .................................... 15
   Protozoa .......................................................... 15
   Roundworms ....................................................... 17
   Tapeworms ....................................................... 20
   Flukes ............................................................ 21
   Arthropods ....................................................... 22

ROUTES OF INFECTION OF INTERNAL PARASITES ............... 23

MOLLUSKS OF PARASITOLOGICAL IMPORTANCE IN HAWAII ....... 24

PARASITES OF MAN .................................................... 27

PARASITES OF ANIMALS ............................................... 43
   Cat ............................................................... 43
   Cattle ........................................................... 46
   Chicken and Other Avian Hosts ............................... 57
   Deer ............................................................. 71
   Dog ............................................................... 71
   Goat ............................................................. 75
   Horse ............................................................ 75
   Mongoose ........................................................ 77
   Mouse ............................................................ 78
   Rabbit ............................................................ 79
   Rat ............................................................... 80
   Sheep ............................................................. 86
   Swine ............................................................. 87

HOST LIST OF PARASITES AND INTERMEDIATE HOSTS RECORDED FROM
HAWAII ................................................................. 95

BIBLIOGRAPHY ........................................................ 119

INDEX ................................................................. 132
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THE AUTHOR

JOSEPH E. ALICATA, Ph.D., is Parasitologist at the Hawaii Agricultural Experiment Station, College of Tropical Agriculture, University of Hawaii. He has been Head of the Department of Parasitology from 1935 to the present (1964). He has served as Senior Scientist (R), U. S. Public Health Service, since 1950. He was Junior Zoologist for the U. S. Department of Agriculture, Washington, D.C., from 1928 to 1935; conducted parasitological investigations for the Hawaii Territorial Board of Health (1936–37), for the Health Committee of the Honolulu Chamber of Commerce (1941–42), and for the Hawaiian Sugar Planters' Association (1943). In addition to Hawaii and other parts of the United States, he has carried out parasitological research in Europe, the Middle East, and several island groups of the Pacific including Polynesia, Melanesia, and Micronesia.

The major fields of research by the author have included the following: life history and control of livestock parasites; use of X-irradiation in pork for the control of human trichinosis; use of hexachloroethane for the control of bovine fascioliasis; survey of parasites of man and animals in the Pacific islands; epidemiology of typhus and leptospirosis in Hawaii; search for evidences pertaining to the theory first proposed by the author, that Angiostrongylus cantonensis may be the causative agent of eosinophilic meningencephalitis of man in the Pacific.
PARASITIC INFECTIONS OF MAN AND ANIMALS IN HAWAII*

JOSEPH E. ALICATA

INTRODUCTION

There are no indications that parasites of man and animals were known or recognized by the early Hawaiians. Although it is likely that the first parasites in the Hawaiian Islands were introduced by migratory birds hundreds of years ago, in all probability most of those now present postdate the discovery of the Islands by Captain Cook in 1778. Since that time there has been in Hawaii a gradual influx of a variety of people, animals, and plants, largely from North America and Asia. All of these importations have resulted in the introduction of many species of parasites and their vectors. To what extent new ones will enter the Hawaiian Islands, especially with the expansion of modern air and ocean transportation, remains to be determined. Much is being done, however, through inspection, quarantine, and other State, Federal, and private agricultural regulatory measures, to prevent the introduction of additional disease-producing organisms and their vectors.

The Hawaiian Islands proper, with which this report is concerned, include the islands of Kauai, Niilhau, Oahu, Maui, Molokai, Lanai, Kahoolawe, and Hawaii. Geographically, these islands represent the summit parts of the southeastern portion of the Hawaiian Archipelago, which extends in the mid-Pacific for a distance of about 2,000 miles. The mountains of these islands may be coastal, acentric, or centric in position, and they divide each island into windward and leeward sides. The windward areas are largely eroded to precipitous cliffs or penetrating valleys, and rainfall is greater there than on the leeward areas. This condition results in increased soil moisture, dense vegetation, and swampiness in the windward areas. Moreover, it

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favors the maintenance and dissemination of parasites, and the abundance of aquatic invertebrates which serve as vectors. On the other hand, the leeward areas do not show severe erosion, rainfall is less, and conditions are less favorable for the dissemination of parasites. However, the Islands do have an equable subtropical climate, and in areas below 2,500 feet, where most livestock are raised, the annual mean temperature is approximately 70° F. In general, therefore, topographical and climatic conditions in the Islands are highly favorable the year round for the propagation and spread of parasites and intermediate hosts.

Parasites have considerable veterinary and public health importance. In livestock raising, they may become so severe as to affect production and profits. Constant vigilance must therefore be maintained by the animal producer to see that infection is kept at a minimum. Parasitic diseases are often not spectacular, nor do they cause high mortality eventually. For this reason a stockman either may not be aware of their presence or may not place much importance on them. Frequently the major symptom associated with parasitism is that of general unthriftness. In most instances this is a slow and gradual process which does not attract attention. The losses usually result from slow and stunted growth, uneconomical use of feed, reduced production, condemnation of meat carcasses in whole or in part, and a waste of time and effort on the part of the owner in maintaining unthrifty and unproductive animals.

Under ordinary animal-raising conditions it is almost impossible to maintain livestock that are free of parasites. However, much can be done to prevent parasites from becoming sufficiently numerous to endanger the health of the animals. This calls for sanitary practices and other specific preventive measures. Drug treatment often plays an important part in parasite control. Destruction of the parasites in the host assists not only in improving the health of the animal but also in reducing pasture or ground contamination. Drug treatment, however, should not be considered as the only means of parasite control, but as an adjunct to good sanitary and management practices.

In the field of public health, awareness of the presence and spread of disease-producing organisms in a community is important in that it assists in the detection, treatment, and prevention of the causative agent. With the exception of enterobiasis, parasitic infections of man acquired in the Hawaiian Islands are comparatively uncommon, and there exist none of the debilitant cases, such as malaria, filariasis, and schistosomiasis, so often found in tropical areas. The low incidence of human parasitism in Hawaii is undoubtedly due to the average high economic, educational, and sanitary levels of the population, plus the active surveillance of the local medical and veterinary authorities, and related workers. Constant alertness by the popula-
tion in the use of preventive measures does go a long way in the control of parasitic infections which have become endemic. These include not only enterobiasis, but also angiostrongylosis, ascariasis, cysticercosis, fascioliasis, heterophyidiasis, toxocarosis, trichinosis, and trichiuriasis, all of which occasionally occur in the Islands.

Of further public health importance is the fact that several parasites of man are acquired from animals. Indeed it is possible that some of the obscure ailments of man are due to these parasites. In Hawaii, parasites which are transmissible directly or indirectly from animals to man include at least the following: *Toxocara canis* and *Dipylidium caninum* from cats and dogs; *Fasciola gigantica* from cattle; *Angiostrongylus cantonensis, Capillaria hepatica,* and *Hymenolepis nana* from rats; *Taenia solium* and *Trichinella spiralis* from swine. Consideration should be given also to the fact that many parasites of man and animals are transmitted by various pests such as insects, mollusks, and rats. Public awareness and active control of these pests in a community are, therefore, important in the control of parasitic diseases.

In this bulletin, an attempt has been made to summarize information by various authors concerning the internal and external parasites of man and animals in the Hawaiian Islands. A list of intermediate hosts, whenever they are present, is also included. For the most part these reports have been compiled noncritically and have been cited in the body of the text and in the bibliography. Greater emphasis has been given to those parasites which have special medical and veterinary importance. It has become evident during the preparation of this report that parasitological information is either inadequate or entirely lacking in a number of important areas. It is hoped that subsequent investigators will be able to enlarge this useful field of knowledge.

**CLASSIFICATION OF INTERNAL PARASITES OF MAN AND ANIMALS IN HAWAII**

The following is a taxonomically arranged list of internal parasites of land vertebrates reported from the Hawaiian Islands. The classification of the Protozoa is based on that used by Honigberg *et al.* (1964) and Levine (1961), and, for the Nematoda, Cestoda, and Trematoda, for the most part that used by Yamaguti (1958, 1959, 1961). The parasites are classified with reference to the phylum, class, order, family, subfamily (when known), genus, and species. The final host is indicated at the right.

1 All orders are designated by an asterisk (*), all families by a single dagger (*), all subfamilies by a double dagger (**); generic and specific names are italicized.
PHYLUM: PROTOZOA

Class: Zoomastigophorea (flagellates)

Diplomonadida* Wenyon, 1926

Hexamitidae† Saville Kent, 1880

Giardia lamblia (Stiles, 1915) Man

Kinetoplastida* Honigberg, 1963

Trypanosomatidae† Doflein, 1901

Trypanosoma conorhini (Donovan, 1909) Mouse or Rat (?)

Trypanosoma lewisi (Kent, 1880) Rat

Rhizomastigida* Doflein, 1916

Mastigamoebidae†

Histomomas meleagridis (Smith, 1895) Chicken, Turkey

Trichomonadida* Kirby, 1947

Trichomonadidae† Chalmers and Pekkola, 1918 emend.

Kirby, 1946

Pentatrichomonas hominis (Davaine, 1860) Man

Trichomonas gallinae (Rivolta, 1878) Pigeon

Class: Rhizopodea (amoebae)

Amoebida* Kent, 1880

Endamoebidae† Calkins, 1926

Dientamoeba fragilis Jepps and Dobell, 1918 Man

Endolimax nana (Wenyon and O'Connor, 1917) Man

Entamoeba coli (Grassi, 1879) Man

Entamoeba hartmanni Von Prowazek, 1912 Man

Entamoeba histolytica Schaudinn, 1903 Man

Class: Telosporia (coccidia and haemosporidia)

Eucoccida* Léger and Duboscq, 1910

Eimeriidae† Poche, 1918

Eimeria bovis (Zublin, 1908) Cattle

Eimeria bukidnonensis Tubangui, 1931 Cattle

Eimeria cylindrica Wilson, 1931 Cattle

Eimeria debliecki Douwes, 1921 Swine

Eimeria scabra Henry, 1931 Swine

Eimeria spinoza Henry, 1931 Swine

Eimeria stiedae (Lindemann, 1865) Rabbit

Eimeria tenella (Railliet and Lucet, 1891) Chicken

Eimeria zurnii (Rivolta, 1878) Cattle
Plasmodiidae† Mesnil, 1903

*Haemoproteus columbae* Kruse, 1890

*Plasmodium vaughani* Novy and McNeal, 1904

Class: Microsporidea (microsporidia)

*Haemoproteus columbae* Kruse, 1890

*Plasmodium vaughani* Novy and McNeal, 1904

Class: Microsporida* Balbiani, 1882

Toxoplasmatidae†

*Encephalitozoon cuniculi* Levaditi, Nicolau, and Schoen, 1923

Class: Ciliata (ciliates)

*Toxoplasma gondii* Levine, 1908

Class: Ciliata (ciliates)

*Encephalitozoon cuniculi* Levaditi, Nicolau, and Schoen, 1923

Class: Toxoplasmatidae†

*Toxoplasma gondii* Levine, 1908

PHYLUM: NEMATHELMINTHES

Class: Nematoda (roundworms)

*Rhabdiasidae* Yamaguti, 1961

Strongyloidae† Chitwood and McIntosh, 1934

*Strongyloides papillosus* (Wedl, 1856)

*Strongyloides ransomi* Schwartz and Alicata, 1930

*Strongyloides ratti* Sandground, 1925

*Strongyloides stercoralis* (Bavay, 1876)

Ascarididea* Yamaguti, 1961

Ascarididae† Blanchard, 1849

*Ascaris lumbricoides* Linnaeus, 1758

*Parascaris equorum* (Goeze, 1782)

*Toxascaris leonina* (Linstow, 1902)

Ascaridiinae† Travassos, 1919

*Ascaridia galli* (Schrank, 1788)

*Toxocarinae* Yamaguti, 1961

*Toxocara canis* (Werner, 1782)

Oxyuridea* Weinland, 1858

Heterakisidae† Railliet and Henry, 1914

*Heterakis gallinarum* (Schrank, 1788)

*Heterakis spumosa* Schneider, 1866

FINAL HOST

Pigeon-fly

Mosquito (?)

Rabbit

Swine

Cattle

Swine

Rat

Man

Man, Swine

Horse

Dog

Chicken

Cat, Dog

Chicken, Pheasant, Turkey

Rat
<table>
<thead>
<tr>
<th>Family</th>
<th>Genus</th>
<th>Host(s)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Oxyuridae† Cobbold, 1864</td>
<td><em>Enterobius</em> vermicularis (Linnaeus, 1758)</td>
<td>Man</td>
</tr>
<tr>
<td></td>
<td><em>Oxyuris equi</em> (Schrank, 1788)</td>
<td>Horse</td>
</tr>
<tr>
<td></td>
<td><em>Passalurus ambiguus</em> (Rudolph, 1819)</td>
<td>Rabbit</td>
</tr>
<tr>
<td></td>
<td><em>Lauroinae† Skrjabin and Shikhobalova, 1951</em></td>
<td>Horse</td>
</tr>
<tr>
<td></td>
<td><em>Probstmayria vivipara</em> (Probstmayr, 1865)</td>
<td>Horse</td>
</tr>
<tr>
<td></td>
<td><em>Syphaciinae† Railliet, 1916</em></td>
<td>Rat</td>
</tr>
<tr>
<td></td>
<td><em>Syphacia obvelata</em> (Rudolphi, 1802)</td>
<td>Rat</td>
</tr>
<tr>
<td></td>
<td><em>Subuluridae† Yorke and Maplestone, 1926</em></td>
<td>Chicken, Pheasant</td>
</tr>
<tr>
<td></td>
<td><em>Subulura brunpfi</em> (López-Neyra, 1922)</td>
<td></td>
</tr>
<tr>
<td><em><em>Trichuridea</em> Yamaguti, 1961</em>*</td>
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<td></td>
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<tr>
<td>Trichinellidae† Ward, 1907</td>
<td><em>Trichinella spiralis</em> (Owen, 1835)</td>
<td>Man, Mongoose, Rat, Swine</td>
</tr>
<tr>
<td>Trichosomoididae† Yorke and Maplestone, 1926</td>
<td><em>Trichosomoides crassicaudaa</em> (Bellingham, 1840)</td>
<td>Rat</td>
</tr>
<tr>
<td>Trichuridae† Railliet, 1915</td>
<td><em>Capillaria hepatica</em> (Bancroft, 1893)</td>
<td>Man, Rat</td>
</tr>
<tr>
<td></td>
<td><em>Capillaria traversae</em> Ash, 1962</td>
<td>Rat</td>
</tr>
<tr>
<td></td>
<td><em>Trichurinae† Ransom, 1911</em></td>
<td></td>
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<tr>
<td></td>
<td><em>Trichurus ovis</em> (Abildgaard, 1795)</td>
<td>Cattle</td>
</tr>
<tr>
<td></td>
<td><em>Trichurus trichiura</em> (Linnaeus, 1771)</td>
<td>Man, Swine</td>
</tr>
<tr>
<td></td>
<td><em>Trichurus vulpis</em> (Froclich, 1789)</td>
<td>Dog</td>
</tr>
</tbody>
</table>

| Strongylidea* Diesing, 1851   |                              |                          |
| Ancylostomatidae† Nicoll, 1927 | *Ancylostomatinae† Nicoll, 1927* |                          |
|                               | *Ancylostoma caninum* (Ercolani, 1859) | Dog                     |
|                               | *Ancylostoma tubaeforme* (Zeder, 1800) | Cat                     |
|                               | *Bunostominae† Looss, 1911* |                          |
|                               | *Bunostomum phlebotomum* (Railliet, 1900) | Cattle                  |

| Cyathostomidae† Yamaguti, 1961 |                              |                          |
| Cyathostominae† Nicoll, 1927  | *Cyathostomum asymmetricum* Theiler, 1923 | Horse                  |
|                               | *Cyathostomum bicoronatum* Looss, 1900 | Horse                   |
|                               | *Cyathostomum calicatum* Looss, 1900 | Horse                   |
|                               | *Cyathostomum cattinatum* Looss, 1900 | Horse                   |
|                               | *Cyathostomum coronatum* (Looss, 1900) | Horse                   |
|                               | *Cyathostomum euproctum* (Boulenger, 1917) | Horse                  |
Parasitic Infections of Man and Animals in Hawaii

Final Host

Cyathostomum goldi (Boulenger, 1917) Horse
Cyathostomum insignis (Boulenger, 1917) Horse
Cyathostomum leptostomum (Kotlan, 1920) Horse
Cyathostomum longibursatum (Yorke and Maclie, 1918) Horse
Cyathostomum nassatum Looss, 1900 Horse
Cyathostomum pateratum (Yorke and Maclie, 1919) Horse
Gyalocephalus capitatus Looss, 1900 Horse
Poteriotomum impavidentatum Quiel, 1919 Horse

Oesophagostominae† Railliet, 1916
Oesophagostomum dentatum (Rudolphi, 1803) Swine
Oesophagostomum radiatum (Rudolphi, 1803) Cattle

Stephanuridae† Travassos and Vogelsang, 1933
Stephanurus dentatus Diesing, 1839 Swine

Strongylidea† Baird, 1853
Strongylinae† Railliet, 1893
Alfortia edentata (Looss, 1900) Horse
Delafondia vulgaris (Looss, 1900) Horse
Strongylus equinus Mueller, 1780 Horse
Triodontophorus brewanda Boulenger, 1916 Horse
Triodontophorus serratus (Looss, 1900) Horse

Globocephalinae† Travassos and Vogelsang, 1932
Globocephalus wroblewskii (Alessandrini, 1909) Wild pig

Trichostrongylidea† Leiper, 1912
Haemoncinidea† Yamaguti, 1961
Haemonchus contortus (Rudolphi, 1803) Cattle, Sheep

Nematodirinae† Skrjabin and Orloff, 1934
Nematodirus spathiger (Railliet, 1896) Sheep

Ornithostrongylinae† Travassos, 1937
Ovithiostrongylus quadiradiatus (Stevenson, 1904) Pigeon

Trichostrongylinae† Leiper, 1908
Cooperia pectinata Ransom, 1907 Cattle
Cooperia punctata (Von Linstow, 1907) Cattle, Dec;
Cooperia spatulata Baylis, 1938 Cattle
Hyostrongylus rubidus (Hassall and Stiles, 1892) Swine
Ostertagia circumcincta (Stadelmann, 1894) Goat
Ostertagia ostertagi (Stiles, 1892) Cattle
Trichostrongylus axei (Cobbold, 1789) Horse
Trichostrongylus colubriformis (Giles, 1892) Goat, Man, Sheep

Vianninae† Neveu-Lemaire, 1934
Nippostrongylus brasiliensis (Travassos, 1914) Rat
Protostrongylidae† Leiper, 1926

Angiostrongylineae† Bohm and Gebauer, 1934

*Angiostrongylus cantonensis* (Chen, 1935) Rat (immature parasites in man)

Dictyocaulinae† Skrjabin, 1933

*Dictyocaulus viviparus* (Block, 1782) Cattle

Metastrongylineae† Leiper, 1908

*Choeostrongylus pudendotectus* (Wostkow, 1905) Swine

*Metastrongylus elongatus* (Dujardin, 1845) Swine

Protostrongylineae† Kamensky, 1905

*Aelurostrongylus abstrusus* (Railliet, 1898) Cat

Pseudaliidae† Railliet, 1916

Filaroidinae† Skrjabin, 1932

*Anafilaroides rostratus* Gerichter, 1949 Cat

*Filaroides osleri* (Cobbold, 1879) Dog

*Spiruridea* Diesing, 1861

Acuariidae† Seurat, 1913

*Acuaria hamulosa* (Diesing, 1851) Chicken, Pheasant, Turkey

*Dispharynx nasuta* (Molin, 1858) Chicken

Physalopteridae† Leiper, 1908

Physalopterinae† Stossich, 1898

*Physaloptera muris-brasiliensis* Diesing, 1861 Rat

*Physaloptera praeputialis* Linstow, 1899 Cat

*Spiruridae* Oerley, 1885

Ascaropsinae† Alicata and McIntosh, 1933

*Ascarops strongyлина* (Rudolphi, 1819) Swine

*Physcocephalus sexalatus* (Molin, 1860) Swine

Gongylonematinae† Nicoll, 1927

*Gongylonema ingluvicola* Ransom, 1904 Chicken

*Gongylonema neoplasticum* (Fibiger and Dittersen, 1914)

*Gongylonema pulchrum* Molin, 1857 Cattle

*Spirurinae*† Railliet, 1935

*Cyrnea graphophasiani* (Yamaguti, 1935) Pheasant

*Habronema microstoma* (Schneider, 1886) Horse

*Habronema muscae* (Carter, 1961) Horse

Thelaziidae† Skrjabin, 1915

Oxyspirurinae† Yamaguti, 1961

*Oxyspirura mansoni* (Cobbold, 1879) Chicken, Dove, English sparrow, Mynah, Pheasant, Quail
<table>
<thead>
<tr>
<th>Class</th>
<th>FINAL HOST</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Tropisuridae</strong>† Yamaguti, 1961</td>
<td></td>
</tr>
<tr>
<td><em>Microtetrameris</em> sp.</td>
<td>Mynah</td>
</tr>
<tr>
<td><em>Tropisurus americanus</em> (Cram, 1927)</td>
<td>Chicken</td>
</tr>
<tr>
<td><em>Tropisurus</em> sp.</td>
<td>English sparrow</td>
</tr>
<tr>
<td><strong>Filariidea</strong>† Yamaguti, 1961</td>
<td></td>
</tr>
<tr>
<td><strong>Dipetalonematidae</strong>† Wehr, 1935</td>
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</tr>
<tr>
<td><strong>Dipetalonematinae</strong>‡ Wehr, 1935</td>
<td>Dog</td>
</tr>
<tr>
<td><strong>Dirofilarinae</strong>‡ Wehr, 1935</td>
<td>Cat, Dog</td>
</tr>
<tr>
<td><em>Dirofilaria immitis</em> (Leidy, 1856)</td>
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</tr>
<tr>
<td><strong>Stephanofilaridae</strong>† Wehr, 1935</td>
<td>Cattle</td>
</tr>
<tr>
<td><em>Stephanofilaria stilesi</em> Chitwood, 1934</td>
<td></td>
</tr>
<tr>
<td><strong>Phylum: Platyhelminthes</strong></td>
<td></td>
</tr>
<tr>
<td>Class: Cestoda (tapeworms)</td>
<td></td>
</tr>
<tr>
<td><strong>Cyclophyllidea</strong>* Beneden in Braun, 1900</td>
<td></td>
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<tr>
<td><strong>Anoplocephalidae</strong>† Cholodkovsky, 1902</td>
<td></td>
</tr>
<tr>
<td><strong>Anoplocephalinae</strong>‡ Blanchard, 1891</td>
<td>Horse</td>
</tr>
<tr>
<td><em>Anoplocephala magna</em> (Abildgaard, 1789)</td>
<td>Horse</td>
</tr>
<tr>
<td><em>Anoplocephala perfoliata</em> (Goeze, 1782)</td>
<td>Horse</td>
</tr>
<tr>
<td><em>Moniezia benedeni</em> (Moniez, 1879)</td>
<td>Horse</td>
</tr>
<tr>
<td><em>Moniezia expansa</em> (Rudolphi, 1805)</td>
<td>Horse</td>
</tr>
<tr>
<td><strong>Davaineidae</strong>† Furhmann, 1907</td>
<td></td>
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<tr>
<td><strong>Davaineinae</strong>‡ Braun, 1900</td>
<td></td>
</tr>
<tr>
<td><em>Raillietina cesticillus</em> (Molin, 1858)</td>
<td>Horse</td>
</tr>
<tr>
<td><em>Raillietina tetragona</em> (Molin, 1858)</td>
<td>Horse</td>
</tr>
<tr>
<td><em>Raillietina sp.</em></td>
<td>Horse</td>
</tr>
<tr>
<td><strong>Dilepididae</strong>† Railliet and Henry, 1909</td>
<td></td>
</tr>
<tr>
<td><strong>Dipylidiinae</strong>‡ Stiles, 1896</td>
<td></td>
</tr>
<tr>
<td><em>Choanotaenia infundibulum</em> (Block, 1779)</td>
<td>Horse</td>
</tr>
<tr>
<td><em>Dipylidium caninum</em> (Linnaeus, 1758)</td>
<td>Horse</td>
</tr>
<tr>
<td><strong>Hymenolepididae</strong>† Railliet and Henry, 1909</td>
<td></td>
</tr>
<tr>
<td><strong>Hymenolepidinae</strong>‡ Perrier, 1897</td>
<td></td>
</tr>
<tr>
<td><em>Cloacotaenia megalops</em> (Nitzsch in Creplin, 1829)</td>
<td>Hawaiian duck</td>
</tr>
<tr>
<td><em>Echinolepis carioca</em> (Magalhaes, 1898)</td>
<td>Chicken</td>
</tr>
<tr>
<td><em>Hymenolepis diminuta</em> (Rudolphi, 1819)</td>
<td>Rat</td>
</tr>
<tr>
<td><em>Hymenolepis nana</em> Siebold, 1852</td>
<td>Man, Rat</td>
</tr>
<tr>
<td><em>Hymenosphenacanthus exiguis</em> (Yoshida, 1910)</td>
<td>Chicken</td>
</tr>
<tr>
<td><strong>Taeniidae</strong>† Ludwig, 1886</td>
<td></td>
</tr>
<tr>
<td><em>Hydatigera taeniaeformis</em> (Batsch, 1786)</td>
<td>Cat</td>
</tr>
<tr>
<td><em>Taenia hydatigena</em> Pallas, 1766</td>
<td>Dog (see p. 74)</td>
</tr>
<tr>
<td><em>Taenia saginata</em> Goeze, 1782</td>
<td>Man</td>
</tr>
<tr>
<td><em>Taenia solium</em> Linnaeus, 1758</td>
<td>Man (see p. 37)</td>
</tr>
</tbody>
</table>
Class: Trematoda (flukes)

**Digenea***

- **Brachylaemidae**† Joyeux and Foley, 1930
  - **Brachylaeminae**† Joyeux and Foley, 1930
    - *Postharmostomum gallinum* (Witenberg, 1923)
      - Chicken

- **Dicrocoeliidae**† Odhner, 1911
  - **Dicrocoeliinae**† Looss, 1899
    - *Platyneosomum fastosum* Kossack, 1910
      - Cat

- **Echinostomatidae**† Poche, 1926
  - **Echinostomatinae**† Faust, 1929
    - *Echinostoma* sp.
      - Rat (experimental)

- **Fasciolidae**† Railliet, 1895
  - **Fasciolinae**† Stiles and Hassall, 1898
    - *Fasciola gigantica* Cobbold, 1855
      - Cattle, Man, Swine
    - *Fasciola hepatica* Linnaeus, 1758
      - Cattle

- **Heterophyidae**† Odhner, 1914
  - **Ascoctylinae**† Yamaguti, 1961
    - *Phagicola longus* Ransom, 1920
      - Cat, Dog

  - **Centrocestinae**‡ Looss, 1899
    - *Centrocestus formosanus* Nishigori, 1924
      - Night heron, Rat
  
    - *Haplorchis taichui* (Nishigori, 1924)
      - Night heron
    - *Haplorchis yokogawai* (Katsuda, 1932)
      - Night heron

  - **Metagoniminae**‡ Ciurea, 1924
    - *Stellantchasmus falcatus* Onji and Nishio, 1915
      - Cat, Dog, Man, Rat

- **Philophthalamidae**† Travassos, 1918
  - **Philophthalmineae**‡ Looss, 1899
    - *Philophthalmus gralli* Mathis and Legar, 1910
      - Hawaiian coot

- **Plagiorchiidae**† Ward, 1917
  - **Plagiorchiinae**‡ Pratt, 1902
    - *Plagiorchis muris* Tanabe, 1922
      - Rat

- **Schistosomatidae**† Poche, 1907
  - **Schistosonatinae**† Stiles and Hassall, 1898
    - *Austrobilharzia variglandis* (Miller and Northup, 1926)
      - Ruddy turnstone
THE LIFE CYCLES OF PARASITES

A knowledge of the life cycles of parasites and their methods of infecting the final host is of paramount importance in determining methods of prevention and in formulating effective control measures. It helps to ascertain the weakest or most vulnerable point at which the parasite can best be attacked. Parasites may be controlled while in the final host by the use of chemotherapeutics, or when outside the final host by the use of chemicals and other means such as mechanical or biological destruction of the eggs, larvae, or their vectors. As a rule, each parasite has a specific course of development; however, the general pattern which the parasite follows is more or less similar to that of the group to which it belongs. Parasites have either a direct life cycle, in which they require only one host, or an indirect cycle, in which one or more intermediate hosts are required. An intermediate host is the one in which the young parasite undergoes development leading to the stage which is infective to the final host. When only one intermediate host is required, the infective stage is reached in the first host; when two are necessary, it is attained in the second. The final host is the one in which the parasite reaches the adult or sexual stage.

Animal parasites belong to five major groups; these are the unicellular organisms (Protozoa), and the multicellular forms which include the roundworms (Nematoda), the tapeworms (Cestoda), the flukes (Trematoda), and certain parasitic insects, mites, and ticks (Arthropoda). The roundworms, tapeworms, and flukes discussed here are all internal parasites, whereas the parasitic arthropods are primarily external parasites except for a relatively few insects which, during their larval stage, live in the body of the host. A summary of the life cycle pattern of each of the above groups of parasites which are represented in this bulletin is given below. For a detailed account of the life cycle of a variety of internal parasites, see Olsen (1962); for the arthropods, the reader should refer to various books on medical entomology.

Life Cycle of Protozoa

The known protozoan parasites of man and animals in Hawaii include the amoebae (Rhizopoda), the flagellates (Zoomastigophorca), the ciliates (Ciliatea), and the coccidia and haemosporidia (Telosporea). The haemosporidia are blood parasites which have an indirect life cycle. Included with the blood parasites are the flagellates, Trypanosoma lewisi, of rats, which utilize fleas as their vector, and the flagellates, Trypanosoma conorhini, believed to be parasites of rodents, which utilize reduviid bugs as their vector. All the other protozoa of man and animals in Hawaii live, for the most part, in the digestive tract, although some live in other tissues of the body and have a direct life cycle.
The amoebae, in the active or trophozoite stage, reproduce by fission in the body of the host. Some of the trophozoites pass out in the feces unchanged while others undergo encystment and are eliminated as infective cysts. Cysts are somewhat resistant to external environment, while most of the trophozoites degenerate rather quickly. Forms like *Dientamoeba fragilis* of man have no cysts, and the mode of infection is not clear; it has been suggested that it may be transmitted through the egg of *Enterobius vermicularis* (see Burrows and Swerdlow, 1956). Others, such as *Entamoeba coli* and *E. histolytica*, form multinucleated cysts. When these are ingested with contaminated food or water, they excyst, grow, divide, and establish an infection in the digestive tract.

The intestinal flagellates and ciliates, like the amoebae, multiply in the host by fission and are eliminated both as trophozoites and as cysts. Forms like *Giardia lamblia*, *Chilomastix mesnili*, and *Balantidium coli* have infective cysts. The trichomonads and histomonads do not form cysts and are infective in the trophozoite stage in which they are eliminated in the feces. By far the more common form of transmission of *Histomonas meleagridis*, however, is in the egg of the cecal worm, *Heterakis gallinarum* (see Parasites of Chicken and Other Avian Hosts). The blood flagellates, *Trypanosoma lewisi*, multiply by longitudinal division in the body of the host. Upon reaching the stomach of the flea vector, they assume the crithidial form, enter the epithelial cells, and multiply. Gradually they travel backwards towards the rectum where they change to the infective or metacyclic forms and are eliminated in the feces. Infection of the rat is believed to take place by eating infected fleas or their feces.

The coccidia have a characteristic nonsporulated oocyst stage in which they are eliminated in the feces of the host. In a few days each oocyst gives rise to spores (sporoblasts) which in turn divide into sporozoites, at which time it is known as a sporulated or an infective oocyst. When ingested by the definitive host, each sporozoite enters an intestinal cell and grows into a mature schizont which gives rise to a large number of merozoites. When the latter are liberated, they enter other intestinal cells and repeat the asexual cycle (schizogony). Some of these merozoites which enter new cells, however, become either macro- or microgametocytes and represent the first developmental stages of the sexual cycle. Each macrogametocyte, upon maturing, becomes a macrogamete (mature ovum). Each microgametocyte, on the other hand, gives rise to numerous comma-shaped, biflagellate microgametes (sperm cells). Following fertilization, the ovum (zygote) passes out in the feces of the host as a nonsporulated oocyst.

In the case of the bird-protozoa, *Haemoproteus columbae*, the infective sporozoites are transmitted through the bite of the pigeon-fly. The sporozoites start the schizogony cycle upon entering the reticulo-endothelial cells of
the liver and spleen of the avian host. They grow, multiply, and eventually form a large number of merozoites. When liberated from the host cells, some of the merozoites enter other endothelial cells and repeat the schizogyony cycle. The others enter the red blood cells and begin the sexual cycle. In the blood cells, the parasites become sausage-shaped and develop into micro- and macrogametocytes. When ingested by the pigeon-fly, these cells go to the stomach and undergo further growth. The macrogametocyte matures into a macrogamete and each microgametocyte differentiates into a number of filamentous microgametes. Upon fertilization, the zygote thus formed becomes a motile ookinete. This enters the stomach wall and forms a pigmented oocyst. The latter gives rise to a large number of sporozoites which eventually escape and reach the salivary glands of the fly. The sporozoites are then transferred to susceptible birds through the bite of the infected fly.

*Plasmodium vaughani* is the only species of bird malaria known in Hawaii. It is probably mosquito-borne and its cycle is only partially known (Manwell, 1947; Laird, 1953).

**Life Cycle of Roundworms**

Roundworms are usually referred to as having five developmental stages in their life cycle, namely: (a) the first, second, third, and fourth larval stages and (b) the fifth or adult stage. There are four molts, one between each stage. The adult female worms are either oviparous or ovoviviparous. Oviparous nematodes (*Ascaris, Trichuris*, etc.) usually lay undeveloped eggs which are eliminated in the feces of the host. Ovoviviparous nematodes (*Acarops, Enterobius*, etc.) usually lay eggs containing immature first-stage larvae which are also eliminated in the feces of the host. In some instances, the eggs of ovoviviparous worms develop to immature first-stage larvae which hatch out in the uterus of the female worms. Such immature larvae may be deposited under the skin (*Stephanosilicaria*) or in the circulatory system (*Dirofilaria*) to be picked up by biting insects. The immature first-stage larvae of filarioid nematodes are referred to as "microfilariae."

Roundworms have either a direct or an indirect life cycle depending on whether they have an intermediate host.

**Roundworms with a Direct Life Cycle**

Roundworms that have a direct life cycle known to be present in Hawaii include members of the families Trichuridae, Trichosomoididae, Ascarididae, Heterakidae, Oxyuridae, Ancylostomatidae, Cyathostomidae, Stephanuridae, Strongylidae, Trichostrongylidae, Strongyloidiidae, and a member of the Protostrongylidae (*Dictyocaulus viviparous*).
The Trichuridae (*Trichuris* and *Capillaria*) are oviparous. The eggs of *Trichuris*, which are eliminated in the feces, become fully embryonated (contain young first-stage larvae) within a few weeks and are infective to the final host. Among the capillarids, there is considerable variation in the manner by which the transmission of the eggs from one host to another is accomplished. In *C. columbae*, occurring elsewhere, the eggs are eliminated in the feces and after embryonation are infective to the final host as in *Trichuris*. In *C. hepatica*, the eggs, which are laid and remain in the liver tissue, seldom escape in the feces of the rat (Momma, 1930). However, when the liver is eaten by another animal such as a cat or a rat, the eggs, once freed in the feces of these animals, become embryonated and are infective to the final host (Shorb, 1931). In certain species of capillarids (*C. caudin-flata*), occurring elsewhere, an earthworm serves as an intermediate host (Wehr and Allen, 1945).

The Trichosomoididae (*Trichosomoides*) are ovoviviparous and the infective eggs are eliminated in the urine of the host.

The Ascarididae (*Ascaridia, Ascaris, Parascaris, Toxascaris*, and *Toxocara*) and the Heterakidae (*Heterakis*) are oviparous. The eggs, which are eliminated in the feces, become embryonated within 2 weeks and the larvae molt once before becoming infective (Alicata, 1934). Among the Oxyuridae (*Enterobius* and *Oxyurus*), the female worms are ovoviviparous and the eggs are believed to be infective soon after they are deposited. When the infective eggs of heterakids and oxyurids are ingested, they hatch in the body of the host and the young parasites migrate and develop solely in the digestive tract. Among the ascarids [except *Ascaridia* and *Toxascaris* (see Wright, 1935)], the hatched larvae migrate and develop partially in the lungs of the host before traveling and maturing in the intestinal tract.

Among members of the families Ancylostomatidae (*Ancylostoma* and *Bunostomum*), Cyathostomidae (*Cyathostomum*, etc.), Stephanuridae (*Stephanurus*), Strongylidae (*Alfortia*, etc.), and Trichostrongylidae (*Haemonchus, Nematodirus*, etc.), the female worms are oviparous and the eggs are eliminated from the body of the host in the feces, except in Stephanuridae, in which the eggs are eliminated in the urine. Usually after a few hours the eggs become fully embryonated and hatch. Within a few days the first three larval stages develop in the soil and these represent the free-living or pre-parasitic stages. In species of *Nematodirus*, the larvae develop slowly within the egg to the third stage and eventually hatch under suitable climatic conditions (Kates and Turner, 1955). Among parasites of the above five families, only the third-stage larvae are infective to the final host. These larvae are usually enclosed within the sheath of the second molt. In the first two stages the larvae of these parasites are referred to as rhabditiform larvae because their esophagus is composed of a corpus, isthmus, and bulb. The
third-stage larvae, on the contrary, do not have such a bulb and are referred to as filariform larvae. The infective larvae usually enter the final host through contaminated food or water, but some are able to enter by boring through the intact skin (Ancylostoma, Nippostrongylus, Stephanurus). Such larvae migrate and develop partially in the lungs or other tissues before settling in the specific site of the body where they reach sexual maturity.

Members of the Strongyloididae (Strongyloides) are oviparous and develop somewhat like the above Ancylostomatidae and related strongyles. The parasitic cycle, however, may alternate with a free-living cycle. In the latter case, the larvae at all stages and the adults are rhabditiform and develop in the soil as free-living organisms. Their offspring, however, may follow either the free-living or the parasitic cycle. If the parasitic cycle is followed, the third-stage larvae become filariform and are able to penetrate the intact skin of the host; they are not ensheathed.

Most members of the family Protostrongylidae have an indirect life cycle, except Dictyocaulus viviparus which is direct. In D. viviparus, the female worms are ovoviviparous. The eggs, which are deposited in the lungs, frequently hatch in the digestive tract of the host and are eliminated as first-stage larvae which develop in the soil to the third or infective stage. When ingested by cattle, the larvae migrate to the mesenteric lymph nodes and later to the lungs, where they reach sexual maturity (Porter and Cauthen, 1942).

Roundworms with an Indirect Life Cycle

Roundworms which have an indirect cycle usually undergo their first three larval stages in an intermediate host. In the latter, the third-stage larva may or may not encyst, but this is the stage which is infectious to the final host. The intermediate host is usually an annelid, a mollusk, or an arthropod.

In some cases, the infected intermediate host is ingested whole or in part by other invertebrate or vertebrate scavengers and the infective larvae are transferred and may re-encyst in the body of these scavengers. Such carriers are known as paratenic hosts. An example is the land planarian, which acquires the third-stage larvae of the rat lungworm, Angiostrongylus cantonensis, by feeding on the body of infected snails, the latter being the true intermediate hosts.

In Hawaii, roundworms which have an indirect life cycle include members of the families Acuaridae, Physalopteridae, Spiruridae, Subuluridae, Thelaziidae, Tropisuridae, Protostrongylidae (except D. viviparus), Pseudaliiidae, Dipetalonematidae, Stephanofilariidae, and Trichinellidae.

In the Acuaridae (Acuaria, Dispharynx), Physalopteridae (Physaloptera), Spiruridae (Ascarops, Cyrnea, Habronema, Gongylonema, Physocephalus),
Subuluridae (Subulura), Thelaziidae (Oxyspirura), and Tropisuridae (Tropisurus), the adult females are ovoviviparous and the eggs are eliminated in the feces of the host. Whenever these eggs are ingested by an arthropod, they hatch and the larvae develop to the third or infective stage. These larvae then develop to maturity whenever the infected intermediate host is ingested by the final host.

Among the Protostrongylidae, some are oviparous (Angiostrongylus, Aelurostrongylus), and some ovoviviparous (Choerostrongylus, Metastrongylus). In Angiostrongylus and Aelurostrongylus, the eggs develop and hatch in the pulmonary capillaries. The first-stage larvae travel from the lungs to the trachea and out of the body in the feces. Certain land snails and slugs serve as intermediate hosts. In Choerostrongylus and Metastrongylus, the embryonated eggs are eliminated in the feces. Earthworms serve as intermediate hosts. When the eggs are ingested by earthworms, they hatch and the larvae develop to the third or infective stage. In the Pseudaliidae (Anafilaroides), the females lay young, first-stage larvae which travel from the lungs to the trachea and out of the body in the feces. Certain land snails and slugs serve as intermediate hosts. Whenever mollusks harboring infective larvae of Anafilaroides and Aelurostrongylus are ingested by rodents, the larvae re-encyst in the body of the rodent and retain the power to infect the final host. Rodents, therefore, can serve as paratenic hosts for these parasites.

In the Dipetalonematidae (Dirofilaria, Dipetalonema), the adult females produce microfilariae which enter the circulatory system. Various biting arthropods such as mosquitoes (for Dirofilaria) and fleas (for Dipetalonema) serve as the intermediate hosts. The females of the Stephanofilariididae (Stephanofilaria) also produce microfilariae which are deposited in the skin. Biting horn flies of cattle are believed to serve as intermediate hosts for S. stilesi.

In the Trichinellidae (Trichinella), the females are ovoviviparous and the immature first-stage larvae are formed in utero. Soon after birth the young larvae penetrate the intestinal wall, reach the circulatory system, and are distributed throughout the body. After undergoing a period of development, the larvae become encysted in the voluntary muscles, at which time they are infective to the final host. Trichinae differ from other nematodes having an indirect life cycle in that they utilize their final host as intermediate host also.

**Life Cycle of Tapeworms**

All the adult tapeworms known to occur in Hawaii are found in the digestive tract of the final host. The eggs of the parasites, and sometimes the terminal gravid segments containing the eggs, are eliminated in the feces
of the host. Each egg possesses a well-developed hexacanth embryo which is infective to the intermediate host. In the latter, the embryo develops to the stage which is infective to the final host. All tapeworms reported in this bulletin require either an arthropod or a mammalian intermediate host, except *Hymenolepis nana*, which usually develops without an intermediate host.

The following families of tapeworms are recorded from Hawaii, and the type of intermediate host used by them is shown in parentheses: Anoplocephalidae (free-living oribatid mites), Davaineidae (insects), Dilepididae (insects), Hymenolepididae (insects), and Taeniidae (mammals). Among the above-mentioned families (except Taeniidae), the hexacanth embryo develops in the intermediate host to the cysticercoid stage, which is infective to the final host. In the case of *H. nana*, when the egg is ingested by the final host, the hexacanth embryo enters an intestinal villus and forms a tailless cysticercoid. This then escapes to the lumen of the intestine and develops to maturity. In members of the Taeniidae (*Taenia saginata*, *T. hydatigena*, etc.), the egg, when ingested by a suitable mammalian intermediate host, develops into a comparatively large infective larva known as cysticercus or bladderworm stage.

**Life Cycle of Flukes**

There are comparatively few species of flukes reported as established in terrestrial vertebrates in Hawaii. These include members of the families Brachylaemidae (*Postharmostomum gallinum*), Dicrocoeliidae (*Platynosomum fastosum*), Echinostomatidae (*Echinostoma* sp.), Fasciolidae (*Fasciola hepatica* and *F. gigantica*), Heterophyidae (*Stellantchasmus falcatus*, etc.), Philophthalmidae (*Philophthalmus gralli*), Plagiorchiidae (*Plagiorchis muris*), and Schistosomatidae (*Austrobilharzia variiglandis*).

All the above parasites are digenetic and require a snail as an intermediate host. In the Heterophyidae, two intermediate hosts are required, a fresh-water snail and a fish. The echinostomid and plagiorchid flukes possibly utilize fresh-water snails as both first and second intermediate hosts.

All the above-named flukes, except *Fasciola*, are ovoviviparous since the eggs contain a well-formed miracidium. In the fasciolids, the eggs develop after they have been eliminated in the feces of the final host. In the case of *Fasciola, Philophthalmus*, and *Austrobilharzia*, the eggs hatch in water and the miracidia actively bore into the tissue of the snail. In the other flukes reported herein, the eggs hatch after they are ingested by the snail host.

In the suitable snail host, each miracidium undergoes growth and asexual propagation which is terminated with the formation of numerous cercariae. In *Postharmostomum*, the mature cercariae reach the infective stage (meta-
cercaria) by entering the pericardial cavity of the same or of other land
snails (Alicata, 1940). In other species of flukes, the cercariae emerge from
the snail and swim in water. In the case of *Austrobilharzia*, the motile cer-
cariae are able to enter the final host by active penetration of the skin. In
*Fasciola* and *Philophthalmus*, the cercariae encyst as metacercariae on the
surface of vegetation or other objects. In the heterophyids, the cercariae
encyst as metacercariae in the musculature of the secondary intermediate
host, which is a fish. In *Echinostoma* and *Plagiorchis*, the cercariae may
encyst as metacercariae in the body of the same or another host. The en-
cysted metacercariae are infective to the final host.

**Life Cycle of Arthropods**

Arthropods which are parasitic on man and animals in Hawaii include,
for the most part, a variety of surface-feeding and blood-sucking insects (bed
bugs, fleas, biting flies, biting and sucking lice, mosquitoes) and acarines
(mites and ticks). Botflies, sheep nasal flies, and warble flies are, however,
internal parasites in their larval stages.

Of the above insects, the fleas, flies, and mosquitoes have four phases in
their life cycle, namely, egg, larva, pupa, and adult (complete metamorpho-
sis). In the life cycle of the bed bugs and lice, on the other hand, there are
three phases represented, namely, egg, nymph, and adult (incomplete meta-
morphosis). Among the species of insects considered in this bulletin, the lice
are parasitic during their whole life cycle; others, such as fleas, horseflies,
and mosquitoes, are temporarily parasitic during the adult stage only;
others, such as warble flies, botflies, and sheep nasal flies, are parasitic only
during the larval stage.

The life cycle of mites and ticks is divided into four phases, namely, egg,
larva, nymph, and adult. Whereas the nymphs and adults have four pairs of
legs, the larvae have three pairs. Parasitic mites of the genera *Demodex*,
*Psoroptes*, and *Sarcoptes* reported herein spend their whole life cycle on
the body of the host. Others, such as those of the genera *Laelaps*, *Ornithony-
sus*, and *Pyemotes*, are free-living a great deal of the time and attack the host
temporarily during the nymphal and adult stages. They drop from the host
after each feeding.

The life cycles of two species of soft ticks (*Otobius megnini*, *Ornithodorus
capensis*) and a species of hard tick (*Rhipicephalus sanguineus*) known in
Hawaii differ in a few respects. *O. megnini* and *O. capensis* are parasitic
only during the larval and nymphal stages, whereas *R. sanguineus* is para-
sitic during the larval, nymphal, and adult stages. Following engorgement,
the soft ticks in the nymphal stage drop to the ground and after a few days
molt and become adults. Following mating of the sexes, the gravid females
lay eggs. After hatching, the larvae (also known as seed ticks) attach themselves to hosts with which they come in contact and repeat the cycle. As for the hard tick, *R. sanguineus*, the important feature of its life cycle is that each of the three phases of the cycle (larva, nymph, adult) requires a separate animal. Both the larvae and nymphs, following engorgement, drop and molt into the next stage. The adult females, following engorgement, drop and lay eggs. After hatching, the larvae attach to another animal with which they come in contact and repeat the life cycle.

**ROUTES OF INFECTION OF INTERNAL PARASITES**

The final host may become infected with parasites by various methods, including oral, cutaneous, and prenatal routes, or combinations of these routes.

In oral infections, the infective cysts, eggs, or larvae are ingested with contaminated or infected food or water.

In cutaneous infections, the infective larvae of certain nematodes (*Strongyloides stercoralis, Ancylostoma caninum*, etc.) or trematodes (*Schistosoma mansoni*, etc.) are able to penetrate the intact skin of the host. Included also are certain protozoans (*Haemoproteus columbae*, etc.) or nematodes (*Dirofilaria immitis*, etc.) which are transmitted by the bite of arthropod vectors.

In prenatal infections, the unborn final host may become infected while *in utero*. Although this is rare among internal parasites, it is true among certain nematodes which in the larval stage migrate to various parts of the body and find their way from the maternal circulatory system to the fetal tissue. Prenatal infection has been reported for the nematodes, *Ancylostoma caninum* (see Yutuc, 1949), *Strongyloides ransomi* (see Stewart et al., 1963), *Strongyloides westeri* (see Taylor, 1955), and *Toxocara canis* (see Sprent, 1958; Webster, 1958; Yutuc, 1949), and the blood fluke, *Schistosoma japonicum* (see Fujinami and Nakamura, 1911).
MOLLUSKS OF PARASITOLOGICAL IMPORTANCE IN HAWAII

<table>
<thead>
<tr>
<th>MOLLUSK (NATURAL SIZE)</th>
<th>LOCATION FOUND</th>
<th>IMPORTANCE</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Achatina fulica</strong></td>
<td>On land</td>
<td>Intermediate host for the rat lungworm, <em>Angiostrongylus cantonensis</em>, which produces cerebral angiostrongylosis (parasitic eosinophilic meningoencephalitis) in man (see pp. 31-36).</td>
</tr>
<tr>
<td><strong>Bradybaena similaris</strong></td>
<td>On land</td>
<td>Intermediate host for: (a) the cat lungworms, <em>Anasiiaroides rostratus</em> and <em>Aelurostrongylus abstrusus</em>; (b) the chicken cecal fluke, <em>Postharmostomum gallinum</em>; (c) the rat lungworm, <em>Angiostrongylus cantonensis</em>, which produces cerebral angiostrongylosis (parasitic eosinophilic meningoencephalitis) in man (see pp. 31-36).</td>
</tr>
<tr>
<td><strong>Fossaria ollula</strong></td>
<td>On banks of freshwater streams and swamps</td>
<td>Intermediate host for the cattle liver flukes, <em>Fasciola gigantica</em> and <em>F. hepatica</em>.</td>
</tr>
</tbody>
</table>
# Mollusks of Parasitological Importance in Hawaii

<table>
<thead>
<tr>
<th>Mollusk</th>
<th>Location Found</th>
<th>Importance</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Littorina pintado</em></td>
<td>On marine rocks</td>
<td>Intermediate host for the blood fluke, <em>Austrobilharzia variglandis</em>, of certain fish-eating birds. The larvae (cercariae) emerging from the snails are potentially able to produce dermatitis in man.</td>
</tr>
<tr>
<td><em>Opeas javanicum</em></td>
<td>On land</td>
<td>Intermediate host for the rat lungworm, <em>Angiostrongylus cantonensis</em> (see <em>Achatina fulica</em>).</td>
</tr>
<tr>
<td><em>Pseudosuccinea columella</em></td>
<td>On banks of fresh-water streams and swamps</td>
<td>*(Same as <em>Fossaria ollula)</em></td>
</tr>
<tr>
<td><em>Stenomelania newcombi</em></td>
<td>In fresh-water streams</td>
<td>Intermediate host for the following intestinal flukes: (a) <em>Centrocestus formosanus</em> in the night heron and rat; (b) <em>Haplorchis yokogawai</em> in the night heron; (c) <em>Stellantchasmus falcatus</em> in the cat, dog, man, and rat. Also intermediate host for the eye-fluke, <em>Philophthalmus gralli</em>, in the Hawaiian coot, and also capable of developing in mammals.</td>
</tr>
<tr>
<td><em>Subulina octona</em></td>
<td>On land</td>
<td>*(Same as <em>Bradybaena similaris)</em></td>
</tr>
</tbody>
</table>
MOLLUSKS OF PARASITOLOGICAL IMPORTANCE IN HAWAII

<table>
<thead>
<tr>
<th>MOLLUSK (NATURAL SIZE)</th>
<th>LOCATION FOUND</th>
<th>IMPORTANCE</th>
</tr>
</thead>
<tbody>
<tr>
<td>Thiara granifera</td>
<td>In fresh-water streams</td>
<td>Intermediate host for the following intestinal flukes: (a) Centrocestus formosanus in the night heron and rat; (b) Haplorchis taichui and H. yokogawai in the night heron; (c) Stellantchasmus falcatus in the cat, dog, man, and rat. Also intermediate host for the eye-fluke, Philophthalmus gralli, in the Hawaiian coot, and also capable of developing in mammals. Potential host for the lung fluke, Paragonimus westermani, in man.</td>
</tr>
</tbody>
</table>

SLUGS

<table>
<thead>
<tr>
<th>Slug</th>
<th>Location Found</th>
<th>Importance</th>
</tr>
</thead>
<tbody>
<tr>
<td>Deroceras laeve</td>
<td>On land and vegetation</td>
<td>Intermediate host for the rat lungworm, Angiostrongylus cantonensis (see Achatina fulica).</td>
</tr>
<tr>
<td>Veronicaella alte</td>
<td>On land and young ones occasionally on vegetation</td>
<td>Intermediate host for the rat lungworm, Angiostrongylus cantonensis (see Achatina fulica).</td>
</tr>
</tbody>
</table>
According to a survey by Ching (1961a), seven species of intestinal protozoa in man have been recorded from Hawaii. These species, and the percentages of infection found among 1,380 persons examined, are as follows: Dientamoeba fragilis, 0.36; Endolimax nana, 0.84; Entamoeba coli, 1.96; E. hartmanni, 0.14; E. histolytica, 0.29; Giardia lamblia, 1.01; Trichomonas hominis, 0.07. Amoebic dysentery appears to be rare in Hawaii.

Human malaria has been diagnosed from civilian and military personnel who acquired the infection elsewhere. Natural transmission is not possible in Hawaii because of the absence of anopheline mosquitoes.

Various species of roundworms occur in Hawaii, and the incidence is more likely to be higher in the rural areas, especially among plantation laborers who have migrated from the Orient. In a stool examination of 1,009 persons living on a plantation on the island of Maui, Powers (1937) found the following percentages of roundworm infection: Ascaris lumbricoides, 0.99; Enterobius vermicularis (fig. 1a), 3.5; hookworm (species unknown), 13.6; Trichuris trichiura, 17.1. Of the persons examined, 387 were Filipino, 269 Japanese, 216 Portuguese, 100 Puerto Rican, and 37 unclassified.

Hall (1936) reports specimens of Trichostrongylus colubriformis from man which were given to him by a medical technician in Hilo, on the island.

FIGURE 1. Pinworms, Enterobius vermicularis: a, adults, natural size; b, eggs, highly magnified. (After Alicata and Kartman, 1948, courtesy of Hawaii Medical Journal.)
of Hawaii. These are parasites which frequently occur in goats and sheep. Trichostrongyle eggs were also recorded from stool examinations on Maui.

In a survey involving the stool examination of 1,195 persons from the island of Oahu and 185 persons from the island of Hawaii, the following percentages of roundworm infection were found (Ching, 1961a): *Ascaris lumbricoides*, 0.94; *Enterobius vermicularis*, 0.58; *Strongyloides stercoralis*, 1.16; *Trichostrongylus* sp., 0.07; *Trichuris trichiura*, 2.03; hookworms, 0.58.

Infection produced by the pinworm, *Enterobius vermicularis*, appears to be common especially among children. In an examination (anal swab technique) of a group of 138 children from Honolulu in the age range of 2 to 7 years, 31.1 percent showed pinworm eggs (fig. 1b) (Alicata and Kartman, 1948).

Infections of the nonperiodic form of the filarioid, *Wuchereria bancrofti*, occur among Samoan immigrants to Hawaii. There is no record, however, that the infection has been transmitted to local inhabitants. Jackowski (1950) examined 72 Samoans who had lived in Hawaii less than 6 years and 8 showed microfilariae in the blood. None of 62 other Samoans who were born and reared in Hawaii showed infection. He postulated that, following last infection, microfilaremia persists for at least 5 years and disappears within 10 years. In Samoa, this parasite is transmitted by *Aedes scutellaris pseudoscutellaris* (see Byrd et al., 1945), a mosquito which is not in Hawaii. Of the species of mosquitoes in Hawaii, the nonperiodic strain of *W. bancrofti* has been shown to develop in *Culex pipiens quinquefasciatus* (see Eyles and Most, 1947). This species appears, however, to be less receptive to the nonperiodic forms of *W. bancrofti* than *A. s. pseudoscutellaris*. Although *C. p. quinquefasciatus* is a potential vector of filariasis in Polynesia, its importance is not yet completely known (Rosen, 1953). Nelson et al. (1946) found that *C. p. quinquefasciatus* in Hawaii is susceptible to experimental infection with the periodic form of microfilariae of *W. bancrofti* from Okinawa. However, the microfilariae of *Brugia malayi* obtained from Koreans failed to develop to the third stage in this species of mosquito.

The first laboratory-proved case of human trichinosis, caused by *Trichinella spiralis* (fig. 32a), was found in Hawaii in 1936 (Alicata, 1938e). This led to a survey in the various islands to determine the infection among animals in nature (see Parasites of Swine). From 1936 to 1964, there were 112 cases of human trichinosis reported in the annual reports of the Hawaii Department of Health. Most of the infected persons had eaten, or were suspected of having eaten, improperly cooked wild pork or products made from wild pork. According to records of the Hawaii Department of Agriculture, during the 8-year period from 1933 through 1940 inclusive (Tinker, 1941), 32,724 wild hogs, or an average of 4,090 a year, were killed on the five larger
islands. Because of the moderately high incidence of trichinosis in local wild hogs (see Parasites of Swine), meat from these animals is believed to constitute a health menace unless proper precautions are taken in cooking, preserving, or refrigerating. In connection with the prevalence of local human infection with trichinae, in an examination of 133 diaphragms from autopsy cases in Honolulu, 7.4 percent showed encysted trichina larvae (Alicata, 1942b). The number of persons of the various ethnic groups examined and the number found positive (shown in parentheses) were as follows: Caucasian, 28 (3); Caucasian-Hawaiian, 7 (1); Caucasian-Japanese, 1 (0); Chinese, 9 (1); Chinese-Hawaiian, 2 (0); Chinese-Korean, 1 (0); Fijian, 1 (0); Filipino, 17 (1); Hawaiian, 24 (1); Japanese, 23 (0); Korean, 7 (2); Puerto Rican, 2 (0); unknown, 1 (0).

A fatal case of Capillaria hepatica infection was described in a 15-month-old child in Hawaii (Ewing and Tilden, 1956). The child had been known to eat dirt. The diagnosis was established before death by means of a liver biopsy which revealed many parasites and their ova (fig. 2a) within the liver. C. hepatica is normally a common parasite of the rat. Outside of Hawaii, it has also been reported from other mammals. Infection with this parasite results from the ingestion of embryonated eggs with contaminated food or water. Normally, in the rat, the adult worms deposit undeveloped eggs in the liver tissue (fig. 31e), and these eggs seldom escape in the feces of the rat (Momma, 1930). However, when the infected liver is eaten through cannibalism by another rat or predation by a cat, the eggs escape in the feces of these animals and eventually become embryonated and infective (Shorb, 1931). Although only four cases of C. hepatica have been recorded from man (Ewing and Tilden, 1956), there is a likelihood that the parasite may be more common in man than is suspected. The diagnosis is difficult because the eggs are seldom found in the feces.

A case of toxocarosis (visceral larva migrans) caused by the larvae of the dog and cat roundworm, Toxocara canis (fig. 28d), was reported from a 2-year-old boy in Hawaii with a history of eating dirt (Bruton and Jaffurs, 1957). The second-stage larvae of the parasite were recovered from a liver biopsy. According to the above writers, criteria which may aid in the diagnosis of this disease include the following: eosinophilia greater than 20 percent; history of eating dirt contaminated with embryonated eggs of T. canis; symptoms of the respiratory tract and nervous system; enlarged liver and spleen. Two other cases of larva migrans in Hawaii were reported by Ash (1962a). Outside of Hawaii, the syndrome of larva migrans appears to have been recognized only in children 15 to 36 months of age, and the majority at approximately 2 years of age (Beaver, 1962). Eosinophilic infiltration of the meninges resulting from invasion of larvae of T. canis has been reported.
Figure 2. a. Section of human liver showing eggs and cross section of *Capillaria hepatica*. (After Ewing and Tilden, 1956, courtesy of *Journal of Pediatrics*); b, cross section of the cysticercus stage of *Taenia solium* from human liver, highly magnified. (Section, courtesy of Queen’s Hospital, Honolulu.)
in a child in the continental United States (Dent et al., 1956); consequently, therefore, cerebral toxocarosis may give rise to eosinophilic meningoencephalitis.

In 1961, the rat lungworm, *Angiostrongylus cantonensis*, was recovered in Hawaii at an autopsy on the brain of a Filipino with a history of eosinophilic meningoencephalitis (Rosen et al., 1961, 1962). This finding followed the discovery by Ash (1962b; see Parasites of Rat) of the adult stage of *A. cantonensis* (fig. 31d) in the lungs of local rats, and confirmed the speculation originally made by Alicata (1961, 1962a) that this parasite may be the causative agent of eosinophilic meningoencephalitis in the Pacific (see also Alicata and McCarthy, 1964). A case of this disease, also referred to as parasitic meningoencephalitis and cerebral angiostrongylosis, occurred in a Japanese laborer in Hawaii following ingestion of two garden slugs, *Veronicella alte* (see Horio and Alicata, 1961).

*A. cantonensis* is normally a parasite of rats and utilizes mollusks (pp. 24–26) as intermediate hosts (see Parasites of Rat). Land planarians (*Geoplanina septemlineata*) in Hawaii, and fresh-water prawns (*Macrobrachium* sp.) and land crabs in other Pacific areas, have been found to serve as paratenic or transport hosts for the infective larvae (Alicata and McCarthy, 1964); experimentally, pigs and calves have also been found to serve in that capacity (Alicata, 1964b).

Human infection with *A. cantonensis* most likely occurs as a result of eating uncooked food (fig. 3) containing infective larvae of the parasite. Eating habits and customs of people may play an important part. In Tahiti, the common occurrence of eosinophilic meningoencephalitis has been traced to the customary habit of eating raw prawns, including, possibly, "taioro." The latter consists of grated coconut to which is added prawn juice, prepared by grinding the stomach and surrounding portions of the prawns in fresh water (Alicata and Brown, 1962). In Thailand, the disease is believed to be acquired as a result of eating the large amphibious snail, *Pila ampullacea* (see Punyagupta, 1964). The fleshy head-foot part of the snails is cut and then either dipped in boiling water or stored in an icebox to keep it fresh. It is then eaten after being chopped into small pieces, seasoned with lime juice, and mixed with vegetables. In New Caledonia and Hawaii, where eosinophilic meningoencephalitis occurs sporadically, it is probably acquired through the accidental ingestion of an infected small garden slug, or a carrier host such as a land planarian, with contaminated salad greens (Alicata, 1963a; Mead, 1963). Furthermore, in some areas, human infection may possibly take place from eating raw land crabs (Alicata, 1964a) and improperly cooked liver or other internal organs of swine or calves (Alicata, 1964b). These animals in their foraging habits are believed to ingest live mollusks.
Figure 3. Life cycle of the rat lungworm, *Angiostrongylus cantonensis*, and possible avenues of human infection. (Original.)
Experimentally, living larvae of *A. cantonensis* have been found in the stomach wall, liver, lungs, and spleen of pigs and calves 2 weeks after infection. In the pig, however, the larvae were found encapsulated and dead in the above organs 5 weeks after infection. The comparatively early encapsulation of the larvae, therefore, appears to minimize the importance of the pig as a carrier host. Experimentally, these larvae have not been found to migrate to the voluntary muscles of pigs or calves (Alicata, 1963c, 1964b). To what extent pigs and calves are infected with larvae of *A. cantonensis* under natural conditions and thus serve as sources of human infection remains to be determined.

Eosinophilic meningoencephalitis is a syndrome characterized by the presence of eosinophils in the cerebrospinal fluid. In man this syndrome has at times been noted in connection with cases of nonhelminthic and helminthic infections involving the central nervous system. Nonhelminthic infections have been observed in some cases of cerebral tumors, epidemic cerebrospinal meningitis, neurosyphilis, purulent meningitis, and tuberculous meningitis (Kaczynski, 1936). Helminthic infections include cerebral angiostrongylosis (Horio and Alicata, 1961; Rosen *et al*., 1962; Alicata, 1963a), cerebral cysticercosis (Kulkov, 1930), cerebral echinococcosis (Applebaum and Wexberg, 1944), cerebral paragonimiasis (Uematsu and Shiozaki, 1935; Nonomura, 1941), and cerebral schistosomiasis (Castaigne *et al*., 1959).

In the Pacific Basin, cases of eosinophilic meningoencephalitis have been reported from Micronesia, Polynesia, and Melanesia. A few additional cases have been reported from Japan (Nonomura, 1941) and the Philippines (Sison *et al*., 1951). In Southeast Asia, cases have occurred in Thailand (Punyagupta, 1964) and Sumatra (Smit, 1962). Laboratory and field evidence suggests that *A. cantonensis* is in most cases the causative agent of eosinophilic meningoencephalitis in Hawaii and other Pacific islands. This evidence includes: (a) recovery of young adult *A. cantonensis* from Inan in two cases of eosinophilic meningoencephalitis (Nomura and Lin, 1945; Rosen *et al*., 1962); (b) capability of the larvae of *A. cantonensis* to travel to the central nervous system of simian primates and to give rise to eosinophilic meningoencephalitis (Alicata, 1962a; Alicata, Loison, and Cavallo, 1963; Weinstein *et al*., 1963); (c) record of two human cases of eosinophilic meningoencephalitis following the willful ingestion of raw slugs from endemic areas (Horio and Alicata, 1961; Alicata, 1963a); (d) record of a human case of the disease in Honolulu following the ingestion of six raw giant African snails, *Achatina fulica* (see Mookini, 1964); (e) presence of lungworms among rats in all the Pacific islands (fig. 4; see also Parasites of Rat) in which eosinophilic meningoencephalitis has been recorded, namely, Cook Islands (Alicata and McCarthy, 1964), Formosa (Nomura and Lin, 1945), Guam (Loison, 1963), Hawaii (Horio and Alicata, 1961; Rosen *et al*.,
FIGURE 4. Geographical distribution of the rat lungworm, *Angiostrongylus cantonensis*, in the Pacific islands and Southeast Asia (indicated by stars), and its relationship to the distribution of eosinophilic meningoencephalitis in man (underscored). (See text, pp. 31-36.)
PARASITIC INFECTIONS OF MAN AND ANIMALS IN HAWAII

1962), New Caledonia (Trubert, 1952), New Hebrides (Loison, 1963), Pohnape (Bailey, 1948), Saipan (Allison, 1962), and Tahiti (Franco et al., 1960); (f) absence of the disease in areas of the Pacific where the rat lungworm is not known to occur, namely, Fiji, Samoa, Tonga, and Wallis (Loison, 1963); (g) high incidence of eosinophilic meningoencephalitis in Tahiti correlated with the frequent consumption of raw prawns, 4 percent of which have been found infected with the larvae of *A. cantonensis* (see Alicata and Brown, 1962); and (h) widespread incidence of the disease in parts of Thailand correlated with consumption of insufficiently cooked amphibious snails, *Pila ampullacea* (see Punyagupta, 1964).

The cause of eosinophilic meningoencephalitis reported from Japan and possibly from the Philippines, where *A. cantonensis* is not known to occur, in all probability is due to cerebral paragonimiasis. In Japan, Uematsu and Shiozaki (1935) reported a pleocytosis of 1,441 cells per cubic millimeter, consisting of practically all eosinophils, in the cerebrospinal fluid of an individual who showed meningeal irritations, cloudiness of both lungs in the X-ray examination, and numerous *Paragonimus* eggs in the sputum. In the same way, Nonomura (1941) reported a pleocytosis with 98 percent eosinophils in the cerebrospinal fluid of another patient in Japan. Although no fluke eggs were found in the sputum of this patient, Nonomura concluded that the pleocytosis was most likely produced by cerebral paragonimiasis. Furthermore, the sporadic cases of eosinophilic meningoencephalitis, which have been reported from Europe and North and South America, where *A. cantonensis* is not known to occur, are possibly caused by one or more species of helminths which occasionally invade the central nervous system (Smit, 1962). Of importance in this connection is the finding of eosinophilic infiltration of the meninges, resulting from larval infection of *Toxocara canis*, which has been observed in a child in the continental United States (Dent et al., 1956). Etiologically, however, infection with larvae of *Toxocara* occurs most commonly in young children, whereas eosinophilic meningoencephalitis in the Pacific area occurs chiefly among adults.

Of interest is the apparent absence of *A. cantonensis* among rats in Fiji, the Philippines, Samoa, Tonga, and Wallis Islands, whose climatic conditions and fauna are generally similar to those of other Pacific islands in which the parasite occurs. In all probability, this condition points out that the parasite is a recent immigrant to the Pacific islands and one which as yet has not become more widely distributed. Its original source of dispersal appears to be Eastern Asia. It was first recorded from Canton, China, by Chen in 1935, and in 1937 it was reported by Matsumoto and Yokogawa from Formosa. It appears to have gradually spread to various Pacific islands either through importation of infected mollusks or infected rats. This has probably been brought about by recent increased commercial and military shipping.
operations, especially during World War II, from Eastern Asia to various Pacific ports. Further evidence of the recent dispersal of the rat lungworm in the Pacific region appears to be the recent occurrence of eosinophilic meningoencephalitis in the Pacific islands. This syndrome was first noted in Formosa in 1944 (Nomura and Lin, 1945), Ponape in 1947 (Bailey, 1948), New Caledonia in 1951 (Trubert, 1952), and Tahiti in 1958 (Franco et al., 1960).

As indicated above, \textit{A. cantonensis} was first discovered in East Asia in 1935. Furthermore, the first case of eosinophilic meningoencephalitis in the Pacific was reported from Formosa in 1944. In this connection, it is of importance to note that these findings followed shortly after the introduction of the giant African snail, \textit{Achatina fulica}, in the areas. \textit{A. fulica} is an ideal intermediate host of \textit{A. cantonensis}. According to Mead (1961), during the nineteenth century, the achatinid snails became dispersed from their East African home to Southeast Asia and from there to East Asia and the Pacific islands. They were first found in Malaya in 1911, Indonesia in about 1930, China in 1931, Formosa in 1932, the Mariana and Hawaiian Islands in 1936. These data point out that \textit{A. fulica} might have imported or assisted in the spread of the rat lungworm in Asia and in the Pacific islands. If this is true, it is possible that the original habitat of the parasite is East Africa, the same as that of \textit{A. fulica}. Although \textit{A. fulica} is not known to occur in Australia, New Caledonia, or Tahiti, where \textit{A. cantonensis} is now found, it is possible that the parasite was imported in these areas through infected land mollusks or infected rats from Southeast Asia or Indonesia after it had become established there. The probability that \textit{A. cantonensis} might have originated from East Africa or nearby areas is being further investigated by the author.*

The geographical area in which \textit{A. cantonensis} is presently known to occur in man and rodents is limited to the tropical belt which extends approximately from the Tropic of Cancer (23° North latitude) to the Tropic of Capricorn (23° South latitude) (fig. 4), and from Thailand (100° East longitude) to the island of Tahiti (150° West longitude). This area is characterized by tropical and subtropical climate, moderate to heavy rainfall, and considerable vegetation. All these factors are highly conducive for the propagation and spread of mollusks and rodents.

\textbf{TAPEWORMS}

Most cases of tapeworm infection that have occurred in Hawaii probably represent infections acquired elsewhere. In a survey carried out by Powers

* After this manuscript was submitted for publication, Dr. Kenichi Nishimura and Dr. Mariano G. Yogore reported to the writer of finding \textit{Angiostrongylus cantonensis} among rats in Manila. The writer has also found \textit{A. cantonensis} in the lungs of rats on the islands of Mauritius, Madagascar, and Ceylon.
(1937) among 1,009 plantation workers on the island of Maui, 0.3 percent harbored *Taenia saginata* and 0.8 percent *Hymenolepis nana*.

Cattle are the intermediate host to *T. saginata*. Man acquires the infection by eating raw or undercooked beef containing the infective bladderworm stage (cysticercus) encysted in the musculature. Infection with *H. nana* usually results either from ingesting the infective eggs of the parasite or from accidentally ingesting one of the infected arthropod intermediate hosts (see Parasites of Rat).

In Hawaii, *T. saginata* has been reported mostly from immigrant Filipino workers. Price (1946) reported that, of 126 cases of *T. saginata* diagnosed at Queen’s Hospital in Honolulu during 1942 and 1945, the following ethnic distribution was represented: Caucasian, 7; Chinese, 2; Filipino, 110; Hawaiian and Part-Hawaiian, 4; Japanese, 2; Syrian, 1.

In a survey of parasites of man in Hawaii, Ching (1961a), found *Taenia* sp. and *H. nana* in 6 and 1 cases, respectively, out of 1,380 persons examined for parasite eggs.

A case of hydatid infection (larval stage of *Echinococcus granulosus*) was reported by Giles (1947), but the infection was believed to have been acquired outside of Hawaii. The 1954 annual report of the Hawaii Department of Health records the finding of *Dipylidium caninum* eggs in the stools of two patients. This is a normal and common parasite of dogs and cats (see Parasites of Dog); fleas and lice are the known intermediate hosts.

A fatal case of human cysticercosis (caused by the larval stage of *Taenia solium*) (fig. 2b) involving the brain, heart, lungs, and liver was diagnosed in 1954 at Queen’s Hospital (autopsy No. A-2316) in a 30-year-old Japanese woman born on Kauai, who had not taken a trip away from the Islands. The adult stage of this parasite occurs only in man. Swine, as well as man, may serve as an intermediate host. Man usually acquires the adult worm by eating raw or undercooked pork containing the infective cysticerci (also known as bladderworms) encysted in the musculature. Swine and man acquire the cysticercus infection by ingesting food or water contaminated by human feces containing the eggs of the parasite. Man can also acquire cysticercosis through autoinfection. The above case of human cysticercosis implies, therefore, that the adult stage of *T. solium* has been introduced into Hawaii, perhaps by Filipino workers, since the parasite is widely distributed in the Philippines (Tubangui, 1947). Its presence among local inhabitants, therefore, constitutes a public health hazard.

**FLUKES**

There are two species of flukes known to occur in man in Hawaii, namely, *Fasciola gigantica* (fig. 5) and *Stellantchasmus falcatus* (fig. 6a, d). Hall (1936) reported a case of *F. hepatica* from man in Hawaii, but this proved
to be *F. gigantica* (see Alicata, 1953a). *F. gigantica* is the common liver fluke of cattle, and infection in man is regarded as accidental. Such infection usually takes place by ingesting raw vegetation containing encysted fluke larvae (metacercariae, fig. 14b). In 1953, the writer summarized information concerning 19 cases which were known to have occurred in the Islands over a period extending from 1904 to 1951 (1953a). Three additional cases, together with their clinical and laboratory findings, were reported by Steßmann (1953a, b). He was of the opinion that human fascioliasis in Hawaii is probably more frequent than the above figures indicate. Herbert (1907) and Hall (1936) pointed out that watercress may be a source of human infection. A study carried out subsequently by Alicata and Bonnet (1955, 1956) confirmed such a possibility. It was suggested that basic methods of preventing metacercariae contamination of watercress consist of raising this vegetation with artesian water rather than using stream water and of preventing cattle from grazing on or near areas where watercress is grown for human consumption.

Intestinal heterophyidiasis, caused by *S. falcatus*, was first recorded in Hawaii in 1938 (Alicata and Schattenburg, 1938). Heavy infections of this parasite may produce severe intestinal inflammation. Local fresh-water snails, *Stenomelania newcombi* and *Thiara granifera*, serve as the first intermediate hosts (Martin, 1958; Noda, 1959). Mullet, *Mugil cephalus*, serve as
the second intermediate host, and infection with the metacercariae (fig. 6c) of this parasite is of frequent occurrence (Alicata and Schattenburg, 1938; Martin, 1958). Local cats, dogs, and rats are known to harbor this parasite in the adult stage (Ash, 1962a, b).

Martin (1958) reported natural infection of the night heron, Nycticorax nycticorax, with three species of heterophyid flukes, namely, Centrocestus formosanus, Haplorchis taichui, and H. yokogawai. C. formosanus was also recovered from the rat. Since these flukes are able to develop in man, they are of public health importance. The melanid snail, Stenomelania newcombi, serves as the first intermediate host for C. formosanus and H. yokogawai. The melanid snail, T. granifera, serves as intermediate host for H. taichui. The mosquito fishes, Gambusia affinis and Xiphophorus helleri, and, rarely, the mullet, Mugil cephalus, are natural second intermediate hosts for C. formosanus and H. taichui. Experimentally, the fish, Kuhlia sandwicensis, is also susceptible to C. formosanus infection. The fishes, G. affinis, Mollienesia formosus, and, rarely, M. cephalus, serve as natural intermediate hosts for H. taichui. Experimentally, the Chinese catfish, Clarias fuscus, has been determined as a second intermediate host for H. yokogawai.

In a survey of parasites of man in Hawaii, Ching (1961a) records the following numbers of fluke infections found among 1,380 persons examined: heterophyids, 41; Clonorchis (= Opisthorchis) sinensis, 15; Opisthorchis sp., 1. The percentages of heterophyid flukes found among the various local ethnic groups examined, listed in order of their frequency, are as follows: Hawaiian or Part-Hawaiian, 7.9; Filipino, 7.2; Chinese and Japanese, less than 0.1; Caucasian, all negative. The findings indicate that these parasites are more common among those groups that customarily consume raw fish. In most instances the heterophyid infections were probably acquired in Hawaii.

In the above study by Ching, the Oriental liver fluke, Opisthorchis sinensis, was found only among persons of Chinese ancestry born in China or who had at some time resided there. The single infection with Opisthorchis sp. was from an elderly man born in Japan. Binford (1934) reported four cases of O. sinensis among persons born and reared in Hawaii. The diagnosis was based on fluke eggs found in the feces of these patients. The illustration of the eggs published by Binford, however, is not convincing evidence that the eggs are those of O. sinensis. They do not show the pronounced shoulders at the region where the operculum rests, a feature characteristic of O. sinensis eggs. It is likely that the eggs were those of heterophyids (fig. 6b), which are somewhat similar to liver fluke eggs and which were unknown in the Islands at the time of Binford’s publication.
FIGURE 6. Heterophyid intestinal flukes, *Stellantchasmus falcatus*: a, adult, ×50; b, egg, highly magnified; c, metacercaria, encysted in the musculature of mullet, highly magnified; d, cross section of human intestine showing adult parasite embedded in mucosa, ×30. (a–c, After Alicata and Schattenburg, 1938, courtesy of *Journal of the American Medical Association*; d, original.)
The possibility of transmission of *O. sinensis* in Hawaii is uncertain. This parasite utilizes certain fresh-water snails as the first intermediate host, and a certain fresh-water fish as the second intermediate host. Man acquires the infection from eating the raw infected fish. In the Orient, three species of operculated snails of the family Amnicolidae serve as first intermediate hosts; in addition, three species of melanid snails including *Thiara granifera* have been implicated as hosts, although this is doubtful (Abbott, 1948). Of the species of operculated snails that are found in Hawaii, *Bulinus robustus minor*, *Thiara granifera*, and *Stenomelania newcombi* warrant investigation as possible intermediate hosts of *O. sinensis*. The spread and propagation of this parasite in Hawaii, however, does not seem likely because of the adequate disposal of local sewage and the usual unavailability of fresh-water fish in the markets.

A case of lung fluke, *Paragonimus westermani*, in Honolulu was reported to the writer by Min (1949), from a patient who had resided in Korea. This fluke requires certain fresh-water snails and crustaceans as first and second intermediate hosts, respectively. The melanid snail, *Thiara granifera*, has been reported as intermediate host for *P. westermani* in the Orient (Abbott, 1948). The snail, *T. granifera*, crayfish, and crabs present in the Islands could probably serve as intermediate hosts. Infection in man is acquired by eating infected fresh-water crayfish or crabs that are improperly cooked. As in *O. sinensis*, transmission of lung flukes in Hawaii is not likely because of the rarity of the infection here, and the adequate disposal of sewage.

Chu (1952) reported that certain schistosome cercariae recovered from the littorine snail, *Littorina pintado*, produce dermatitis when experimentally applied to the skin of man. These cercariae were later identified as the larval stage of *Austrobilharzia variglandis* (see Chu and Cutress, 1954). As adults, these flukes have been found in Hawaii in the mesenteric veins of the shore bird, *Arenaria interpres morinella*, known as the ruddy turnstone. Experimentally, chickens, ducks, and sooty and noddy terns have proved to be susceptible to infection. No case of human dermatitis has thus far been traced to natural infection with this parasite.

*Philophthalmus gralli*, located under the nictitating membrane of certain water birds (see Parasites of Chicken and Other Avian Hosts), and *Plagiorchis muris*, an intestinal fluke of rats (see Parasites of Rats), are potential parasites of man. Experimentally, *P. gralli* has been determined to develop in the eyes of rats and rabbits (Alicata and Ching, 1960). In Hawaii, these flukes utilize the melanid snails, *Stenomelania newcombi* and *Thiara granifera*, as intermediate hosts (Alicata and Noda, 1960). Related parasites have been reported in man for two areas, *P. lacrymosus* in Belgrade (Markovic, 1939) and *Philophthalmus* sp. in Ceylon (Dissanaike, 1958).
Several species of arthropods in the Islands are known either to be permanent external parasites or occasionally to bite or sting man intermittently or by accident. These include the following (Pemberton, 1943): the human head louse, *Pediculus humanus capitis*; the human body louse, *P. h. humanus*; the pubic louse, *Phthirus pubis*; the bed bug, *Cimex lectularius*; the human flea, *Pulex irritans*; the Oriental rat flea, *Xenopsylla cheopis*; the Hawaiian field rat flea, *X. vexabilis hawaiiensis*; the northern rat flea, *Nosopsyllus fasciatus*; the cat flea, *Ctenocephalides felis*; the mouse flea, *Lepotopsylla segnis*; the sticktight flea, *Echidnophaga gallinacea*; the daylight-feeding mosquitoes, *Aedes aegypti* and *A. albopictus*; and the night-feeding mosquito, *Culex pipiens quinquefasciatus*; the cone-nose or “kissing bug,” *Triatoma rubrofasciata*; the common stable fly, *Stomoxys calcitrans*; the honey bee, *Apis mellifera*; a species of hornet, *Vespa occidentalis*; several species of wasps of the genus *Polistes*; the common fire ant, *Solenopsis geminata*; and the antlike insect, *Holepyrus hawaiiensis*; the common centipede, *Scolopendra subspinipes*; the common brown dog tick, *Rhipicephalus sanguineus*; and the black widow spider, *Latrodectus mactans*. Included also are the brown widow spider, *Latrodectus geometricus* (see Bonnet, 1948), and the clubionid spider, *Cheiracanthium diversum*, which are known to have bitten man (Bianchi, 1959; Gressitt, 1957; Hardy, 1957). The mosquito, *Aedes vexans nocturnus*, which is a voracious feeder and attacks at dusk and in the early evening or morning, was recently accidentally introduced into Hawaii (Hardy, 1962). The thrips, *Frankliniella sulphurea*, and possibly other species, have been reported to have bitten man (Carter, 1959).

Several species of mites, which occasionally bite or are capable of biting man, include *Ornithonyssus bacoti* (see Haramoto, 1964); *Laelaps echi- dinus* and *L. nuttalli*, common on rats; and *Pediculoides ventricosus* (= *P. boylei*), a common ectoparasite of grain insects (Pemberton, 1943). Other mites found in stored grain and feeds which may cause dermatitis are *Acarus siro*, *Dermatophagoïdes scheremetewskyi*, and *Glycyphagus domesticus* (see Haramoto, 1964). Included also is the tropical fowl mite, *Ornithonyssus bursa*, commonly found on poultry, poultry houses, nests of English sparrows and mynah birds, and also occasionally found infesting human dwellings (Zimmerman, 1944). The follicular mite, *Demodex folliculorum*, appears to be of common occurrence on man in Hawaii (Haramoto, 1964).

On at least one occasion, the “sheep head maggot,” *Oestrus ovis*, larviposited in the eyes and nostrils of three men in Hawaii (Hardy, 1956).

Several of the above arthropods in Hawaii serve as intermediate hosts of parasites or as vectors of bacterial, rickettsial, and virus diseases of man. The fleas, *Ctenocephalides felis* and *Pulex irritans*, are known to be inter-
mediate hosts of the tapeworm, *Dipylidium caninum* (see Hall, 1929). According to epidemiological data, the fleas, *Xenopsylla cheopis* and *X. v. hawaiiensis*, are the principal vectors of bubonic plague (Eskey, 1934). *X. cheopis* and *Nosopsyllus fasciatus* are believed to be the principal vectors of endemic typhus (Doolittle, 1941). Experimentally, the chicken sticktight flea, *Echidnophaga gallinacea*, has been shown to serve as a suitable vector of endemic typhus (Alicata, 1942a). Dengue fever, which has occurred in Hawaii, is believed to be transmitted by the mosquito, *Aedes aegypti*.

**PARASITES OF ANIMALS**

**CAT**

**PROTOZOA**

No reports are available.

**ROUNDWORMS**

Of 107 stray cats examined, the following species and percentages of roundworms were found (Ash, 1962a): stomach worms, *Physaloptera praeputialis* (fig. 7a), 23 percent; ascarids, *Toxocara canis* (fig. 28), 8 percent; lungworms, *Aehurostrongylus abstrusus* and *Anafilaroides rostratus*, 4.0 and 2.0 percent, respectively; hookworms (fig. 7b), 58 percent. The hookworms were listed as *Ancylostoma caninum*, but these are actually *A. tubaeforme* (see Burrows, 1962). The microfilarial stage of the *Dirofilaria immitis* was seen by Ash in the peripheral blood of one cat, but no adult worms were found. Immature acanthocephalans, identified by Van Cleave (1947) as *Arythmorhynchus* sp., have been collected by the writer from the small intestine of a cat. Acanthocephalans of this genus are predominantly parasites of water birds. The cat, therefore, is believed to be an unnatural host. The ascarid, *T. canis*, is of public health interest, since its larvae are one of the chief causes of visceral larva migrans in children (see Parasites of Man).

Of the above parasites, the stomach worms and the lungworms require an intermediate host in their larval development. Experimental infection carried out elsewhere has shown that *P. praeputialis* is able to utilize the German cockroach, *Blatella germanica*, the camel cricket, *Centophilus* sp., and the field cricket, *Gryllus assimilis*, as intermediate hosts (Petri and Ameel, 1950).

The lungworm, *A. abstrusus*, is known to require snails and slugs as intermediate hosts (Hobmaier and Hobmaier, 1955; Gerichter, 1949). The land snail, *Subulina octona*, was reported by Van Volkenberg (1937) to be an intermediate host in Puerto Rico; in Hawaii, the writer (1947) infected the
land snail, *Bradybaena similari*s, with larvae of *A. abstrusus*. Cats may acquire this lungworm by eating infected mice (Cameron, 1927) or rats (Ash, 1962b). Rodents serve as paratenic hosts and acquire the infective larvae of this parasite by eating infected mollusks. In mice, the infective larvae of the lungworm migrate to and encyst in various parts of the body including musculature and omentum (Cameron, 1927; Mackerras, 1957). Ash (1962b) reported finding infective larvae of this parasite in the intercostal muscles of a rat 1 week after experimental infection.

The lungworm, *A. rostratus*, also utilizes land snails and slugs as intermediate hosts; rodents and birds can serve as paratenic hosts (Seneviratna, 1959). In rodents, the infective larvae (fig. 7c) encyst in the liver (Alicata, 1963b). Both *Subulina octona* and *Bradybaena similari*s can serve as inter-
mediate hosts in Hawaii (Ash, 1962a). Since cats do not usually eat mollusks, it appears most likely that they normally acquire lungworms from eating rodents harboring the infective lungworm larvae.

**TAPEWORMS**

Of 107 stray cats examined in Honolulu, 81 percent harbored *Dipylidium caninum* (fig. 8a) and 21 percent *Hydatigera taeniaeformis* (see Ash, 1962a). *D. caninum* utilizes fleas and lice as intermediate hosts. The adult stage of this parasite may also develop in man (see Parasites of Man). The tapeworm, *H. taeniaeformis* (fig. 8b), utilizes rats or mice as intermediate hosts. The infective larval stage (strobilocercus) is commonly found in the liver of rodents (fig. 31f) in Hawaii (see Parasites of Rat).

**FLUKES**

Two species of heterophyid flukes, *Stellantchasmus falcatus* (fig. 6a) and *Phagicola longus*, were found in the intestinal tract of stray cats examined in Honolulu. They occurred in 8 and 1 percent, respectively, of 107 cats examined. The liver fluke, *Platynosomum fastosum* (fig. 7d), was recovered from 15 percent of the same cats examined (Ash, 1962a).
S. falcatus utilizes melanid snails as the first intermediate host and mullet as the second intermediate host (see Parasites of Man). P. longus also requires fresh-water snails and fish as first and second intermediate hosts, respectively, but these have not been determined in Hawaii.

P. fastosum has been shown, in Puerto Rico, to utilize the land snail, Subulina octona, as the first intermediate host, and the lizard, Anolis cristatellus, as the second intermediate host (Maldonado, 1945). In Hawaii, in addition to S. octona, Bradybaena similaris has also been determined to be a suitable first intermediate host (Ash, 1962a).

According to the above information, cats in Hawaii normally acquire heterophyid infection from eating raw mullet and possibly other fish, and they acquire the liver fluke, P. fastosum, from eating lizards.

ARTHROPODS

The cat flea, Ctenocephalides felis, and the sticktight flea, Echidnophaga gallinacea, are common on cats in Hawaii, the former occurring under houses and in other areas frequented by cats. The biting louse, Felicola subrostrata (see Zimmerman, 1944), and the mange mite, Notoedres cati (see Haramoto, 1964), have been collected from cats.

CATTLE

PROTOZOA

Four species of coccidia, Eimeria bovis, E. bukidnonensis, E. cylindrical, and E. zurnii, have been recovered from the feces of young calves (Cuckler and Alicata, 1943). Severe cases of coccidiosis, associated with emaciation, have occasionally been noted among calves in the Islands.

Anaplasmosis, an infectious disease of cattle characterized by anemia and the presence of marginal bodies in the red cells, has been recognized in Hawaii since 1954 (Hawaii Bd. Commrs. Agr. and Forestry, 1956). The exact nature of the causative organism, Anaplasma marginale, is still undetermined, but it is believed to be a one-celled body of complex nature (Roby, 1960). It is transmitted mechanically by ticks, horseflies, and probably by other biting insects, as well as by unclean instruments (Stiles, 1946; Dikmans, 1950) which are used in dehorning, earmarking, or other surgical operations.

ROUNDWORMS

In a survey involving the examination of about 375 adult cattle raised on various Hawaiian islands and slaughtered in Honolulu, the following peri-
Figure 9. Adult parasites of cattle, natural size: a, intestinal roundworms, *Cooperia punctata*; b, stomach worms, *Haemonchus contortus*; c, hookworms, *Bunostomum phlebotomum*; d, nodular worms, *Oesophagostomum radiatum*; e, tapeworm, *Moniezia benedeni*. (Original.)
percentages of roundworm infections were found (Cuckler and Alicata, 1943):
gullet worms, *Gongylonema pulchrum*, 54.3; stomach worms, *Haemonchus contortus* (fig. 9b), 0.9; intestinal roundworms, *Bunostomum phlebotomum* (fig. 9c), 6.7; *Cooperia punctata* (fig. 9a), 4.0; *C. pectinata*, 0.3; skin filarioids, *Stephanofilaria stilesi* (fig. 10b), 89.8. *C. punctata* has also been reported from axis deer in Hawaii (see Parasites of Deer).

**Figure 10.** *a*, Skin of cattle from the region of the mid-ventral line showing characteristic lesions caused by the presence of *Stephanofilaria stilesi*; *b*, section of skin lesion showing presence of adult *S. stilesi*, highly magnified. (Original.)
In a study of parasites of 60 calves from 4 to 12 months of age, the following percentages of roundworms were found (Alicata, 1960): stomach worms, *Haemonchus contortus*, 38.3; *Ostertagia ostertagi*, 55.6; *Trichostrongylus axei*, 55.6; intestinal worms, *Bunostomum phlebotomum*, 40.0; *Cooperia punctata*, 95.0; *C. spatulata*, 13.3; *Oesophagostomum radiatum* (fig. 9d), 61.6; *Trichuris* sp., 30.0; *Strongyloides papillosus*, 56.7; lungworms, *Dictyocaulus viviparus*, 1.6.

The above study indicated that *C. punctata* is more common and more numerous among calves than any other helminth present in Hawaii and is more prevalent in areas of high rainfall. It is most common among calves 4 to 8 months of age. This parasite appears to be a major contributing factor to the unthriftiness often noted among local calves (Alicata, 1960). Calves experimentally infected with *C. punctata* have shown intermittent or continued diarrhea, progressive emaciation, reduced feed consumption, weight loss, and listlessness (Alicata and Lynd, 1961). Lungworm infection, although apparently uncommon, has on occasion been reported as producing death among calves in the Islands (Willers, 1945).

Of the above roundworms, *Gongylonema pulchrum* and *Stephanofilaria stilesi* require an intermediate host in their development. *G. pulchrum* is known to utilize one of various coprophagous beetles and cockroaches as an intermediate host in the continental United States (Alicata, 1935). Of the insects reported as hosts, *Aphodius lividus*, *Dermestes vulpinus*, and *Blatella germanica* occur in Hawaii. *S. stilesi* is known to utilize horn flies of cattle as intermediate hosts (Ivashkin et al., 1963). In the continental United States, horn flies, *Haematobia irritans*, have been found naturally infected with third-stage larvae resembling those recovered from skin lesions of infected cattle (Hibler, 1964). *S. stilesi* usually occurs under the skin in lesions along the midline on the abdominal surface of cattle (fig. 10a). The lesions vary in size up to several inches. They are hairless and the skin is frequently broken and moist with blood and serum. These lesions may sometimes dry, forming scabs.

**TAPEWORMS**

Tapeworms identified as *Moniezia benedeni* (fig. 9e) have been recovered from beef and dairy calves in Hawaii (Alicata, 1960). Experimentally, in Russia, this species of tapeworm has been reported to utilize the free-living oribatid mites, *Galumna obvius* and *Scheloribates laevigatus* (fig. 11), as intermediate hosts (Kates and Runkel, 1948; Kates, 1965). Oribatid mites of the same genera are known to occur in Hawaii (Jacot, 1934) and may serve as vectors under local conditions.

The infective larval stage (cysticercus) of *Taenia saginata* has been found occasionally in the musculature of cattle in the Islands, according to Julien.
Oribatid mites containing the infective larval stage (cysticercoids) of the cattle tapeworm, Moniezia expansa, magnified. (After Kates and Runkel, 1948, courtesy of Proceedings of the Helminthological Society of Washington.)

(1947), federal meat inspector. The larvae reach maturity in the small intestine of man following ingestion of improperly cooked beef. Cattle acquire the cysticercus infection as a result of eating vegetation contaminated with human feces containing eggs of this parasite. It is generally believed that most cases of human infection occur among immigrants from the Orient, especially from the Philippines.

FLUKES

Three species of flukes have been recorded from beef and dairy cattle in the Islands, namely, the liver flukes, Fasciola gigantica (fig. 12b) and F. hepatica, and an unidentified rumen fluke reported by Hall (1936). Of these flukes, F. gigantica is the most common. F. hepatica is probably only rarely found in Hawaii. It has been observed only once in a cow born and raised on the island of Oahu, and its introduction into Hawaii is believed to be recent (Alicata, 1952b). No liver flukes were found in 14 wild cattle examined by Swanson (1939).

Liver fluke infection is the most important parasitic disease of cattle in the Hawaiian Islands (figs. 12, 13). Of 21,198 beef and dairy cattle slaughtered in 1962 at the Hawaii Meat Company, Honolulu, 3,395, or 16 percent, showed fluke infection of the liver. The origin, number examined, and percent infection of these animals were as follows: (beef cattle) Hawaii, 13,352, 9.6 percent; Kauai, 593, 59.0 percent; Maui, 2,770, 21.4 percent; Molokai, 448, 16.0 percent; Niihau, 333, 0 percent; Oahu, 1,775, 31.5 percent; unknown origin, 558, 8.9 percent; (dairy cattle) Hawaii, 57, 10.5 percent; Maui, 93, 16.1 percent; Oahu, 1,217, 39.3 percent; unknown origin, 6, 83.3 percent (Alicata, 1964c).
Figure 12. a, Cow heavily infected with liver flukes; b, adult liver flukes, *Fasciola gigantica*, natural size; c, eggs of *F. gigantica*, highly magnified. (Original.)
Liver fluke infection in cattle was first reported by Lutz (1892) as being common on four of the larger Hawaiian islands. Although at that time the parasites were reported as *Fasciola hepatica*, more recent study has shown them to be *F. gigantica* (see Alicata and Swanson, 1937). The importation of
this fluke into Hawaii is not clearly understood, but it is believed to have come from the Orient with the introduction of water buffaloes. It is of interest to note that the limnaeid snail, *Fossaria ollula*, which serves as the intermediate host, has Japan and China as its geographic range (Alicata, 1938a). Experimentally, *F. ollula* has also been shown to serve as intermediate host for *Fasciola hepatica* (see Alicata, 1952b). This snail is widely distributed in Hawaii and is common along banks of streams and swampy lowlands (fig. 14a). In addition, another snail, *Pseudosuccinea columella*, apparently introduced recently into Hawaii, has been shown experimentally to be a suitable intermediate host for *F. gigantica* (see Alicata, 1953b).

The continuance of bovine fascioliasis in Hawaii, as elsewhere, is dependent on various factors of which topography, climatic conditions, and agricultural practices are very important. In Hawaii, the mountains descend to the ocean either abruptly or by gradual decline over relatively flat land with very little drainage. These poorly drained lowlands and valleys, especially on the windward side, often present rather extensive swamps. Rainfall is heaviest in winter months, but showers during other seasons of the year are sufficient to maintain swampy conditions. These wet areas and the mild Hawaiian climate encourage snail propagation the year round, as well as the development and hatching of the fluke eggs. Moreover, agricultural practices in the Islands have encouraged rather than hindered the maintenance of fluke infection. With ample supplies of vegetation, cattle have often been allowed to graze continuously. Many dairymen have long been in the habit of feeding cut forage from wet or swampy areas to cattle. These practices have been largely responsible for the widespread fluke infection. This disease is gradually being brought under control largely through (1) use of copper sulfate for the control of the snail vector in swamps or streams, (2) use of forage grass cut from dry areas, and (3) treatment of infected animals with hexachloroethane. This synthetic compound was first used for fluke control in Europe in 1926 (Thienel, 1926). In the United States it was first utilized on a large scale in Hawaii (Alicata, 1941a).

The possible use of snail predators for the control of limnaeid snails in Hawaii has been advocated by the writer since 1939 (Alicata, 1941a). Japanese fireflies, *Luciola cruciata* and *L. lateralis*, were introduced into Hawaii from Japan during 1939, 1940, and 1952 for the control of the snails, but none became established (Alicata and Bess, 1953). In recent years, two species of scomyzid flies, *Sciomyza dorsata* and *Sepenedon macropus*, have been imported and released in the Hawaiian Islands for the control of limnaeid snails (Davis *et al.*, 1961; Chock *et al.*, 1961). The value of these flies remains to be determined.
FIGURE 14. a, Muddy bank adjacent to a swamp where lymnaeid snails, *Fossaria ollula*, are found; b, a stem and blade of *Commelina diffusa* containing encysted metacercariae of liver flukes, *Fasciola gigantica*, ×3. (Original.)
Several arthropods are known to be parasitic on cattle. In an examination of 303 animals (Cuckler and Alicata, 1943), 26.1 percent showed evidence of the cattle grub, *Hypoderma lineatum*. This fly was first reported in the Islands in 1906 (Bryan, 1934). It appears to be restricted to the island of Hawaii at regions between 1,500 and 6,000 feet (Bess, 1951). According to Sherman (1964), entomologist, the northern cattle grub, *H. bovis*, is also present in most of the Hawaiian Islands.

Cattle grubs (fig. 15) are recognized as injurious to cattle, causing loss of flesh and decreasing the value of the skin for leather. Pickerill (1935), of the Hawaii Meat Company, reported that, during the year 1934, of 15,099 hides of island cattle examined, 4,252, or 28.16 percent, were grubby.

In recent years, a report was made of attacks of blowflies on young calves on the island of Kauai (Holdaway, 1943, 1945). Observations indicated that three species of flies were involved, *Chrysomyia megacephala*, *C. rufifacies*, and, possibly, *Lucilia sericata*. These flies ordinarily breed in carcasses and other animal matter. However, they may deposit eggs in a number of differ-
FIGURE 16.  

a. Ear of cattle cut to show ticks, Otobius megnini, in the deeper portion of the ear canal; b, fully engorged young (nymph) ear tick, greatly enlarged; c, third-larval stage of cattle grubs, Hypoderma lineatum, from the skin of cattle, slightly enlarged. (Original.)
ent places on recently born calves. The eggs hatch and the larvae or maggots feed on the surface layer and cause an inflamed, malodorous wound. Infested calves become debilitated and, unless treated, often die in a few days.

Auricular myiasis of cattle, caused by the larvae of *C. megacephala*, *C. rufifacies*, and *Fannia* sp., has been reported by Zimmerman (1944). Species of adult flies which are pestiferous on cattle in the Islands include the horn fly, *Haematobia irritans*, and the stable fly, *Stomoxys calcitrans* (see Bryan, 1934). The horn flies are known to serve as intermediate hosts for *Stephanofilaria stilesi* (see Ivashkin et al., 1963; Hibler, 1964).

The sucking louse, *Haematopinus eurysternus* (see Cuckler and Alicata, 1943), and the biting louse, *Bovicola bovis* (see Zimmerman, 1944), have occasionally been found on cattle. General emaciation or unthriftiness is usually associated with these infestations.

The spinose ear tick, *Otodectes megnini* (fig. 16), which was first noted in recent years (Alicata, 1941b; Cuckler and Alicata, 1943; Zimmerman, 1944), is widespread on beef cattle. Of 357 animals examined from Hawaii, Oahu, and Maui, 160, or 44.8 percent, showed infestation (Cuckler and Alicata, 1943). In several instances, the ticks were seen in large numbers in the ear canal (fig. 16a). These ectoparasites are known to puncture the tender skin of the ear and to suck blood. The wounds thus caused often ulcerate and a condition known as ear canker results.

**CHICKEN AND OTHER AVIAN HOSTS**

**PROTOZOA**

The coccidium, *Eimeria tenella*, which causes cecal coccidiosis, is the most important protozoan affecting chickens (fig. 17). Infection with this parasite is as troublesome in Hawaii as it is anywhere else. Undoubtedly, other species of coccidia occur among chickens in the Islands, but they have not been determined.

Pigeons in Hawaii are commonly infected with the blood parasite, *Haemoproteus columbae* (see Alicata, 1939c). Blood smears from 101 adult pigeons in Honolulu showed 83 to be infected (Kartman, 1949). *H. columbae* may produce anemia and lowered vitality. It is transmitted among pigeons through the bite of the pigeon-fly, *Pseudolynchia canariensis*, which is widely distributed (Bryan, 1954). Of a total of 25 *P. canariensis* dissected, 9, or 36 percent, were infected with *H. columbae* (see Kartman, 1949). The avian malarial organism, *Plasmodium vaughani*, in the Pekin nightingale, *Liothrix lutea*, was reported by Fisher and Baldwin in 1947. This parasite was noted in 1 of 11 birds examined from Hawaii National Park, island of Hawaii. *P. vaughani* is probably mosquito-borne and its life cycle is only partially known (Manwell, 1947; Laird, 1953).
The flagellate, *Trichomonas gallinae*, has been associated with pathological conditions of the upper digestive tract of pigeons in Hawaii (Yager and Gleiser, 1946). Trichomoniasis is ordinarily considered a disease of squabs and young pigeons and is generally less important among older birds. The disease is known to be transmittable to chickens and turkeys, and pigeons may be the source of infection.
Histomonas meleagridis, the causative organism of "blackhead," is responsible for sporadic outbreaks of this disease in turkeys (fig. 18) in various parts of the Islands. The disease is more frequent among turkeys than chickens. The parasite may be transmitted either by feed or water contaminated with droppings of infected chickens or turkeys or, most commonly, through the ingestion of eggs of the cecal worm, Heterakis gallinarum, which carry the small "blackhead" parasite (see Roundworms, below).

ROUNDWORMS

The gizzard-worm, Acuaria hamulosa (fig. 19b), is an important parasite of chickens and turkeys in Hawaii. Depending upon the degree of infection, the lining of the gizzard may contain from small to large ulcerations
which may involve the muscular tissue. Soft nodules enclosing parasites are frequently noted in the thinner muscular portion of the gizzard. When heavily parasitized, the gizzard becomes enlarged and may assume a saccular appearance (fig. 20a). Up to several years ago, these parasites were very common in the Islands and were responsible for anemia, emaciation, and death, especially among chickens. The spread and incidence of the parasite was checked following the control of various arthropods which serve as intermediate hosts. The following serve as natural or experimental hosts: (Order: Coleoptera) Carphophilus dimidiatus, Dactylosternum abdominale, Dermestes vulpinus, Epitragus direptus, Euxestus sp., Gonocephalum seri­atum, Litargus bulbeatus, Oxydema fusiforme, Palorus ratzeburgi, Sitophilus oryzae, Tenebroides nana, Tribolium castaneum, and Typhaea stercorea; (Order: Orthoptera) Atractomorpha ambigua, Conocephalus saltator, and Oxya chinensis; (Order: Amphipoda) Orchestia platensis. (See Alicata, 1936, 1938b, 1939c.)

A. hamulosa has also been reported from the Chinese ring-necked pheasant, Phasianus colchicus torquatus, but no ill effects were noted (Schwartz and Schwartz, 1951). Another species of gizzard-worm, Cyrnea graphophasi­ani, was reported from the gizzard of one of the pheasants; this was regarded as a new host for this parasite.
Lesions on the gizzard of chicken caused by *Acuaria hamulosa*: 

- **a.** Heavily infected gizzard showing enlargement of the anterior and posterior portions; 
- **b.** Lesions on the inner lining of the gizzard; 
- **c.** Cross section of gizzard showing sections of the worms within the wall. 
- **d.** Enlarged; **d.** Greatly enlarged. 

(After Alicata, 1938.)
The eyeworm, *Oxyspirura mansoni* (fig. 21a), which utilizes the burrowing roach, *Pycnoscelus surinamensis* (fig. 21b), as intermediate host, was reported by the writer (1936) as being common among chickens in the Islands. However, in more recent years, with the availability and use of several powerful insecticides such as benzene hexachloride, "Parathion," and others (Kartman et al., 1950), it has been possible to reduce the roach infestation and, as a result, to decrease the incidence of eyeworms. This parasite has also been recovered from the Chinese ring-necked pheasant (Swanson, 1939; Schwartz and Schwartz, 1951), the English sparrow (Illingworth, 1931), the mynah bird, the Chinese or lace-necked dove, and the Japanese quail (Schwartz and Schwartz, 1949, 1951). There is thus good reason to believe that wild birds serve as reservoir hosts from which infection can be acquired. An account of the life history of the eyeworm in the intermediate and final hosts in Hawaii was given by Schwabe in 1951. In the bird host, the parasites are located in the inner corner of the eye socket, between the eyeball and the nictitating membrane. In heavy infections, there is a puffiness around the eye and inflammation of the surrounding tissue (Schwabe, 1950). Infected birds blink their eyes continually, and the irritation causes the birds to scratch at their eyes with their claws for relief. The process of scratching frequently causes mechanical injury to the eyeball, which may lead to secondary bacterial and viral infections (Schwabe, 1950). Eyeworms are most
PARASITIC INFECTIONS OF MAN AND ANIMALS IN HAWAII

prevalent in dry areas with loose sandy soil, in which the roaches thrive. As a means of controlling this disease, the writer has advocated the maintenance of the giant toad, *Bufo marinus*, in poultry yards. This toad is insectivorous and devours roaches readily. Insecticides, such as lindane or chlordane, when dusted on roach-infested ground, are also effective in the control of the burrowing roach (*Alicata*, 1952a).

The poultry ascarid, *Ascaridia galli* (fig. 22), and two species of cecal worms, *Heterakis gallinarum* (fig. 19a) and *Subulura brumpti*, are common among chickens. *S. brumpti* is the most prevalent, and, unlike *H. gallinarum*, requires an intermediate host, which may be any one of the following (*Alicata*, 1939a; *Cuckler and Alicata*, 1944): (Order: Coleoptera) *Alphitobius diaperinus, Ammophorus insularis, Dermestes vulpinus, Gonocephalum seriatum*, and *Tribolium castaneum*; (Order: Orthoptera) *Conocephalus saltator, Oxya chinensis*; (Order: Dermaptera) *Euborellia annulipes, Subulura brumpti, Heterakis gallinarum*, and an undetermined species of *Ascaridia*.

Figure 22. Adult intestinal roundworms, *Ascaridia galli*, natural size. (Original.)
(probably *A. galli*) have been reported from the Chinese ring-necked pheasant also (Schwartz and Schwartz, 1951). *A. galli* has been reported from a wide variety of birds, both domestic and wild; however, very little cross-infection work has been done with this ascarid from either domestic or wild game birds (Wehr, 1940). In chickens and turkeys, the cecal worm, *H. gallinarum*, is known to play an important part in the spread of “blackhead” disease although it is not the only factor. The blackhead organism, *Histomonas meleagridis*, may pass out of the bird’s body enclosed in the egg of the cecal worm, and in this way it is protected from adverse climatic or environmental conditions.

The intestinal roundworm, *Ornithostrongylus quadriradiatus*, occurs commonly in pigeons in Hawaii and is believed responsible for unthriftiness and losses among these birds (Alicata, 1939c). The biology and pathogenicity of this parasite were described by Cuvillier (1937).

Other roundworms of chickens which require an intermediate host include the crop worm, *Gongylonema ingluvicola*, and the proventricular worms, *Tropisurus americanus* and *Dispharynx nasuta* (fig. 23a). *D. nasuta* is also known elsewhere to infect turkeys and pigeons. In the continental United States, *G. ingluvicola* utilizes the beetle, *Copris minutus*, as an intermediate host; in Hawaii, the related beetle, *C. incertus*, serves as a suitable host. *T. americanus* utilizes any of the following arthropods as intermediate hosts:

![Figure 23. a, Adult stomach worms, Dispharynx nasuta, natural size; b, adult sow bugs, Porcellio laevis, which serve as intermediate hosts, natural size. (Original.)](image)
hosts in the Islands (Alicata, 1938c): (Order: Coleoptera) *Dendrophilus* sp. (probably *D. punctatus*), *Der- mestes vulpinus*, *Epitragus duremptus*, and *Gonocephalum seriatum*; (Order: Orthoptera) *Blatella germanica* and *Conocephalus saltator*; (Order: Dermaptera) *Euborellia angulipes*; (Order: Amphipoda) *Orchestia platensis*. The sow bug, (Order: Isopoda) *Porcellio laevis* (fig. 23b), serves as intermediate host for *D. nasuta*; this parasite often produces deep ulcerations of the proventricular wall (Alicata, 1938c). In the continental United States, *D. nasuta* is an important parasite of several wild game birds, including the grouse, Hungarian partridge, bobwhite quail, and pheasant (Wehr, 1940).

Kartman (1951) reported a species of *Tetramera* (= *Tropisurus*) in the proventriculus of 22.2 percent of English sparrows examined, and he experimentally determined that two species of grasshoppers, *Conocephalus saltator* and *Oxya chinensis*, may serve as intermediate hosts. Attempts to infect chicks with the infective third-stage larvae of this parasite proved unsuccessful. Alicata, Kartman, and Fisher (1948) reported a species of *Microtetramer* from the proventriculus of mynahs.

**Tapeworms**

Tapeworms are of common occurrence in chickens. Those known in Hawaii include the following: *Choanotaenia infundibulum* (fig. 24b), *Echinolepis carioca*, *Hymenosphenacanthus exigus* (fig. 24a), *Raillietina cesticillus*, and *R. tetragona* (see Alicata, 1938c).

Various arthropods in Hawaii serve as intermediate hosts for the above-mentioned tapeworms, as follows (Alicata, 1938c; Hall, 1929): for *C. infundibulum*: (Order: Coleoptera) *Der- mestes vulpinus*, *Epitragus duremptus*, *Gonocephalum seriatum*, and (Order: Diptera) *Musca domestica*; for *E. carioca*: (Order: Coleoptera) *Aphodius granarius* and (Order: Diptera) *Stomoxys calcitrans*; for *H. exigus*: (Order: Amphipoda) *Orchestia platensis*; for *R. cesticillus*: (Order: Coleoptera) *Der- mestes vulpinus* and *Gonocephalum seriatum*; and for *R. tetragona*: probably various species of ants (Order: Hymenoptera), especially those of the genera *Pheidole* and *Tetramorium*. Members of this group of ants (*Pheidole* sp. and *T. caespitum*) are known to be intermediate hosts of *R. tetragona* in the continental United States (Jones and Horsfall, 1935).

Tapeworm infection has been common among some of the local wild game birds, namely the Chinese dove, *Streptopelia chinensis*, and the feral pigeon, *Columba livia*, according to Schwartz and Schwartz (1949), but the names of the parasites were not determined. *Raillietina* sp. has been noted among local pigeons (Yager and Gleiser, 1946). A tapeworm collected from the intestine of the Hawaiian duck has been identified by McIntosh (1935) as *Hymenolepis* (= *Clacotaenia*) megalops.
FLUKES

The cecal fluke, *Postharmostomum gallinum* (fig. 25b), commonly infects adult chickens raised on the ground. Extensive cecal hemorrhages have been found to be associated with this parasite. Studies have shown that the snail, *Bradybaena similaris*, serves as first and second intermediate host, and *Subulina octona* as second intermediate host (Alicata, 1940). The adolescaria, which is infective to chickens, occurs in the pericardial cavity of the second intermediate host (fig. 25a).
FIGURE 25. *a*, Life cycle of the cecal fluke, *Postharmostomum gallinum*, which is as follows: Fluke egg (1) containing miracidium is swallowed by the snail, *Bradybaena similaris*. Egg hatches in the digestive tract; miracidium (2) migrates to the liver and develops into a branched sporocyst (3), which gives rise to cercariae (4). Cercariae leave the sporocyst and crawl out of the snail. Cercariae, then, may enter the renal duct of the same snail or that of another snail such as *Subulina octona*. From the renal duct the cercariae reach the pericardial cavity and develop to the adolescara stage (5) which is infective to chickens. *b*, Adult cecal fluke, *P. gallinum*, ×1½. (Original.)

Martin (1958) reported the night heron as natural host for the heterophyid flukes, *Centrocestus formosanus*, *Haplorchis taichui*, and *H. yokogawai*. All of these utilize melanid snails and fish as first and second intermediate hosts, respectively (see Parasites of Man).

Figure 26. a, Adult eye-fluke, *Philophthalmus gralli*, ×20; b, encysted infective larval stage (metacercariae), ×20; c, head of chicken, experimentally infected with eye-flukes, showing congestion of the eyelids. (After Alicata. 1962, courtesy of *Journal of Parasitology*.)
Young chickens and ducks have been infected experimentally with the eye-fluke, *Philophthalmus gralli* (fig. 26a), by feeding the birds encysted cercariae (fig. 26b) derived from naturally infected melanid snails, *Thiara grantera* and *Stenomelania newcombi* (see Alicata and Noda, 1960; Ching, 1961b; Alicata, 1962b). Rabbits have been infected experimentally by placing the active cercariae on the surface of the eye. Heavily infected chickens and rabbits show considerable congestion and exudate (fig. 26c). Natural infections of this parasite occur among the Hawaiian coot, *Fulica americana alai* (see Alicata and Noda, 1960). This is a large bird inhabiting the emergent aquatic vegetation or heavy stands of grass of coastal areas. Accidental human infection with this parasite is possible (see Parasites of Man).

**ARTHROPODS**

Various species of biting lice infest poultry in the Islands. These include the following (Illingworth, 1928a): chicken body louse, *Menacanthus stramineus*; chicken head louse, *Cyclotogaster heterographa*; common hen louse, *Menopon gallinae*, also found on ducks, guinea fowl, pigeons, and turkeys; fluff louse of chickens and turkeys, *Goniocotes gallinae*; large chicken louse, *Goniodes gigas*; large turkey and chicken louse, *Chelopistes melagridis*; peafowl and guinea fowl louse, *Menopon phaeostomum*; turkey louse, *Oxyliepeurus polytrapezius*; and the chicken wing louse, *Lipeurus caponis*. Also, the biting lice of pigeons, *Columbicola columbae* and *Colpocephalum turbinatum*, and the chicken louse, *Goniodes dissimilis*, have been reported (Zimmerman, 1948).


Several species of biting lice and mites have been listed by Zimmerman (1948) from the following wild birds: “amakih,” *Philopterus macgregori* and *Machaerilaemus hawaiensis*; “apapane,” *Myrsidea cyrtostigma*; barred dove, *Cumbicola columbae* and *Goniocotes chinensis*; “dove,” *Goniodes lativentris*; English sparrow, *Brueelia vulgata*; California valley quail, *Goniodes mammillatus*; Chinese dove, *Myrsidea invadens*, *Cumbicola columbae*, and *Goniocotes chinensis*; Chinese ring-necked pheasant, *Lipeurus caponis* and
Goniocotes hologaster; guinea fowl, Goniodes gigas; gray-backed tern, Saemundssonia snyderi and Quadraceps birostris; Hawaiian coot, Rallicola advena and Quadraceps oraria; Hawaiian owl, Colpocephalum brachysomum; "iiwi," Myrsidea cyrtostigma and Colpocephalum hilensis; linnet, Colpocephalum discrepans and Philopterus subflavescens; mynah, Myrsidea invadens; noddy tern, Colpocephalum discrepans, Actornithophilus epiphanes, and Quadraceps separata; Pacific golden plover, Colpocephalum brachysomum, Quadraceps birostris, and Saemundssonia conicus; ricebird, Philopterus subflavescens and Brueelia stenozona; wandering tattler, Saemundssonia conicus and Actornithophilus kilauensis.

Several species of biting lice and mites have been reported from the following wild birds trapped on poultry farms (Alicata, Kartman, and Fisher, 1948): barred dove, (lice) Columbicola columbae and Goniodes sp., (mites) Ornithonyssus bursa; Brazilian cardinal, (lice) Myrsidea incerta; Chinese dove, (mites) Pterolichus sp.; English sparrow, (lice) Myrsidea sp., (mites) Atricholaelaps megaventralis (= Haemolaelaps casalis), Neonyssus sp., and Proctophyllodes truncatus; Japanese white-eye, (mites) Dermoglyphus elongatus, Megninia sp., Pteronyssus sp. (?), and Trouessartia sp.; mynah, (lice) Menacanthus spinosus and Myrsidea invadens, (mites) Montesauria sp., Pteronyssus sp., and Trouessartia sp.

Mites found on chickens include: the red mite, Dermanyssus gallinae, the wing mite, Pterolichus obtusus, and the body mite, Megninia cubitalis (see Alicata et al., 1946). Also included is the tropical fowl mite, Ornithonyssus bursa; this has also been reported as common in nests of English sparrows and mynahs. It is known to invade houses, where it bites human beings and causes skin irritation (Zimmerman, 1944).

The following species of mites have been reported on the Chinese ring-necked pheasant in Hawaii (Schwartz and Schwartz, 1951): Megninia ginglymura and M. columbae. The latter was also recovered from pigeons. Two other species of mites include Gabucinia sp. (see Baker, 1956) and the feather mite, Falculifer rostratus (see Yager and Gleiser, 1946).

Other arthropods reported include: the sticktight flea, Echidnophaga gallinacea, on chickens (Illingworth, 1916) and on the California quail (Schwartz and Schwartz, 1951); the pigeon-fly, Pseudolynchia canariensis, generally widespread among pigeons (Bryan, 1934); the hippoboscid fly, Ornithoiea vicina, on the Chinese ring-necked pheasant, the Japanese quail, the jungle fowl (Schwartz and Schwartz, 1951), the English sparrow (Alicata, Kartman, and Fisher, 1948), and on Zosterops sp. (see Hardy, 1952), presumably the Japanese white-eye; the soft-bodied tick, Ornithodorus capensis, which feeds on a variety of birds, has been found in crevices on Manana Island (Strandtmann, 1963). O. capensis has also been recorded from Laysan Island of the Hawaiian Archipelago (Butler, 1961).
DEER

PROTOZOA

No reports are available.

ROUNDWORMS

*Cooperia punctata* is the only roundworm recorded from axis deer in the Hawaiian Islands. The infection was noted in one deer each from the islands of Lanai and Molokai (Ash, 1961). *C. punctata* is a common parasite of calves in Hawaii (*see Parasites of Cattle*). It is likely that the deer acquired the infection from foraging in localities formerly grazed by cattle. This finding leads to the belief that, under field conditions, deer can serve as a reservoir host for this parasite.

TAPEWORMS AND FLUKES

No tapeworms or flukes have been reported. Swanson (1939) examined 8 deer, 14 wild cattle, and about 20 goats for the common cattle liver fluke, *Fasciola gigantica*, but none were found. He was of the opinion (1936) that since these animals do not usually range in fluke-infested areas, they are probably of little importance as reservoir hosts for the cattle liver fluke.

ARTHROPODS

No reports are available.

DOG

PROTOZOA

Canine coccidiosis occurs in dogs in Hawaii, but the species involved is unknown.

ROUNDWORMS

In a study of helminths from 96 dogs in Honolulu, the following species and percentages of roundworms were found (Ash, 1962a): hookworms, *Ancylostoma caninum*, 71.0 percent; subcutaneous tissue filarioids, *Dipetalonema* sp., 19.0 percent; heartworms, *Dirofilaria immitis* (fig. 27a, b), 23.0 percent; ascarids, *Toxocara canis* (fig. 28), 24.0 percent; whipworms, *Trichuris vulpis*, 3.0 percent. The ascarid, *Toxascaris leonina*, was reported by Hall (1936). In a check list of parasites of dogs and cats, Becklund (1964) lists the dog lungworm, *Filaroides osleri*, from Hawaii. *T. canis* is of public health interest since the infective (second-stage) larvae (fig. 28d) are one of the chief causes of visceral larva migrans in children (*see Parasites of Man*). At room
Figure 27. Dog heartworms, *Dirofilaria immitis*: a, heart of a dog, cut open to show a cluster of adult heartworms, slightly enlarged; b, adult male (*top*) and female worms, natural size. (a, Courtesy of U. S. Department of Agriculture; b, original.)
Figure 28. Intestinal roundworms, *Toxocara canis*: a, adults, natural size; b, egg, undeveloped, ×335; c, egg, embryonated, ×335; d, infective (second-stage) larva, ×400. (Original.)
temperature (26° C) the eggs of *T. canis* embryonate and contain infective larvae in about 9 days; these larvae are approximately 0.35–0.38 millimeter long by 0.02 millimeter wide (Alicata, 1964c).

Of the above roundworms, the heartworm, *D. immitis*, utilizes mosquitoes as intermediate hosts. Of the four species of mosquitoes known to be present in Hawaii, namely, *Culex pipiens quinquefasciatus*, *Aedes aegypti*, *A. albopictus*, and *A. vexans nocturnus*, all have been incriminated as intermediate hosts of heartworms (Hall, 1929; Kartman, 1953; Yen, 1938). In recent years, *A. aegypti* has not been found present on the islands of Oahu and Kauai (Nakagawa and Hirst, 1959). The filarioid nematode, *Dipetalonema* sp. (possibly *D. reconditum*), is known to utilize the fleas, *Ctenocephalides canis* and *C. felis*, as intermediate hosts (Newton and Wright, 1956). Of these fleas, *C. felis* occurs in Hawaii. The life history of *Filaroides osleri* is unknown; it is probable that it utilizes mollusks as intermediate hosts, as does the related species, *F. martis* (see Dubnitski, 1955, cited by Yamaguti, 1961a).

**TAPEWORMS**

*Dipylidium caninum* is the only tapeworm recorded from dogs in Hawaii. Of 82 dogs examined in Honolulu, 85.0 percent were infected (Ash, 1962a). This tapeworm utilizes fleas and lice as intermediate hosts (Hall, 1929). *Ctenocephalides felis* (see Pemberton, 1926) and *Trichodectes latus* (see Swezey, 1931), which could serve as hosts, are found on dogs in Hawaii. *D. caninum* also occurs in man. The infective larval stage (cysticercus) of *Taenia hydatigena* has been observed on several occasions attached to the liver and omentum of swine (fig. 40b) and sheep in the Islands; from this finding it may be inferred that the adult stage of this parasite is present in dogs.

**FLUKES**

Two species of heterophyid flukes, *Stellantchasmus falcatus* and *Phagicola longus*, were found in 5 and 3 percent, respectively, of 96 dogs examined by Ash (1962a). Dogs, as well as cats, may, therefore, serve as an important reservoir for human infection in this area.

*S. falcatus* utilizes melanid snails as the first intermediate host and mullet as the second intermediate host (see Parasites of Man). *P. longus* also requires a fresh-water snail and fish as first and second intermediate hosts, respectively, but the species of these hosts have not been determined in Hawaii.

**ARTHROPODS**

Arthropods present on dogs in Hawaii include the following: fleas, *Ctenocephalides felis* (see Pemberton, 1926) and *Echidnophaga gallinacea*; biting
lice, Trichodectes canis (see Swezey, 1931); species of kangaroo lice, Heterodoxus longitarus, collected from a dog in Honolulu (Pemberton, 1934); sucking lice, Linognathus setosus (see Zimmerman, 1948); and ticks, Rhipicephalus sanguineus (see Van Zwaluwenburg, 1934).

GOAT

PROTOZOA

No reports are available.

ROUNDWORMS

The following species of roundworms, collected by Lyman Nichols, wildlife biologist, Hawaii Department of Agriculture, and identified by the writer, were found in wild goats in Hawaii: stomach worms, Haemonchus contortus; stomach and intestinal worms, Ostertagia circumcincta and Trichostrongylus colubriformis; and whipworms, Trichuris ovis.

TAPEWORMS AND FLUKES

No tapeworms or flukes have been reported from goats in the Islands. Swanson (1939) examined about 20 goats for the common cattle liver fluke, Fasciola gigantica, but none were found.

ARTHROPODS

The sucking louse, Linognathus africanus, and the biting louse, Bovicola caprae, have been reported from goats (Zimmerman, 1944).

HORSE

PROTOZOA

No reports are available.

ROUNDWORMS

According to a survey (Foster and Alicata, 1939), horses in Hawaii harbor at least 25 species of roundworms as follows: Alfortia edentata, Cyathostomum asymmetricum, C. bicornatum, C. calicatum, C. catinatum, C. coronatum, C. euproctum, C. goldi, C. leptostomum, C. longibursatum, C. nassatum, C. pateratum, Delafondia vulgaris, Gyaloccephalus capitatus, Habronema microstoma, H. muscae, Oxyuris equi, Parascaris equorum, Proteostomum imprudentatum, Probstrmayria vivipara, Strongylus equinus, Trichostrongylus axei, Triodontophorus brevicauda, and T. serratus. Cyathostomum insigne has also been recorded (Swanson, 1939).
Of the above parasites, *D. vulgaris* (fig. 29a) is of somewhat common occurrence. This fact suggests that verminous arthritis and aneurysm, believed to be caused by the larval stages of this roundworm (La Page, 1956), may not be infrequent among horses in the Islands.

The roundworms of the genus *Habronema* listed above utilize various species of flies as intermediate hosts (Hall, 1929). In Hawaii, the house fly, *Musca domestica*, may transmit *H. muscae* and *H. microstoma*, and the stable fly, *Stomoxys calcitrans*, may transmit *H. microstoma*.

**TAPEWORMS**

Two species of tapeworms, *Anoplocephala perfoliata* (see Foster and Alicata, 1939) and *A. magna* (fig. 29b) (Swanson, 1939), have been reported from horses in the Islands. In Russia, under laboratory conditions, free-living oribatid mites of the genus *Scheloribates* have been found to serve as intermediate hosts for these two species of tapeworms. In Russia, experimentally, mites of the genera *Galumna* and *Achipteria* have been found to serve as intermediate hosts for *A. perfoliata* (see Kates and Runkle, 1948; Kates, 1965). Oribatid mites of the genera *Galumna* and *Scheloribates*, which are known to occur in Hawaii (Jacot, 1934), may possibly serve as vectors under local conditions.

**FLUKES**

According to a report by Hall (1936), liver flukes collected in 1894 from a horse in Honolulu were sent to the United States Bureau of Animal Industry. These flukes were originally diagnosed as *Fasciola hepatica*, but a more recent re-examination by McIntosh (1935) revealed that they are *F. gigantica*. Moreover, veterinarians on the island of Kauai have verbally reported to the writer the finding of fascioloid flukes in livers of horses. In recent years, examination by the writer of the livers of five horses pastured in areas known to be fluke-infested failed to reveal liver fluke infection. In addition, a horse and a mule experimentally fed 650 to 2,300 infective liver fluke cysts, respectively, failed to show evidence of flukes or fluke lesions when autopsied a few months later (Alicata and Swanson, 1938). It appears that horses only rarely become infected with liver flukes.

**ARTHROPODS**

The larvae of the botflies, *Gasterophilus intestinalis* and *G. nasalis*, are commonly found attached to the stomach wall (fig. 30) of horses in the Islands (Foster and Alicata, 1939). The adult stable fly, *Stomoxys calcitrans*, is also pestiferous on horses. The biting horse-louse, *Bovicola equi*, and the sucking louse, *Haematopinus asini*, have been seen on U. S. Army horses brought to the Islands (Zimmerman, 1948).
Fig. 29. a, Portion of cecum of a horse showing a mixed infection of Strongylus equinus and Delafondia vulgaris, natural size; b, adult tapeworms, Anoplocephala magna, from a horse, about one-half natural size. (After Schwartz et al., 1948, U. S. Department of Agriculture, Circular 148.)

MONGOOSE

PROTOZOA

No reports are available.

ROUNDWORMS

The infective stage of Trichinella spiralis was found in 21.4 percent and 9.1 percent of the mongooses examined on the islands of Hawaii and Maui, respectively (Alicata, 1938e).
TAPEWORMS AND FLUKES

No reports are available.

ARTHROPODS

The following species of fleas have been reported from the mongoose in Hawaii (Pemberton, 1945): Ctenocephalides felis, Echidnophaga gallinacea, Leptopsylla segnis, Nosopsyllus fasciatus, Xenopsylla cheopsis, and X. vexabilis hawaiensis. The mange mite, Notoedres cati, has also been collected from the mongoose (Haramoto, 1964).

MOUSE

PROTOZOA

No reports are available on protozoan parasites of mice in Hawaii. Trypanosoma conorhini, a blood parasite of an unknown vertebrate, has been
reported from the reduviid bug, *Triatoma rubrofasciata*, collected under a chicken coop on the island of Oahu (Wood, 1946). This parasite has been experimentally grown in mice and rats and propagated in culture media (Johnson, 1947).

**Roundworms**

Of 343 mice examined from various islands of Hawaii, none showed infection with trichina larvae (Alicata, 1938c).

**Tapeworms**

The liver of mice in Hawaii has been found to be commonly infected with the infective stage (strobilocercus) of the cat tapeworm, *Hydatigera taeniaeformis*.

**Flukes**

No reports are available.

**Arthropods**

The following fleas have been reported from mice in Hawaii (Pemberton, 1945): *Ctenocephalides felis*, *Leptopsylla segnis*, *Nosopsyllus fasciatus*, *Xenopsylla cheopis*, and *X. vexabilis hawaiiensis*. Mites reported from mice in Hawaii include: *Myobia musculi* (see Baker, 1956), *Myocoptes musculinus*, *Radfordia affinis* (see Joyce, 1957), and *Listrophorus* sp. (Joyce, 1959).

**Rabbit**

**Protozoa**

*Encephalitozoon cuniculi* has been found in the apparently normal tissue of the brain of local rabbits (Shoho, 1964). This microsporidian parasite, though frequently innocuous, is able to produce paralysis and death to rabbits. The mode of transmission is not well understood.

Liver coccidiosis caused by *Eimeria stiedae* is of common occurrence in domestic rabbits in Hawaii. Infected animals frequently show whitish spots on the surface of the liver. These spots contain masses of coccidial oocysts. The parasite is usually transmitted either in feed or water contaminated with droppings of infected rabbits.

**Roundworms**

Pinworms, *Passalurus ambiguus*, have occasionally been noted in the cecum and large intestine of domestic rabbits in the Islands (Alicata, 1964c).

**Tapeworms and Flukes**

No reports are available.
The mite, *Psoroptes equi cuniculi*, which causes ear mange, is the most important external parasite affecting domestic rabbits. It is as troublesome in Hawaii as in other areas. The inflammatory reaction produced by the mite causes a brownish discharge which cakes inside of the ears. Affected animals frequently shake their head and try to scratch their ears with their hind feet. The mite, *Notoedres cati cuniculi*, has also been collected from the face of the rabbit (Haramoto, 1961). Mites are transmitted by contact.

**Protozoa**

The blood flagellate, *Trypanosoma lewisi*, has been reported from wild rats inhabiting a gulch in the Hamakua District of the island of Hawaii (Kartman, 1954). The incidence of infection among the field rat, *Rattus exulans*, was said to be almost four times that of *R. norvegicus* and about two times that of *R. rattus* and its subspecies. On the basis of epizootiological evidence, it was suggested that the rat flea, *Xenopsylla vexabilis hawaiensis*, is the principal intermediate host. *Trypanosoma conorhini*, a blood parasite of an unknown vertebrate, has been reported from the reduviid bug, *Triatoma rubrofasciata*, collected under a chicken coop on the island of Oahu (Wood, 1946). This parasite has been grown experimentally in rats and mice, and in culture media (Johnson, 1947).

**Roundworms**

In a survey of parasites of rats in Honolulu, the following species and percentages of roundworms were found (Ash, 1962b): stomach worms, *Gonoglyonema neoplasticum* (fig. 31a), 53; *Physaloptera muris-brazilensis* (fig. 31c), 37; intestinal worms, *Heterakis spumosa*, 46; *Nippostrongylus brasiliensis*, 17; *Strongyloides ratti*, 17; *Syphacia obvelata*, 44; urinary bladderworm, *Trichosomoides crassicauda* (fig. 31b), 17; lungworm, *Angiostrongylus cantonensis* (fig. 31d), 12; liver capillarid, *Capillaria hepatica* (fig. 31e), 28. The intestinal capillarid, *Capillaria travereae*, and the acanthocephalan, *Moniliformis moniliformis*, were also reported. A fatal case of *C. hepatica* infection has also been reported from a child in Hawaii (see Parasites of Man).

In addition to the above, *Trichinella spiralis* (fig. 32a) occurred in 2.7 percent of the rats examined from the island of Hawaii, and in 0.09 percent from the island of Maui (Alicata, 1938e). No trichinae have been found among rats on the islands of Oahu and Kauai. This parasite occurs in man and swine in Hawaii (see Parasites of Man, and Swine).
Figure 31. Parasites of the rat: a, adult Gongylonema neoplasticum; b, adult bladder-worms, Trichosomoides crassicauda; c, adult stomach worms, Physaloptera muris-brasiliensis; d, adult lungworms, Angiostrongylus cantonensis; e, liver showing clusters (arrow) of eggs and adults of Capillaria hepatica; f, liver showing (arrow) encysted infective larval stage (strobilocercus) of the cat tapeworm, Hydatigera taeniaeformis. All natural size. (Original.)
Figure 32. a, Infective larvae of Trichinella spiralis encysted in the diaphragm of rat, highly magnified; b, adult tapeworm, Hymenolepis nana, natural size; c, adult tapeworm, Hymenolepis diminuta, natural size. (Original.)

Of the above roundworms, Gongylonema neoplasticum utilizes certain cockroaches and beetles as intermediate hosts. These include Blatella germanica, Periplaneta americana, and Tenebrio molitor (see Hall, 1929), all of which occur in Hawaii. According to O'Dea (1964), the stomach worm, Physaloptera muris-brasiliensis, has been experimentally determined to utilize the following arthropods as intermediate hosts: (Order: Coleoptera) Dermestes vulpinus, Tenebroides nana, and Tribolium castaneum; (Order: Orthoptera) Nauphoeta cinerea and Periplaneta americana.

The lungworm, Angiostrongylus cantonensis, utilizes a mollusk as intermediate host (pp. 24, 26 and fig. 3). The development of this parasite to the infective or third-larval stage (fig. 33e) in the garden slug, Deroceras laeve, was first described by Mackerras and Sandars (1955). These writers also traced the development of the parasite in the rat host and determined that during larval development it invaded the brain and produced dilation of the meningeal vessels and leucocytic infiltration. The rat lungworm was first found in Hawaii by Ash in November, 1960 (Ash, 1962b). Subsequently the giant African snail, Achatina fulica, the garden snails, Bradybaena similaris and Subulina octona, and the garden slug, Veronicaella alte, were found to be suitable ex-
**Figure 33.** Larvae of *Angiostrongylus cantonensis*: a) first-stage larva recovered from feces of rat, ×300; b) full-grown first-stage larva from snail, ×300; c) second-stage larva enclosed within cuticle of first molt from snail, ×300; d) third-stage larva enclosed within cast cuticles of the first and second molt from snail, ×300; e) third-stage larva coiled in the musculature of snail, ×300; f) anterior end of third-stage larva showing the characteristic sclerotized stomatorhabdions in buccal cavity, ×640. (a–e, Original; f, after Alicata, 1962, courtesy of Canadian Journal of Zoology.)
peridental intermediate hosts (Alicata, 1962a). Included also is the garden snail, *Opeas javanicum*, and possibly other members of this genus. According to Kondo (1961), malacologist, Bishop Museum, seven species of *Opeas* occur in Hawaii as follows: *O. beckianum*, *O. clavulinum*, *O. goodalli*, *O. javanicum*, *O. mawitianum*, *O. opavanum*, and *O. opella*. The fresh-water snail, *Fossaria ollula*, was also found to be a suitable experimental host (Alicata and Brown, 1962). Of the above mollusks, *A. fulica*, *B. similis*, *S. octona*, *O. javanicum*, *V. alte*, and *D. laeve* have been found naturally infected with the larvae of the rat lungworm. The land planarian, *Geoplanus septemlineatus*, in Hawaii also frequently harbors the infective lungworm larvae (Alicata, 1962a). Planarians, however, serve only as paratenic or transport hosts and acquire the larvae from feeding on the bodies of naturally infected snails. *A. cantonensis* is able to invade the brain of man and of the monkey and to produce cerebral angiostrongylosis (parasitic eosinophilic meningoencephalitis) (see Parasites of Man).

In addition to Hawaii, *A. cantonensis* has been reported among rats from other islands of the Pacific and parts of Southeast Asia, as follows (fig. 4): Espiritu Santo, New Hebrides (Alicata, 1963a); Formosa (Yokogawa, 1937); Guadalcanal, Solomon Islands (Loison, 1964); Guam (Lindquist and Li, 1955); Lifou, Loyalty Islands (Alicata, 1963a); Malaya (Schacher and Cheong, 1960); New Caledonia (Alicata, 1963a); Moen, Pingalap, and Ponape, Caroline Islands (Jackson, 1962); Rarotonga, Cook Islands (Alicata and McCarthy, 1964); Rota, Saipan, and Tinian, Mariana Islands (Alicata, 1964c); Tahiti (Alicata, 1962a); China (Chen, 1935); and Thailand (Punyagupta, 1964). In addition to rats of the genus *Rattus*, *A. cantonensis* has also been reported from the bandicoot rat, *Bandicota indica nemorivaga*, in Formosa (Kuntz and Myers, 1964).

**TAPEWORMS**

In a survey conducted by Asli (1962b), *Hymenolepis nana* (fig. 32b) and *H. diminuta* (fig. 32c) were recovered in approximately 50 percent of the rats examined in Honolulu. The infective stage (strobilocercus) of the cat tapeworm, *Hydatigera taeniaeformis*, was found in the liver of about 40 percent of the rats examined (fig. 31f). The high incidence of this larval parasite in the rat corresponded with the frequency of occurrence of the adult parasite in the cat (see Parasites of Cat). *H. nana* also has been found in man in Hawaii (see Parasites of Man).

Although most tapeworms have an indirect life cycle, *H. nana* can have either a direct or an indirect life cycle. In the former, the eggs are ingested by the definitive host and the young larvae penetrate the intestinal wall to form a tailless cysticercoid. These eventually emerge into the lumen of the
intestine and develop into sexually mature adults. In the indirect cycle, species of fleas such as Nosopsyllus fasciatus and Xenopsylla cheopis, or the grain beetle, Tenebrio molitor, serve as intermediate hosts (Hall, 1929). Voge and Heyneman (1957) studied the development of H. nana and H. diminuta in experimentally infected beetles, Tribolium confusum.

H. diminuta utilizes a large number of arthropods as intermediate hosts, including certain species of beetles, dermapterans, fleas, lepidopterans, and myriapods (Hall, 1929).

FLUKES

Plagiorchis muis was found by Ash (1962b) in 1 out of 75 rats he examined in Honolulu. Unpublished data by Noda (1958) stated that rats obtained from rural areas, particularly near bodies of water, were frequently infected with the heterophyid fluke, Stellantchasmus falcatus (fig. 6a). It appears that rats, as well as cats and dogs, serve as reservoir hosts for S. falcatus infection of man in Hawaii. Martin (1958) reported rats as natural hosts for Centrocestus formosanus in Hawaii.

An adult echinostomid fluke, identified as Echinostoma sp. and closely resembling E. ilocanum, was recovered from the small intestine of a white rat experimentally fed encysted metacercariae found in naturally infected fresh-water snails, Fossaria ollula, collected by the writer in the Kailua district of the island of Oahu. The metacercariae possessed about 47 uninterrupted collar spines, and the eggs of the adult worm measured 0.093 to 0.108 millimeter long by 0.070 to 0.074 millimeter wide (Alicata, 1964c).

Of the above flukes, the life history of P. muis in Hawaii has not as yet been determined. The first intermediate host of this parasite in the continental United States is the snail, Stagnicola emarginata angulata (see McMullen, 1937a); and in Japan, Lymnaea viridis (see Tanabe, 1922). The metacercariae encyst either in the snail host or in various species of aquatic insect larvae, which serve as secondary intermediate hosts. The final hosts reported in other areas include birds, man, mice, and rats (McMullen, 1937a, b). S. falcatus and C. formosanus utilize melanid snails and mullet as first and second intermediate hosts, respectively (see Parasites of Man).

ARTHROPODS

Seven species of fleas are known to occur on rats in Hawaii (Pemberton, 1945). These include the following: Ctenocephalides felis, rarely found on rats and mice, but common on cats, dogs, and mongooses; Echidnophaga gallinacea, also found on poultry, dogs, and mongooses; Leptopsylla segnis, uncommon on rats, but common on cats, dogs, and mice; Nosopsyllus fasciatus, found mostly at about 2,500 feet elevation on the islands of Maui and
Hawaii and also rarely found on mice and mongooses; *Pulex irritans*, rare on rats, and also found on cats, dogs, and man; *Xenopsylla cheopis* and *X. vexabilis hawaiiensis*, also rarely found on mice and mongooses.

Mites found on rats include *Laelaps echidinus* and *L. hawaiiensis* (= *L. nuttalli*) (see Pemberton, 1943), *Radfordia ensifera* (see Joyce, 1958), and *Ornithonyssus bacoti* (see Haramoto, 1964), which are also capable of biting man.

Species of sucking lice found on rats include the spinulose rat louse, *Polyplax spinulosa*, and the Pacific rat louse, *Hoplopleura oenomydis* (see Zimmerman, 1948).

Of the above fleas, *X. cheopis* and *X. vexabilis hawaiiensis* are the principal vectors of bubonic plague (Eskey, 1934). *X. cheopis* and *N. fasciatus* are believed to be the principal vectors of endemic typhus (Doolittle, 1941); *E. gallinacea* also serves in the same capacity (Alicata, 1942a). The flea, *C. felis*, is an intermediate host for the tapeworm, *Dipylidium caninum* (see Hall, 1929), and for the subcutaneous tissue filarioid, *Dipetalonema* sp., of dogs (Newton and Wright, 1956).

**SHEEP**

**PROTOZOA**

No reports are available.

**ROUNDWORMS**

An examination of a group of sheep from the island of Kahoolawe revealed the following incidence of roundworms (Cuckler, 1943): stomach worms, *Haemonchus contortus*, in 6 of 15 examined, and *Trichostrongylus colubriformis*, in 3 of 10 examined; intestinal worms, *Cooperia punctata*, in 3 of 10 examined, and *Nematodirus spathiger*, in 1 of 10 examined.

**TAPEWORMS**

Tapeworms identified by the writer as *Moniezia expansa* have been found in the intestines of wild sheep. In the continental United States, this tapeworm is known to utilize several species of free-living oribatid mites of the genera *Galumna*, *Peloribates*, *Protoschelobates*, and *Oribatula* as intermediate hosts (Kates and Runkel, 1948). Members of the genera *Galumna* and *Protoschelobates* are known to occur in Hawaii (Jacot, 1934), and may serve as vectors under local conditions.

Unidentified larval tapeworms (cysticerci), probably those of *Taenia hydatigena*, attached to the liver and peritoneum of sheep, were noted by Julien (1947), federal meat inspector. The larvae of *T. hydatigena* reach maturity in the intestinal tract of dogs.
FLUKES

Specimens of liver flukes collected from sheep in Honolulu were submitted to the United States Bureau of Animal Industry in 1892 (Hall, 1936). These specimens, which originally were identified as *Fasciola hepatica*, are probably those of *F. gigantica*, since the former is only rarely found in the Islands.

ARTHROPODS

In the examination by Cuckler (1943) of 60 sheep from the island of Kaho'olawe, 43 harbored the spinose ear tick, *Otobius megnini*. Reports also indicate the occurrence of the "sheep tick," *Melophagus ovinus* (see Bryan, 1934; Muir, 1928); the "head maggot," *Oestrus ovis* (see Bryan, 1934); and the "Oriental blowfly," *Chrysomyia megacephala* (see Bryan, 1934). The sheep scab mite, *Psoroptes equi ovis*, has been reported from sheep in the Islands (Hawaii Dept. Agr. and Cons., 1960).

SWINE

PROTOZOA

At least four species of protozoa occur in swine in the Islands (Alicata, 1947). They are frequently the cause of dysentery, especially among young animals (fig. 34). Included are the coccidia, *Eimeria debliecki*, *E. scabra* (fig. 35), and *E. spinosa*, and the ciliate, *Balantidium coli*. Various forms of unidentified amoebae and flagellates of unknown pathogenicity are also frequently found in the feces of swine.

ROUNDWORMS

In 1938, an examination of the feces of 103 grown pigs from the islands of Oahu and Hawaii (Alicata, 1939b) revealed the following incidence of parasite eggs: *Ascaris lumbricoides*, 21 percent; *Oesophagostomum dentatum*, 32 percent; *Strongyloides ransomi*, 43 percent; *Trichuris trichiura*, 7 percent.

Adult roundworms which have been recovered at necropsy from swine include the following (Alicata, 1938d): stomach worms, *Ascarops strongylina* and *Hyostrongylus rubidus*; intestinal worms, *A. lumbricoides* (fig. 36), *T. trichiura* (fig. 37b), and *O. dentatum* (fig. 37a); kidney worms, *Stephanurus dentatus* (fig. 38b); lungworms (fig. 40a), *Choerogastropus pudendotectus* and *Metastrongylus elongatus*. Larvae of *Trichinella spiralis* (fig. 32a) also have been found encysted in the musculature of a domestic pig (Alicata, 1938e).
Figure 34. Two pigs of the same litter. Left, pig which had suffered from a severe case of experimental coccidiosis; right, control normal pig. (Original.)

Figure 35. Intestinal coccidia of swine. (After Henry, 1931, courtesy of University of California Press.)

FIGURE 37. a, Large intestine of pig showing adult nodular worms, *Oesophagostomum dentatum* (arrows) and nodules caused by them; b, adult whipworms, *Trichuris trichiura*, natural size. (After Schwartz, 1952, U. S. Department of Agriculture, Farmers’ Bulletin 1787.)
Figure 38. a, Swine heavily infected with kidney worms, *Stephanurus dentatus*; b, adult kidney worms, natural size. (a, After Alicata, 1953, courtesy of *American Journal of Veterinary Research;* b, original.)
The following species of roundworms collected by Lyman Nichols, wildlife biologist, Hawaii Department of Agriculture, and identified by the writer, have been found among wild pigs in Hawaii: lungworms, *Metastrongylus elongatus* and *Choerostrongylus pudendotectus*; stomach worms, *Ascarops strongylina* and *Physocophalus sexalatus*; intestinal worms, *Globocephalus urosubulatus* and *Oesophagostomum dentatum*; kidney worms, *Stephanurus dentatus*. Wild pigs are descendants of domestic stock which escaped and now roam wild in the mountains, swamps, and wastelands of the Islands.

Kidney worms and lungworms are most frequently present among hogs raised in open hog lots. According to Gooch (1952), veterinarian, Hawaii Department of Health, of 16,323 hogs slaughtered between January and April, 1952, on the island of Oahu, 3,466 (21.2 percent) harbored kidney worms. Furthermore, one of the slaughterhouses in Honolulu indicated that during 1947, hog livers valued at about $13,000 were condemned because of parasitism due largely to kidney worms (fig. 39) (Kartman and Alicata, 1948). A method of protecting swine against kidney worm infection by the use of polyborate as a larvicide was suggested by Alicata (1954).

**Figure 39.** Liver of swine showing lesions produced by the migratory, immature stages of the kidney worm, *Stephanurus dentatus*. (Original.)
Of the above roundworms, lungworms require earthworms as intermediate hosts. At least two species of unidentified earthworms recovered from hog lots around Honolulu were found by the writer to harbor infective lungworm larvae. It was reported (Williams, 1931) that in Hawaii there were about a dozen species of earthworms of the genus *Pheretima*. The stomach worms, *A. strongylina* and *P. sexalatus*, utilize one of various coprophagous beetles as intermediate hosts in the continental United States (Alicata, 1935); in Hawaii, coprophagous beetles possibly serve in this capacity.

Because of the occurrence of the first laboratory-proved case of human trichinosis in Hawaii in 1936, the writer, under the auspices of the Hawaii Department of Health, conducted a survey to determine the prevalence of trichina infection in nature. This survey revealed the following information: of 47 domestic hogs and 40 wild hogs examined from the island of Hawaii, 1 (2.1 percent) and 6 (15.0 percent), respectively, were found infected; of 2,130 rats and 70 mongooses examined, 57 (2.7 percent) and 15 (21.4 percent), respectively, harbored trichinae. No trichinae were observed in 92, 130, and 30 domestic hogs examined from the islands of Maui, Oahu, and Kauai, respectively. Of 1,904 rats and 22 mongooses examined on Maui, 1 (0.05 percent) and 2 (9.1 percent), respectively, were found infected. Of 352 and 601 rats examined from Oahu and Kauai, respectively, none were infected (Alicata, 1938e).

It is of interest to point out that, from 1936 to 1964, the Hawaii Department of Health reported 112 cases of human trichinosis in Hawaii. Most of the infected persons had eaten, or were suspected of having eaten, improperly cooked wild pork or products made from wild pork (Alicata, 1938e). According to records of the Hawaii Board of Agriculture and Forestry, during the 8-year period from 1933 through 1940 inclusive (Tinker, 1941), 32,724 wild hogs, or an average of 4,090 a year, were killed on five of the larger islands. Because of the moderately high incidence of trichinosis in wild hogs, meat from these animals constitutes a health menace unless proper precautions are taken in cooking, preservation, and refrigeration. Of 133 human diaphragms examined at autopsy in Honolulu, 7.4 percent harbored trichina larvae (Alicata, 1942b) (see Parasites of Man).

**TAPEWORMS**

No adult tapeworms are present in swine. However, the infective larval stage (cysticercus) of *Taenia hydatigena* (fig. 40b) has been observed attached to the liver and omentum of domestic swine (Alicata, 1938d) and wild pigs. These larvae are known to reach maturity in the intestinal tract of dogs.

In 1954, a case of human cysticercosis (infection with the larval stage of *Taenia solium*) of unknown origin was reported in Honolulu in a person
Figure 40. Parasites of swine. a, adult lungworms in lower portion of lung, natural size; b, infective bladderworm stage of the dog tapeworm, *Taenia hydatigena*, attached to omentum of swine, natural size. (a, After Schwartz, 1952, U. S. Department of Agriculture, Farmers' Bulletin 1787; b, original.)
who had never resided away from the Islands (see Parasites of Man). Since this form of infection usually results from the ingestion of food or water contaminated with the eggs of the parasite, it implies that the adult parasite occurs in man in Hawaii. Man acquires the adult parasite from eating raw or undercooked pork containing the infective bladderworm stage. No record is available of the occurrence of porcine cysticercosis in Hawaii.

FLUKES

Flukes are not known to occur in domestic hogs in Hawaii. However, the liver fluke of cattle, presumably *Fasciola gigantica*, has been reported from wild pigs (Shipley, 1913).

ARTHROPODS

The hog mange mite, *Sarcoptes scabiei suis*, is present among domestic and wild pigs in the Islands (fig. 41). The sucking louse, *Haemopinus suis*, is also present (Illingworth, 1928b). Infestation with these ectoparasites is very often associated with malnutrition and unhygienic surroundings.

![Figure 41. Pig showing lesions of sarcoptic mange. (Original.)](image-url)
PARASITIC INFECTIONS OF MAN AND ANIMALS IN HAWAII

HOST LIST OF PARASITES AND INTERMEDIATE HOSTS
RECORDED FROM HAWAII

<table>
<thead>
<tr>
<th>NAME OF PARASITE</th>
<th>LOCATION IN HOST</th>
<th>INTERMEDIATE HOST*</th>
</tr>
</thead>
<tbody>
<tr>
<td>&quot;AMAKIHI&quot; (Chlorodrepanis virens)</td>
<td></td>
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</tr>
<tr>
<td>Machaerillaenus hawaiensis</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Philopterus macgregori</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>&quot;APAPANE&quot; (Himatione sanguinea)</td>
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<td></td>
</tr>
<tr>
<td>Myrsidea cyrtostigma</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>BARRED DOVE (Geopelia striata striata)</td>
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<td></td>
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<tr>
<td>(See DOVE, BARRED)</td>
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<td></td>
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<tr>
<td>BRAZILIAN CARDINAL (Paroaria cucullata)</td>
<td></td>
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<tr>
<td>(See CARDINAL, BRAZILIAN)</td>
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<tr>
<td>CALIFORNIA VALLEY QUAIL (Lophortyx californica vallicola)</td>
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<tr>
<td>(See QUAIL, CALIFORNIA VALLEY)</td>
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<tr>
<td>CARDINAL, BRAZILIAN (Paroaria cucullata)</td>
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<td></td>
</tr>
<tr>
<td>Myrsidea incerta</td>
<td>External</td>
<td>CAT (Felis domestica)</td>
</tr>
<tr>
<td>Roundworms:</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Aelurostrongylus abstrusus</td>
<td>Lungs</td>
<td>Gastropoda: Subulina octona,^2^ Bradybaena similis^2^ (rodents serve as paratenic hosts)</td>
</tr>
<tr>
<td>Anafilaroides rostratus</td>
<td>Lungs</td>
<td>Gastropoda: Subulina octona,^2^ Bradybaena similis^2^ (rodents serve as paratenic hosts)</td>
</tr>
<tr>
<td>Ancylostoma tubaeforme</td>
<td>Small intestine</td>
<td></td>
</tr>
<tr>
<td>Dirofilaria immitis</td>
<td>Heart and pulmonary artery</td>
<td>(See Parasites of Dog, p. 101)</td>
</tr>
</tbody>
</table>

*Legend: (^1^) = infection found in nature; (^2^) = determined experimentally; (^3^) = reported elsewhere for animals similar to those occurring in Hawaii.
<table>
<thead>
<tr>
<th>NAME OF PARASITE</th>
<th>LOCATION IN HOST</th>
<th>INTERMEDIATE HOST* (IF ANY)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Physaloptera praeputialis</td>
<td>Stomach</td>
<td>Orthoptera: Blatella germanica³</td>
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<tr>
<td>Toxocara canis</td>
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<tr>
<td>Tapeworms:</td>
<td></td>
<td></td>
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<tr>
<td>Dipylidium caninum</td>
<td>Small intestine</td>
<td>(See Parasites of Dog, p. 101)</td>
</tr>
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<td>Hydatigera taeniaeformis</td>
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<td></td>
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<td>Rattus rattus¹</td>
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<tr>
<td>Flukes:</td>
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<tr>
<td>Phagicola longus</td>
<td>Small intestine</td>
<td>Gastropoda (first intermediate host): unknown in Hawaii</td>
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<td></td>
<td>Teleostei (second intermediate host): unknown in Hawaii</td>
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<tr>
<td>Platynosomum fastosum</td>
<td>Liver</td>
<td>Gastropoda (first intermediate host): Subulina octona,² Bradybaena similaris²</td>
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<tr>
<td>Stellantchasmus falcatus</td>
<td>Small intestine</td>
<td>(See Parasites of Man, p. 107)</td>
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<td>Arthropods:</td>
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<td></td>
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<tr>
<td>Ctenocephalides felis</td>
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<td>Echidnophaga gallinacea</td>
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</tr>
<tr>
<td>Felicola subrostrata</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Notoedres cati</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>CATTLE (Bos taurus)</td>
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<tr>
<td>Protozoa:</td>
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</tr>
<tr>
<td>Eimeria bovis</td>
<td>Small intestine and cecum</td>
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<tr>
<td>Eimeria bukidnonensis</td>
<td>Intestine</td>
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<tr>
<td>Eimeria cylindrica</td>
<td>Intestine</td>
<td></td>
</tr>
<tr>
<td>Eimeria zurnii</td>
<td>Small and large intestines</td>
<td></td>
</tr>
</tbody>
</table>

* Legend: (¹) = infection found in nature; (²) = determined experimentally; (³) = reported elsewhere for animals similar to those occurring in Hawaii.
<table>
<thead>
<tr>
<th>NAME OF PARASITE</th>
<th>LOCATION IN HOST</th>
<th>INTERMEDIATE HOST* (IF ANY)</th>
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<tr>
<td><em>phlebotomum</em></td>
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<td></td>
</tr>
<tr>
<td><em>Cooperia pectinata</em></td>
<td>Small intestine</td>
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</tr>
<tr>
<td><em>punctata</em></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Cooperia spatulata</em></td>
<td>Small intestine</td>
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<tr>
<td><em>Dictyocaulus viviparous</em></td>
<td>Lungs</td>
<td>Coleoptera: <em>Aphodius lividus</em>,* Der* <em>mestes vulpinus</em></td>
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<tr>
<td><em>Gongylonema pulchrum</em></td>
<td>Esophagus</td>
<td><em>Orthoptera: Blatella germanica</em></td>
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<tr>
<td><em>Haemonchus contortus</em></td>
<td>Fourth stomach</td>
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</tr>
<tr>
<td><em>Oesophagostomum radiatum</em></td>
<td>Cecum and colon</td>
<td></td>
</tr>
<tr>
<td><em>Ostertagia ostertagi</em></td>
<td>Small intestine</td>
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<tr>
<td><em>Stephano</em> <em>flavia stilesi</em></td>
<td>Skin</td>
<td>Diptera: <em>Haematobia irritans</em></td>
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<td><em>Strongyloides papillosus</em></td>
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<tr>
<td><em>Trichostrongylus axei</em></td>
<td>Fourth stomach</td>
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<tr>
<td><em>Trichuris ovis</em></td>
<td>Cecum</td>
<td></td>
</tr>
<tr>
<td><strong>Tapeworms:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Moniezia benedeni</em></td>
<td>Small intestine</td>
<td>Acarina: Species of oribatid mites</td>
</tr>
<tr>
<td><em>Taenia saginata</em></td>
<td>Muscles</td>
<td>Artiodactyla: <em>Bos taurus</em> (cattle serve as intermediate host; man is the final host)</td>
</tr>
<tr>
<td>(cysticercus)</td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Flukes:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Fasciola gigantica</em></td>
<td>Liver</td>
<td>Gastropoda: <em>Fossaria ollula</em>,* Pseudosucinea columella*</td>
</tr>
<tr>
<td><em>Fasciola hepatica</em></td>
<td>Liver</td>
<td><em>Gastropoda: Fossaria ollula</em> (Unknown)</td>
</tr>
<tr>
<td>(Rumen fluke)</td>
<td>Rumen</td>
<td></td>
</tr>
<tr>
<td><strong>Arthropods:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Bovicola bovis</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Chrysomyia megacephala</em> (larvae)</td>
<td>In wounds and external</td>
<td></td>
</tr>
</tbody>
</table>

* Legend: (1) = infection found in nature; (2) = determined experimentally; (3) = reported elsewhere for animals similar to those occurring in Hawaii.
### NAME OF PARASITE  
**Location in Host**  
**Intermediate Host\* (if any)**

- *Chrysomyia rufifacies*  
  (larvae)  
  In wounds and external

- *Fannia sp.* (larvae)  
  External

- *Haematobia irritans*  
  External

- *Haematopinus eury sternus*  
  External

- *Hypoderma bovis* (larvae)  
  Under skin

- *Hypoderma lineatum* (larvae)  
  Under skin

- *Lucilia sericata* (?)  
  (see p. 55) (larvae)  
  External

- *Otobius megnini* (larvae and nymphs)  
  Ear canal

- *Stomoxys calcitrans*  
  External

---

**CHICKEN (Gallus gallus)**

**Protozoa:**

- *Eimeria tenella*  
  Cecum

- *Histomonas meleagridis*  
  Cecum and liver

**Roundworms:**

- *Ancylostoma hamulosa*  
  Gizzard

- *Ascariidae galli*  
  Small intestine

- *Dispherynx nasuta*  
  Proventriculus

**Amphipoda:**

- *Orchestia pluvialis*

**Coleoptera:**

- *Carpophilus dimidiatus*
- *Dactylo sternum abdominale*
- *Deremestes vulpinus*
- *Epiragus diremptus*
- *Euxestus sp.*
- *Gonocephalus serialum*
- *Litargus baleatus*
- *Oxydema fusiforme*
- *Palorus ratzbergi*
- *Sitophilus oryzae*
- *Tenebroides nana*
- *Tribolium castaneum*
- *Typhaea stercorea*

**Orthoptera:**

- *Atractomorpha ambigua*
- *Gonocephalus saltator*
- *Oxya chinensis*

---

*Legend: (1) = infection found in nature; (2) = determined experimentally; (3) = reported elsewhere for animals similar to those occurring in Hawaii.*
<table>
<thead>
<tr>
<th>Name of Parasite</th>
<th>Location in Host</th>
<th>Intermediate Host* (If Any)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gongylonema ingluvicola</td>
<td>Crop</td>
<td>(Unknown; probably coprophagous beetles)</td>
</tr>
<tr>
<td>Heterakis gallinarum</td>
<td>Cecum</td>
<td>Orthoptera: <em>Pycnoscelus surinamensis</em></td>
</tr>
<tr>
<td>Oxyspirura mansonii</td>
<td>Conjunctival sac</td>
<td>Coleoptera: <em>Amprobias diapinurus</em>, <em>A. insularis</em>, <em>Dermestes vulpinus</em>, <em>Gonocephalum seriatum</em>, <em>Tribolium castanum</em></td>
</tr>
<tr>
<td>Subulura brunpti</td>
<td>Cecum</td>
<td>Orthoptera: <em>Conocephalus saltator</em>, <em>Oxya chinensis</em></td>
</tr>
<tr>
<td>Tropisurus americanus</td>
<td>Proventriculus</td>
<td>Coleoptera: <em>Dendrophiulus sp.</em>, <em>Dermestes vulpinus</em>, <em>Epitragus divemtus</em>, <em>Gonocephalum seriatum</em></td>
</tr>
<tr>
<td>Tapeworms:</td>
<td></td>
<td>Orthoptera: <em>Blastella germanica</em>, <em>Conocephalus saltator</em>, <em>Euboreilla annulipes</em>, <em>Euboreilla annulipes</em>, <em>Orchestia platensis</em></td>
</tr>
<tr>
<td>Choanotaenia infundibulum</td>
<td>Small intestine</td>
<td>Coleoptera: <em>Dermestes vulpinus</em>, <em>Epitragus divemtus</em>, <em>Gonocephalum seriatum</em></td>
</tr>
<tr>
<td>Echinolepis cavioca</td>
<td>Small intestine</td>
<td>Diptera: <em>Musca domestica</em></td>
</tr>
<tr>
<td>Hymenosphenacanthus exigus</td>
<td>Small intestine</td>
<td>Coleoptera: <em>Aphodius granarius</em></td>
</tr>
<tr>
<td>Raillietina cesticillus</td>
<td>Small intestine</td>
<td>Diptera: <em>Stomoxys calcitrans</em></td>
</tr>
<tr>
<td></td>
<td></td>
<td>Amphipoda: <em>Orchestia platensis</em></td>
</tr>
<tr>
<td></td>
<td></td>
<td>Coleoptera: <em>Dermestes vulpinus</em>, <em>Gonocephalum seriatum</em></td>
</tr>
</tbody>
</table>

*Legend: (*) = infection found in nature; (2) = determined experimentally; (3) = reported elsewhere for animals similar to those occurring in Hawaii.
<table>
<thead>
<tr>
<th>NAME OF PARASITE</th>
<th>LOCATION IN HOST</th>
<th>INTERMEDIATE HOST* (IF ANY)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Raillietina tetragona</td>
<td>Small intestine</td>
<td>(Probably ants of the genera Pheidole and Tetramorium)³</td>
</tr>
<tr>
<td>Flukes:</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Postharmostomum gallinum</td>
<td>Cecum</td>
<td>Gastropoda: Bradybaena similaris,¹ Subulina octona¹</td>
</tr>
<tr>
<td>Arthropods:</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Chelopistes meleagris</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Cyclotogaster</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>heterographa</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Dermanyssus gallinae</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Echidnophaga gallinae</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Goniocotes gallinae</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Goniodes dissimilis</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Goniodes gigas</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Lipeurus caponis</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Megninia cubitalis</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Menacanthus stramineus</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Menopon gallinae</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Ornithonyssus busa</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Pterolichus obtusus</td>
<td>External</td>
<td></td>
</tr>
</tbody>
</table>

CHINESE DOVE (Streptopelia chinensis)  
(See DOVE, CHINESE)

CHINESE RING-NECKED PHEASANT (Phasianus colchicus torquatus)  
(See PHEASANT, CHINESE RING-NECKED)

COOT, HAWAIIAN (Fulica americana alai)

Flukes:  
Philophthalmus gralli | Eye | Gastropoda: Stenomelania newcombi,¹ Thiara granifera¹ |

Arthropods:  
Quadraceps oraria | External |
Rallicola advena | External |

* Legend: (¹) = infection found in nature; (²) = determined experimentally; (³) = reported elsewhere for animals similar to those occurring in Hawaii.
<table>
<thead>
<tr>
<th>NAME OF PARASITE</th>
<th>LOCATION IN HOST</th>
<th>INTERMEDIATE HOST* (IF ANY)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Roundworms:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Cooperia punctata</em></td>
<td>Small intestine</td>
<td>DEER (<em>Cervus axis</em>)</td>
</tr>
<tr>
<td><strong>Protozoa:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>(Coccidia of undetermined species)</td>
<td>Small intestine</td>
<td>DOG (<em>Canis familiaris</em>)</td>
</tr>
<tr>
<td><strong>Roundworms:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Ancylostoma caninum</em></td>
<td>Small intestine</td>
<td></td>
</tr>
<tr>
<td><em>Dipetalonema sp.</em></td>
<td>Subcutaneous tissue</td>
<td>Siphonaptera:</td>
</tr>
<tr>
<td><em>Dirofilaria immitis</em></td>
<td>Heart and pulmonary artery</td>
<td><em>Ctenocephalides felis</em></td>
</tr>
<tr>
<td><em>Filaroides osleri</em> (see p. 71)</td>
<td>Lungs</td>
<td><em>Diptera: Aedes aegypti</em></td>
</tr>
<tr>
<td><em>Toxascaris leonina</em></td>
<td>Small intestine</td>
<td><em>Culex pipiens</em></td>
</tr>
<tr>
<td><em>Toxocara canis</em></td>
<td>Small intestine</td>
<td><em>quinquefasciatus</em></td>
</tr>
<tr>
<td><em>Trichuris vulpis</em></td>
<td>Cecum</td>
<td>(Unknown; see p. 74)</td>
</tr>
<tr>
<td><strong>Tapeworms:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Dipylidium caninum</em></td>
<td>Small intestine</td>
<td>Siphonaptera:</td>
</tr>
<tr>
<td><em>Taenia hydatigena</em> (see p. 74)</td>
<td>Small intestine</td>
<td><em>Ctenocephalides felis,</em></td>
</tr>
<tr>
<td><em>Phagicola longus</em></td>
<td>Small intestine</td>
<td><em>Pulex irritans</em></td>
</tr>
<tr>
<td><em>Stellantchasmus falcatus</em></td>
<td>Small intestine</td>
<td><em>Trichodectes latus</em></td>
</tr>
<tr>
<td><strong>Flukes:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Phagicola longus</em></td>
<td>Small intestine</td>
<td>(See Parasites of Cat, p. 96)</td>
</tr>
<tr>
<td><em>Stellantchasmus falcatus</em></td>
<td>Small intestine</td>
<td>(See Parasites of Man, p. 107)</td>
</tr>
<tr>
<td><strong>Arthropods:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Ctenocephalides felis</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Echidnophaga gallinacea</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Heterodoxus longistarsus</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Linognathus setosus</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Rhipicephalus sanguineus</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Trichodectes canis</em></td>
<td>External</td>
<td></td>
</tr>
</tbody>
</table>

*Legend: (1) = infection found in nature; (2) = determined experimentally; (3) = reported elsewhere for animals similar to those occurring in Hawaii.*
<table>
<thead>
<tr>
<th>NAME OF PARASITE</th>
<th>LOCATION IN HOST</th>
<th>INTERMEDIATE HOST* (IF ANY)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Arthropods:</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Goniodes lativentris</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>DOVE (Species unrecorded)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Goniocotes armoredalus</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Goniocotes chinensis</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Goniodes sp.</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Menopon sp.</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Ornithonyssus bursa</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>DOVE, BARRED (Geopelia striata striata)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Columbicola columbae</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Arthropods:</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Roundworms:</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Oxyspirura mansoni</td>
<td>Eye</td>
<td>(See Parasites of Chicken, p. 99)</td>
</tr>
<tr>
<td>Arthropods:</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Columbicola columbae</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Goniocotes chinensis</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Myrsidea invadens</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Pterolichus sp.</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>DUCK (Anas boschas domesticus)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Menopon gallinae</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>DUCK, HAWAIIAN (Anas wyvilliana wyvilliana)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Tapeworms:</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cloacotaenia megalops</td>
<td>Cloaca</td>
<td>(Unknown)</td>
</tr>
<tr>
<td>ENGLISH SPARROW (Passer domesticus)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>(See SPARROW, ENGLISH)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

* Legend: (') = infection found in nature; (') = determined experimentally; (') = reported elsewhere for animals similar to those occurring in Hawaii.
<table>
<thead>
<tr>
<th>Name of Parasite</th>
<th>Location in Host</th>
<th>Intermediate Host* (if Any)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fowl, Guinea (Numida meleagris galeata)</td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Arthropods:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Goniodes gigas</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Menopon gallinae</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Menopon phaeostomiunum</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><strong>Arthropods:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Goniodes dissimilis</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Lipeurus caponis</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Menopon gallinae</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Ornithoica vicina</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><strong>Roundworms:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Haemonchus contortus</td>
<td>Stomach</td>
<td></td>
</tr>
<tr>
<td>Ostertagia circumcincta</td>
<td>Small intestine</td>
<td></td>
</tr>
<tr>
<td>Trichostrongylus colubriformis</td>
<td>Stomach and small intestine</td>
<td></td>
</tr>
<tr>
<td>Trichuris ovis</td>
<td>Cecum</td>
<td></td>
</tr>
<tr>
<td><strong>Arthropods:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Bovicola capræ</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Linognathus africanaus</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><strong>Gray-Backed Tern</strong> (Sterna luna)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>(See Tern, Gray-Backed)</td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Guinea Fowl</strong> (Numida meleagris galeata)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>(See Fowl, Guinea)</td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Hawaiian Coot</strong> (Fulica americana alai)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>(See Coot, Hawaiian)</td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Hawaiian Duck</strong> (Anas wyvilliana wyvilliana)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>(See Duck, Hawaiian)</td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Hawaiian Owl</strong> (Asio flammeus sandwichensis)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>(See Owl, Hawaiian)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

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<table>
<thead>
<tr>
<th>NAME OF PARASITE</th>
<th>LOCATION IN HOST</th>
<th>INTERMEDIATE HOST*</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>HERON, NIGHT (Nycticorax nycticorax)</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Flukes:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Centrocestus formosanus</td>
<td>Small intestine</td>
<td>(See Parasites of Rat, p. 113)</td>
</tr>
<tr>
<td>Haplorchis taichui</td>
<td>Small intestine</td>
<td>Gastropoda (first intermediate host): Thiara granifera¹</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Teleostei (second intermediate host): Gambusia affinis,¹ Mugil cephalus,¹ Xiphophorus helleri²</td>
</tr>
<tr>
<td>Haplorchis yokogawai</td>
<td>Small intestine</td>
<td>Gastropoda (first intermediate host): Stenomelania newcombi,¹ Thiara granifera¹</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Teleostei (second intermediate host): Clarias fuscus²</td>
</tr>
<tr>
<td><strong>HORSE (Equus caballus)</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Alfortia edentata</td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td>Cyathostomum asymmetricum</td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td>Cyathostomum bicoronatum</td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td>Cyathostomum calicatum</td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td>Cyathostomum cattinatum</td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td>Cyathostomum coronatum</td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td>Cyathostomum euproctum</td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td>Cyathostomum goldi</td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td>Cyathostomum insignne</td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td>Cyathostomum leptostomum</td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td>Cyathostomum longibursatum</td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td>Cyathostomum nassatum</td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td>Cyathostomum pateratum</td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td>Delafondia vulgaris</td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td>Gyalcephalus capitatus</td>
<td>Large intestine</td>
<td></td>
</tr>
</tbody>
</table>

*Legend: ¹ = infection found in nature; ² = determined experimentally; ³ = reported elsewhere for animals similar to those occurring in Hawaii.
### Parasitic Infections of Man and Animals in Hawaii

<table>
<thead>
<tr>
<th>Name of Parasite</th>
<th>Location in Host</th>
<th>Intermediate Host*</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Habronema microstoma</em></td>
<td>Stomach</td>
<td>Diptera: <em>Musca domestica</em>, <em>Stomoxys calcitrans</em></td>
</tr>
<tr>
<td><em>Habronema muscae</em></td>
<td>Stomach</td>
<td>Diptera: <em>Musca domestica</em></td>
</tr>
<tr>
<td><em>Oxyuris equi</em></td>
<td>Colon</td>
<td></td>
</tr>
<tr>
<td><em>Parascaris equorum</em></td>
<td>Small intestine</td>
<td></td>
</tr>
<tr>
<td><em>Poteriostomum imparidentatum</em></td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td><em>Probstmayria vivipara</em></td>
<td>Colon</td>
<td></td>
</tr>
<tr>
<td><em>Strongylus equinus</em></td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td><em>Trichostongylus axei</em></td>
<td>Stomach</td>
<td></td>
</tr>
<tr>
<td><em>Triodontophorus brevicauda</em></td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td><em>Triodontophorus serratus</em></td>
<td>Large intestine</td>
<td></td>
</tr>
</tbody>
</table>

**Tapeworms:**

- *Anoplocephala magna* Small intestine Acarina: species of oribatid mites
- *Anoplocephala perfoliata* Cecum and near ileocecal valve Acarina: species of oribatid mites

**Flukes:**

- *Fasciola gigantica* (see p. 76) Liver (See Parasites of Cattle, p. 97)

**Arthropods:**

- *Bovicola equi* External
- *Gasterophilus intestinalis* (larvae) Stomach
- *Gasterophilus nasalis* (larvae) Stomach
- *Haematopinus asini* External
- *Stomoxys calcitrans* External

"HWI" (*Vestiaria cocinea*)

**Arthropods:**

- *Colpocephalum hilensis* External
- *Myrsidea cyrtostigma* External

**Japanese Quail** (*Coturnix coturnix japonica*) *(See Quail, Japanese)*

**Japanese White-eye** (*Zosterops palpebrosus japonicus*) *(See White-eye, Japanese)*

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<table>
<thead>
<tr>
<th>NAME OF PARASITE</th>
<th>LOCATION IN HOST</th>
<th>INTERMEDIATE HOST*</th>
</tr>
</thead>
<tbody>
<tr>
<td>JUNGLE FOWL (Gallus gallus) (See FOWL, JUNGLE)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>LINNET (Carpodacus mexicanus obscurus)</td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Arthropods:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Colpocephalum discrepans</td>
<td>External</td>
<td>(IF ANY)</td>
</tr>
<tr>
<td>Philopterus subflavescens</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><strong>Protozoa:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Dientamoeba fragilis</td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td>Endolimax nana</td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td>Entamoeba hartmanni</td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td>Entamoeba histolytica</td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td>Giardia lamblia</td>
<td>Small intestine</td>
<td></td>
</tr>
<tr>
<td>Pentatrichomonas hominis</td>
<td>Cecum and colon</td>
<td></td>
</tr>
<tr>
<td><strong>Roundworms:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Angiostrongylus cantonensis (developmental stages)</td>
<td>Brain</td>
<td>(See Parasites of Rat, p. 112)</td>
</tr>
<tr>
<td>Ascaris lumbricoides</td>
<td>Small intestine</td>
<td></td>
</tr>
<tr>
<td>Capillaria hepatica</td>
<td>Liver</td>
<td></td>
</tr>
<tr>
<td>Enterobius vermicularis</td>
<td>Cecum and adjacent portions</td>
<td></td>
</tr>
<tr>
<td>Strongyloides stercoralis</td>
<td>Small intestine</td>
<td></td>
</tr>
<tr>
<td>Toxocara canis (larvae)</td>
<td>In tissues</td>
<td></td>
</tr>
<tr>
<td>Trichinella spiralis</td>
<td>Adults in small intestine, larvae in muscles</td>
<td>(Same as final host)</td>
</tr>
<tr>
<td>Trichostrongylus colubriformis</td>
<td>Small intestine</td>
<td></td>
</tr>
<tr>
<td>Trichurus trichiura</td>
<td>Cecum</td>
<td></td>
</tr>
<tr>
<td><strong>Tapeworms:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Dipylidium caninum</td>
<td>Small intestine</td>
<td>(See Parasites of Dog, p. 101)</td>
</tr>
<tr>
<td>Hymenolepis nana</td>
<td>Small intestine</td>
<td>(See Parasites of Rat, p. 113)</td>
</tr>
<tr>
<td>Taenia saginata</td>
<td>Small intestine</td>
<td>Artiodactyla: Bos taurus¹</td>
</tr>
<tr>
<td>Taenia solium (larvae)</td>
<td>Brain and viscera</td>
<td>Artiodactyla: Sus scrofa domestica¹</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Primate: Homo sapiens¹</td>
</tr>
</tbody>
</table>

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### Parasitic Infections of Man and Animals in Hawai'i

<table>
<thead>
<tr>
<th>Name of Parasite</th>
<th>Location in Host</th>
<th>Intermediate Host* <em>(if any)</em></th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Flukes:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fasciola gigantica</td>
<td>Liver</td>
<td>(See Parasites of Cattle, p. 97)</td>
</tr>
<tr>
<td>Stellantchasmus falcatus</td>
<td>Small intestine</td>
<td>Gastropoda (first intermediate host): Stenomelania newcombi, Thuara granifera¹</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Teleostei (second intermediate host): Mugil cephalus¹</td>
</tr>
<tr>
<td><strong>Arthropods:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Aedes aegypti</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Aedes albopictus</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Aedes vexans nocturnus</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Cimex lectularius</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Ctenocephalides felis</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Culex pipiens</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>quinquefasciatus</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Demodex folliculorum</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Dermapthysus gallinae</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Echidnophaga gallinacea</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Laelaps echidninus</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Laelaps nuttalli</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Leptopsylla segnis</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Nosopsyllus fasciatus</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Oestrus ovis (larvae)</td>
<td>Body openings</td>
<td></td>
</tr>
<tr>
<td>Ornithonyssus bacoti</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Ornithonyssus bursa</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Pediculus humanus</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>capitis</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pediculus humanus humanus</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Phthirus pubis</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Pulex irritans</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Pyemotes boylei</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Rhipicephalus sanguineus</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Stomoxys calcitrans</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Triatoma rubrofasciata</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Xenopsylla cheopis</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Xenopsylla vexabilis hawaiensis</td>
<td>External</td>
<td></td>
</tr>
</tbody>
</table>

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**NAME OF PARASITE** | **LOCATION IN HOST** | **INTERMEDIATE HOST***
---|---|---
**MONGOOSE (Herpestes javanicus auropunctatus)**

**Nematodes:**

| **Trichinella spiralis** | Adults in small intestine, larvae in muscles | (Same as final host) |

**Arthropods:**

| **Ctenocephalides felis** | External | |
| **Echidnophaga gallinacea** | External | |
| **Leptopsylla segnis** | External | |
| **Nosopsyllus fasciatus** | External | |
| **Notoedres cati** | External | |
| **Xenopsylla cheopis** | External | |
| **Xenopsylla v. vexabilis hawaiensis** | External | |

**MOUSE (Mus musculus)**

**Protozoa:**

| **Trypanosoma conorhini** | Blood | Heteroptera: Triatoma rubrofasciata¹ |

(see p. 78)

**Tapeworms:**

| **Hydatigera taeniaeformis** | Liver | (See Parasites of Cat, p. 96) |

(larvae)

**Arthropods:**

| **Ctenocephalides felis** | External | |
| **Leptopsylla segnis** | External | |
| **Listrophorus sp.** | External | |
| **Myobia musculi** | External | |
| **Myocoptes musculinus** | External | |
| **Nosopsyllus fasciatus** | External | |
| **Radfordia affinis** | External | |
| **Xenopsylla cheopis** | External | |
| **Xenopsylla v. vexabilis hawaiensis** | External | |

**MYNAH (Acridotheres tristis)**

**Roundworms:**

| **Microtetrameres sp.** | Proventriculus | (Unknown) |
| **Oxyspirura mansoni** | Eye | (See Parasites of Chicken, p. 99) |

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### Parasitic Infections of Man and Animals in Hawaii

<table>
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<tr>
<th>Name of Parasite</th>
<th>Location in Host</th>
<th>Intermediate Host* (If Any)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Arthropods:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Menacanthus spinosus</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Montesauria sp.</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Myrsidea invadens</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Ornithonyssus bursa</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Pteronyssus sp.</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Trouessartia sp.</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><strong>Protozoa:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Plasmodium vaughani</em></td>
<td>Blood</td>
<td><em>Passeriformes: Liothrix lutea</em> (parasites probably transmitted by mosquitoes)</td>
</tr>
<tr>
<td><strong>Birds:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>NIGHT HERON</strong> (Nycticorax nycticorax)</td>
<td><em>(See HERON, NIGHT)</em></td>
<td></td>
</tr>
<tr>
<td><strong>NIGHTINGALE, PEKIN</strong> (Liothrix lutea)</td>
<td><em>(See TERN, NODDY)</em></td>
<td></td>
</tr>
<tr>
<td><strong>OWL, HAWAIIAN</strong> (Asio flammeus sandwichensis)</td>
<td><em>(See PLOVER, PACIFIC GOLDEN)</em></td>
<td></td>
</tr>
<tr>
<td><strong>Arthropods:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Colpocephalum biachysomum</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><strong>PEKIN NIGHTINGALE</strong> (Liothrix lutea)</td>
<td><em>(See NIGHTINGALE, PEKIN)</em></td>
<td></td>
</tr>
</tbody>
</table>

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<thead>
<tr>
<th>NAME OF PARASITE</th>
<th>LOCATION IN HOST</th>
<th>INTERMEDIATE HOST* (IF ANY)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>PHEASANT (Phasianus sp.)</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Cyrnea graphophasiani</em></td>
<td>Gizzard</td>
<td>(Unknown in Hawaii; probably transmitted by several species of arthropods)</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>PHEASANT, CHINESE RING-NECKED (Phasianus colchicus torquatus)</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Acuaria hamulosa</em></td>
<td>Gizzard</td>
<td><em>(See Parasites of Chicken, p. 98)</em></td>
</tr>
<tr>
<td><em>Ascaridia sp.</em> (galli?)</td>
<td>Small intestine</td>
<td></td>
</tr>
<tr>
<td><em>Heterakis gallinarum</em></td>
<td>Cecum</td>
<td><em>(See Parasites of Chicken, p. 99)</em></td>
</tr>
<tr>
<td><em>Oxyspirura mansoni</em></td>
<td>Eye</td>
<td><em>(See Parasites of Chicken, p. 99)</em></td>
</tr>
<tr>
<td><em>Subulura brumpti</em></td>
<td>Cecum</td>
<td></td>
</tr>
<tr>
<td><strong>Arthropods:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Falculifer rostratus</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Gabucinia sp.</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Goniocotes hologaster</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Goniodes colchici</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Goniodes mammillatus</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Lipeurus caponis</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Lipeurus maculosus</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Megninia columbae</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Megninia ginglymura</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Menopon</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>fulvomaculatum</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Ornithoica vicina</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Raphignathus sp.</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Uchida sp.</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><strong>PIGEON (Columba livia domestica)</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Haemoproteus columbae</em></td>
<td>Blood</td>
<td></td>
</tr>
</tbody>
</table>

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### Parasitic Infections of Man and Animals in Hawaii

**NAME OF PARASITE** | **LOCATION IN HOST** | **INTERMEDIATE HOST***
---|---|---
*Trichomonas gallinae* | Upper digestive tract |  
Roundworms:  
*Ornithostrongylus quadriradiatus* | Small intestine |  
Tapeworms:  
*Raillietina* sp. | Small intestine | (Unknown; probably species of arthropods)  
Arthropods:  
*Colpocephalum turbinatum* | External |  
*Columbicola columbace* | External |  
*Goniocotes bidentatus* | External |  
*Megninia columbace* | External |  
*Menopon gallinae* | External |  
*Pseudolynchia canariensis* | External |  

**Plover, Pacific Golden** (*Pluvialis dominica fulva*)

Arthropods:  
*Colpocephalum brachysomum* | External |  
*Quadraceps birostris* | External |  
*Saemundssonia conicus* | External |  

**Quail, California Valley** (*Lophortyx californica vallicola*)

Arthropods:  
*Echidnophaga gallinacea* | Attached to skin |  
*Goniodes mammillatus* | External |  
*Lagopoces docophoroides* | External |  
*Menopon fulvomaculatum* | External |  
*Xoloptes* sp. | External |  

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<table>
<thead>
<tr>
<th>NAME OF PARASITE</th>
<th>LOCATION IN HOST</th>
<th>INTERMEDIATE HOST*</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>QUAIL, JAPANESE (Coturnix coturnix japonica)</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Roundworms:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Microtetrameres</em> sp.</td>
<td>Proventriculus</td>
<td>(Unknown)</td>
</tr>
<tr>
<td>Oxyspirura mansoni</td>
<td>Eye</td>
<td>(See Parasites of Chicken, p. 99)</td>
</tr>
<tr>
<td><strong>Arthropods:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Goniocotes asterocephalus</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Ornithoica vicina</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><strong>RABBIT (Oryctolagus cunicularis)</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Protozoa:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Eimeria stiedae</td>
<td>Liver</td>
<td></td>
</tr>
<tr>
<td>Encephalitozoon cuniculi</td>
<td>Brain</td>
<td></td>
</tr>
<tr>
<td><strong>Roundworms:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Passalurus ambiguus</td>
<td>Cecum and large intestine</td>
<td></td>
</tr>
<tr>
<td><strong>Arthropods:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Notoedres cati cuniculi</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Psoroptes equi cuniculi</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><strong>RAT (Rattus exulans, R. norvegicus, R. rattus)</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><strong>Protozoa:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Trypanosoma conorhini (see p. 80)</td>
<td>Blood</td>
<td>Heteroptera: Triatoma rubrofasciata</td>
</tr>
<tr>
<td>Trypanosoma lewisi</td>
<td>Blood</td>
<td>Siphonaptera: Xenopsylla vexabilis hawaiensis</td>
</tr>
<tr>
<td><strong>Nematodes:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Angiostrongylus cantonensis</td>
<td>Lungs</td>
<td>Gastropoda: Achatina fulica,1 Bradybaena similis,1 Fossaria ollula,2 Subulina octona,1 Deroceras laeve,1 Opeas javanicum,1 Veronicaec alte1 (land planarians, Geoplan filiformis,2 calve,2 and swine,2 serve as paratenic hosts)</td>
</tr>
</tbody>
</table>

*Legend: (*) = infection found in nature; (2) = determined experimentally; (*) = reported elsewhere for animals similar to those occurring in Hawaii.*
<table>
<thead>
<tr>
<th>NAME OF PARASITE</th>
<th>LOCATION IN HOST</th>
<th>INTERMEDIATE HOST*</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Capillaria hepatica</em></td>
<td>Liver</td>
<td></td>
</tr>
<tr>
<td><em>Capillaria traverae</em></td>
<td>Small intestine</td>
<td></td>
</tr>
<tr>
<td><em>Gongylonema neoplasticum</em></td>
<td>Stomach</td>
<td></td>
</tr>
<tr>
<td><em>Heterakis spumosa</em></td>
<td>Large intestine</td>
<td>Orthoptera: <em>Blatella germanica</em>, <em>Periplaneta americana</em>, <em>Tenebrio molitor</em></td>
</tr>
<tr>
<td><em>Nippostrongylus brasiliensis</em></td>
<td>Small intestine</td>
<td></td>
</tr>
<tr>
<td><em>Physaloptera muris-brasiliensis</em></td>
<td>Stomach</td>
<td></td>
</tr>
<tr>
<td><em>Strongyloides ratti</em></td>
<td>Small intestine</td>
<td></td>
</tr>
<tr>
<td><em>Syphacia obvelata</em></td>
<td>Large intestine</td>
<td>Coleoptera: <em>Dermestes vulpinus</em>, <em>Tenebroides nana</em>, <em>Tribolium castaneum</em></td>
</tr>
<tr>
<td><em>Trichinella spiralis</em></td>
<td>Adults in small intestine, larvae in muscles</td>
<td>Orthoptera: <em>Nauphoeta Cinerea</em>, <em>Periplaneta americana</em></td>
</tr>
<tr>
<td><em>Trichosomoides crassicauda</em></td>
<td>Urinary bladder</td>
<td>(Same as final host)</td>
</tr>
</tbody>
</table>

Tapeworms:

*Hydatigera taeniaeformis* (larvae) Liver (See Parasites of Cat, p. 96)

*Hymenolepis diminuta* Small intestine Coleoptera: *Tenebrio molitor*, *Tribolium jerrugineum* Siphonaptera: *Myobia musculi*, *Pulex irritans*, *Xenopsylla cheopis*  

*Hymenolepis nana* Small intestine Coleoptera: *Tenebrio molitor* Siphonaptera: *Nosopsyllus fasciatus*, *Xenopsylla cheopis*

Flukes:

*Centrocestus formosanus* Small intestine Gastropoda (first intermediate host): *Stenomelania newcombi*, *Thiara granifera*

---

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<table>
<thead>
<tr>
<th>NAME OF PARASITE</th>
<th>LOCATION IN HOST</th>
<th>INTERMEDIATE HOST* (IF ANY)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Echinostoma</em> sp. (see p. 85)</td>
<td>Small intestine</td>
<td>Teleostei (second intermediate host): <em>Gambusia affinis</em>,¹ <em>Mugil cephalus</em>,¹ <em>Xiphophorus helleri</em>,¹ <em>Kuhlia sandwicensis</em>²</td>
</tr>
<tr>
<td><em>Plagiorchis muris</em></td>
<td>Small intestine</td>
<td>Gastropoda (first and second intermediate hosts): <em>Fossaria ollula</em>¹ (Unknown in Hawaii; but possibly fresh-water snails as first and second intermediate hosts, and aquatic larvae of midges also as second intermediate hosts)</td>
</tr>
<tr>
<td><em>Stellantchasmus falcatus</em></td>
<td>Small intestine</td>
<td><em>(See Parasites of Man, p. 107)</em></td>
</tr>
</tbody>
</table>

**Arthropods:**
- *Ctenocephalides felis* External
- *Echidnophaga gallinacea* External
- *Hoplopleura oenomydis* External
- *Laelaps echidninus* External
- *Laelaps nuttalli* External
- *Leptopsylla segnis* External
- *Nosopsyllus fasciatus* External
- *Ornithonyssus baroti* External
- *Polyplax spinulosa* External
- *Pulex irritans* External
- *Radfordia ensifera* External
- *Xenopsylla cheopis* External
- *Xenopsylla vexabilis hawaiensis* External

**RICEBIRD** (*Munia nisoria*)

**Arthropods:**
- *Bruelia stenozona* External
- *Philopterus subflavescens* External

**RUDDY TURNSTONE** (*Arenaria interpres interpres*) *(See TURNSTONE, RUDDY)*

---

*Legend: (¹) = infection found in nature; (²) = determined experimentally; (³) = reported elsewhere for animals similar to those occurring in Hawaii.*
<table>
<thead>
<tr>
<th>NAME OF PARASITE</th>
<th>LOCATION IN HOST</th>
<th>INTERMEDIATE HOST*</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>SHEEP (Ovis aries)</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Cooperia punctata</em></td>
<td>Small intestine</td>
<td></td>
</tr>
<tr>
<td><em>Haemonchus contortus</em></td>
<td>Fourth stomach</td>
<td></td>
</tr>
<tr>
<td><em>Nematodirus spathiger</em></td>
<td>Small intestine</td>
<td></td>
</tr>
<tr>
<td><em>Trichostrongylus colubriformis</em></td>
<td>Stomach and small intestine</td>
<td></td>
</tr>
<tr>
<td><strong>Tapeworms:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Moniezia expansa</em></td>
<td>Small intestine</td>
<td>Acarina: species of oribatid mites (See Parasites of Swine, p. 117)</td>
</tr>
<tr>
<td><em>Taenia hydatigena</em></td>
<td>Larval stage attached to liver, mesentery, and omentum</td>
<td></td>
</tr>
<tr>
<td><strong>Flukes:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Fasciola sp. (gigantica?)</em> (see p. 87)</td>
<td>Liver</td>
<td>(See Parasites of Cattle, p. 97)</td>
</tr>
<tr>
<td><strong>Arthropods:</strong></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Chrysomyia megacephala</em></td>
<td>In wounds and external</td>
<td></td>
</tr>
<tr>
<td><em>Melophagus ovinus</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Oestrus ovis</em> (larvae)</td>
<td>Nasal cavities and sinuses of head</td>
<td></td>
</tr>
<tr>
<td><em>Otobius megnini</em> (larvae and nymphs)</td>
<td>Ear canal</td>
<td></td>
</tr>
<tr>
<td><em>Psoroptes equi ovis</em></td>
<td>Skin</td>
<td></td>
</tr>
</tbody>
</table>

**SPARROW, ENGLISH (Passer domesticus)**

<table>
<thead>
<tr>
<th>NAME OF PARASITE</th>
<th>LOCATION</th>
<th>INTERMEDIATE HOST*</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Oxyspirura mansoni</em></td>
<td>Eye</td>
<td>(See Parasites of Chicken, p. 99)</td>
</tr>
<tr>
<td><em>Tropisurus sp.</em></td>
<td>Proventriculus</td>
<td>Orthoptera: Conocephalus saltator,² Oxya chinensis²</td>
</tr>
</tbody>
</table>

*Legend: (¹) = infection found in nature; (²) = determined experimentally; (³) = reported elsewhere for animals similar to those occurring in Hawaii.*
<table>
<thead>
<tr>
<th>NAME OF PARASITE</th>
<th>LOCATION IN HOST</th>
<th>INTERmediate Host* (IF ANY)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Arthropods:</td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Bruelia vulgata</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Haemolaelaps casalis</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Myrsidea sp.</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Neonyssus sp.</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Ornithoica vicina</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Ornithonyssus bursa</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td><em>Proctophyllodes truncatus</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>SWINE (Sus scrofa domestica)</td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Balantidium coli</em></td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td><em>Eimeria debliecki</em></td>
<td>Small and large intestines</td>
<td></td>
</tr>
<tr>
<td><em>Eimeria scabra</em></td>
<td>Intestine</td>
<td></td>
</tr>
<tr>
<td><em>Eimeria spinosa</em></td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td>Roundworms:</td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Ascaris lumbricoides</em></td>
<td>Small intestine</td>
<td></td>
</tr>
<tr>
<td><em>Ascarops strongylina</em></td>
<td>Stomach</td>
<td>Coleoptera: (probably coprophagous beetles)</td>
</tr>
<tr>
<td><em>Choeronyssus pudendotectus</em></td>
<td>Lungs</td>
<td>Oligochaeta: (earthworms, probably of the genus Pheretima)</td>
</tr>
<tr>
<td><em>Globocephalus urosubulatus</em> (see p. 91)</td>
<td>Small intestine</td>
<td></td>
</tr>
<tr>
<td><em>Hyostrongylus rubidus</em></td>
<td>Stomach</td>
<td></td>
</tr>
<tr>
<td><em>Metastrongylus elongatus</em></td>
<td>Lungs</td>
<td>Oligochaeta: (earthworms, probably of the genus Pheretima)</td>
</tr>
<tr>
<td><em>Oesophagostomum dentatum</em></td>
<td>Large intestine</td>
<td></td>
</tr>
<tr>
<td><em>Physostomus sexalatus</em></td>
<td>Stomach</td>
<td>Coleoptera: (probably coprophagous beetles)</td>
</tr>
<tr>
<td><em>Stephanurus dentatus</em></td>
<td>Adults in kidneys and kidney fat; immature forms in liver and other internal organs</td>
<td></td>
</tr>
</tbody>
</table>

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<table>
<thead>
<tr>
<th>NAME OF PARASITE</th>
<th>LOCATION IN HOST</th>
<th>INTERMEDIATE HOST* (IF ANY)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Strongyloides ransomi</td>
<td>Small intestine</td>
<td>(Same as final host)</td>
</tr>
<tr>
<td>Trichinella spiralis</td>
<td>Adults in small intestine; larvae in muscles</td>
<td></td>
</tr>
<tr>
<td>Trichuris trichiura</td>
<td>Cecum</td>
<td></td>
</tr>
<tr>
<td>Tapeworms:</td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Taenia hydatigena</em> (cysticercus)</td>
<td>Attached to liver, mesentery, and omentum</td>
<td>Artiodactyla: <em>Sus scrofa domestica</em>, Ovis aries</td>
</tr>
<tr>
<td><em>Taenia solium</em> (cysticercus, see p. 37)</td>
<td>Larvae in musculature and viscera</td>
<td>Artiodactyla: <em>Sus scrofa domestica</em>, Primate: Homo sapiens</td>
</tr>
<tr>
<td>Flukes:</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Fasciola sp. (gigantica?)</td>
<td>Liver</td>
<td>(See Parasites of Cattle, p. 97)</td>
</tr>
<tr>
<td>Arthropods:</td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Haematopinus suis</em></td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Sarcoptes scabiei suis</td>
<td>External</td>
<td></td>
</tr>
</tbody>
</table>

**TATTLER, WANDERING (Heteroscelus incanus)**

Arthropods:

*Actornithophilus kilauensis* External

*Saemundssonia conicus* External

**TERN, GRAY-BACKED (Sterna lunata)**

Arthropods:

*Quadraceps birostris* External

*Saemundssonia snyderi* External

**TERN, NODDY (Anous stolidus pileatus)**

Arthropods:

*Actornithophilus epiphanes* External

*Colpocephalum discrepans* External

---

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<table>
<thead>
<tr>
<th>Name of Parasite</th>
<th>Location in Host</th>
<th>Intermediate Host* (if Any)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Quadraceps separata</td>
<td>External</td>
<td>TURKEY (Meleagris gallopavo)</td>
</tr>
<tr>
<td>Histomonas meleagrisidis</td>
<td>Cecum and liver</td>
<td></td>
</tr>
<tr>
<td>Acuaria hamulosa</td>
<td>Gizzard</td>
<td>(See Parasites of Chicken, p. 98)</td>
</tr>
<tr>
<td>Heterakis gallinarum</td>
<td>Cecum</td>
<td></td>
</tr>
<tr>
<td>Chelopistes meleagrisidis</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Goniocotes hologaster</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Menopon gallinæ</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Oxylipeurus polytrapezius</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>TURNSTONE, RUDDY (Arenaria interpres interpres)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Austrobilharzia variglandis</td>
<td>Mesenteric veins</td>
<td>Gastropoda: Littorina pintado¹</td>
</tr>
<tr>
<td>WANDERING TATTLER (Heteroscelus incanus) (See TATTLER, WANDERING)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>WHITE-EYE, JAPANESE (Zosterops palpebrosus japonicus)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Dermoglyphus elongatus</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Megninia sp.</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Ornithoica vicina</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Pteronyssus sp. (?)</td>
<td>External</td>
<td></td>
</tr>
<tr>
<td>Trouessartia sp.</td>
<td>External</td>
<td></td>
</tr>
</tbody>
</table>

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BAKER, E. W.

BECKER, WILLARD W.
BESS, H. A.

BIANCHI, F. H.

BINFORD, C. H.

BONNET, D. D.

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BUTLER, G. D., JR.

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Price, A. S.

Punyagupta, S.

Roby, T. O.

Rosen, L.

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Schware, C. W.

and


Seneviratna, P.

Sherman, M.

Shipley, A. E.

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Zimmerman, E. C.

INDEX

Acarus siro, 42
Achatina fulica, 26, 36, 82, 112
Achiterna, 76
Acridothrix tristis, 108
Actornithophilus epiphanes, 70, 117
Actornithophilus kilauensis, 70, 117
Acutaria hamulosa, 12, 59, 60, 61, 98, 110, 118
Acuariidae, 12
Aedes aegypti, 42, 43, 74, 101, 107
Aedes albopictus, 42, 74, 107
Aedes scutellaris pseudoscutellaris, 28
Aedes vexans nocturnus, 42, 74, 107
Aelurostrongylus abstrusus, 12, 24, 43, 95
Alfortia edentata, 11, 75, 104
Alphitobius diaperinus, 63, 99
Amakihi, parasites of, 69, 95
Ananophorus insularis, 63, 99
A. naplasmorbostrongylus rostratus, 12, 24, 43, 44, 95
A. naplasmorbostrongylus marginale, 46
Anaplasmosis, 46
Anas boschas domestica, 102
Anas wyvilliana wyvilliana, 102
Ancylostoma caninuln, 10, 23, 43, 71, 101
Ancylostoma tubaeforrmes, 10, 43, 44, 95
Ancylostomatidae, 10
Ancylostomatinae, 10
Aneurysm, 76
Angiostrongylosis, 7, 31-36
Angiostrongylus cantonel1sis, 7, 12, 24, 26, 31-36, 80-83, 106, 112
Anolis cristatellus, 46
Anoplocephala magna, 13, 76, 77, 105
Anoplocephala perfoliata, 13, 76, 103
Anoplocephalidae, 13
Anoplocephalinae, 13
Anous stolidus pileatus, 117
Artemia, 65
Apapane, parasites of, 69, 95
Aphodius granarius, 65, 99
Aphodius lividus, 49, 97
Apis mellifera, 42
Arenaria interpres interpres, 67, 118
Arenaria interpres morinella, 41
Arthropoda, 15
Arthropods, life cycle of, 22
Arythmorhynchus, 43
Ascaris lumbricoides, 9, 27, 28, 87, 89, 106, 115
Ascarops strongylina, 12, 87, 91, 116
Ascaropsinae, 12
Ascomycotinae, 14
Asia flammaeus sandwichensis, 109
Atractomorpha ambigua, 60, 98
Attepholaelaps megacentralis (see Hymenoleps casalis)
Austrobilharzia variglandis, 14, 25, 41, 67, 118
Balantidiidae, 9
Balantidium coli, 9, 16, 87, 116
Bandicota pudica nemorivaga, 84
Barred dove (see Dove)
Beetle, coprophagous, 99
Blackhead, 59, 64
Blatella germanica, 43, 49, 65, 82, 96, 97, 99, 113
Bos taurus, 96, 97, 106
Bovicola bivis, 57, 97
Bovicola caprae, 75, 103
Bovicola equi, 76, 105
Brachylaenidae, 14
Brachylaeninae, 14
Bradybaena similis, 24, 44, 66, 82, 95, 96, 100, 112
Brazilian cardinal (see Cardinal)
Brudaria stenozona, 70, 114
Brudaria vulgata, 69, 116
Brugia malayi, 28
Bubonic plague, 43
Bufo martius, 63
Bulinus robustus minor, 41
Bunostominae, 10
Bunostomum phlebotomum, 10, 47, 48, 19, 97
California valley quail (see Quail)
Canis familiaris, 101
Capillaria caudinflata, 18
Capillaria hepatica, 7, 10, 29, 30, 80, 81, 106, 113
Capillaria traversae, 10, 80, 113
Capillarinae, 10
Capra hircus, 103
Cardinal, Brazilian, parasites of, 70, 95
Carposobus mexicanus obscurus, 106
Carposobus dimidiatus, 60, 98
Cat, parasites of, 43, 95
Cattle, parasites of, 46, 96
Centropus, 43
Centrocestinae, 14
Centrocestus formosanus, 14, 25, 39, 67, 104, 113
Cerebral angiostrongylosis, 24, 26, 31, 33, 84
Cerebral cysticercosis, 33
Cerebral echinococcosis, 33
Cerebral paragonimiasis, 33
Cerebral schistosomiasis, 33
Cerus eexs, 101
Cestoda, 33
Cheiracanthium diversum, 42
Chelopistes Meleagris, 69, 100, 118
Chicken, parasites of, 57, 58, 65, 98
Chilomastix mesnili, 16
Chinese dove (see Dove)
Chinese ring-necked pheasant (see Pheasant)
Chlorodrepanis virens, 95
Choanotaenia infundibulum, 13, 65, 66, 99
Choerostrongylus pudendotectus, 12, 87, 91, 116
Chrysornia megacephala, 55, 87, 97, 115
Chrysornia rufifacies, 55, 98
Ciliatea, 9
Cinlex lectorius, 42, 107
Clarias fuscus, 39, 10
Cloacotaenia megalops, 13, 65, 102
Clonorchis sinensis (see Opisthorchis sinensis)
Coccidiosis, 57, 58, 71, 88, 101
Colpoccephalum brachysomum, 70, 109, 111
Colpoccephalum discrepans, 70, 106, 117
Colpoccephalum hilensis, 70, 105
Colpoccephalum turbinatum, 69, 142
Columba livia, 65
Colurnba Livia domestica, 110
Colurnbicola columbae, 69, 70, 102, 149
Conocephalus saltator, 60, 63, 65, 98, 99, 115
Cooperia pectinata, 11, 48, 97
Cooperia punctata, 11, 47, 48, 49, 71, 86, 97, 101, 115
Cooperia spatulata, 11, 49, 97
Copris incertus, 64
Copris minutus, 64
Coturnix coturnix japonica, 112
Ctenocephalides canis, 74
Ctenocephalides felis, 42, 46, 74, 78, 79, 85, 96, 101, 107, 118
Culex pipiens quinquefasciatus, 28, 42, 74, 101, 107
Cyathostomum, 10
Cyathostomum asymetricum, 10, 75, 104
Cyathostomum bicoronatum, 10, 75, 104
Cyathostomum calicatum, 10, 75, 104
Cyathostomum catinatum, 10, 75, 104
Cyathostomum coronatum, 11, 75, 104
Cyathostomum endocticum, 11, 75, 104
Cyathostomum gordi, 11, 75, 104
Cyathostomum insigne, 11, 75, 104
Cyathostomum leptostomum, 11, 75, 104
Cyathostomum longibursatum, 11, 75, 104
Cyathostomum nassatum, 11, 75, 104
Cyathostomum pateratum, 11, 75, 104
Cyclophyllidea, 13
Cyrene graphophasianus, 12, 60, 110
Cysticercosis:
  bovine, 49
  human, 37
  porcine, 94
Dactylosternum abdominalis, 60, 98
Davainiidae, 13
Davaineinae, 13
Decr, parasites of, 71, 101
Delafondia vulgaris, 11, 75, 76, 77, 104
Dendrolex folliculorum, 42, 107
Dendrophilus, 65
Dengue, 43
Dermatophagoides scheremetewskyi, 42
Dermestes vilpini, 49, 60, 63, 65, 82, 97, 98, 99, 113
Dermoglyphus elongatus, 70, 118
Deroceras laeve, 26, 82, 112
Dicrocoeliidae, 14
Dicrocoeliinae, 14
Dictyocaulinae, 12
Dictyocaulus vivit)arus, 12, 49, 97
Dientamoeba fragilis, 8, 16, 27, 106
Digenea, 14
Dilectidae, 13
Dipetalonemata, 13, 71, 101
Dipetalonema reconditum, 74
Dipetalonematinae, 13
Dipelostomina, 8
Dipylidiatidae, 13
Dipylidium caninum, 7, 13, 37, 42, 45, 71, 96, 101, 106
Dirofilaria immitis, 13, 43, 71, 72, 74, 95, 101
Dirofilaria immitis, 13, 43, 71, 72, 74, 95, 101
Dirofilaria immitis, 13, 43, 71, 72, 74, 95, 101
Dog, parasites of, 71, 101
Dog, parasites of, 69, 70, 102
Duck, Hawaiian, parasites of, 102
Duck, parasites of, 69, 70, 102
Duck, parasites of, 69, 70, 102
Earthworm, 92
Echinococcus grantulosus, 37
Echinocystis carinica, 65, 99
Echinostoma, 14, 85, 114
Echinostoma ilocanum, 83
Echinostomatidae, 14
Eimeria bovis, 8, 46, 96
I believe the text is a list of species names, possibly highlighting those found in Hawaii. Here is the text in plain text format:

- Eimeria bukidnonensis,
- Eimeria cylindrica,
- Eimeria debbiechi,
- Eimeria saobra,
- Eimeria spinosa,
- Eimeria stiedae,
- Eimeria tenella,
- Eimeria zurnii,
- Eimeria bukidnonensis,
- Eimeria cylindrica,
- Eimeria debbiechi,
- Eimeria saobra,
- Eimeria spinosa,
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- Eimeria cylindrica,
- Eimeria debbiechi,
- Eimeria saobra,
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- Eimeria zurnii,
- Eimeria bukidnonensis,
- Eimeria cylindrica,
- Eimeria debbiechi,
- Eimeria saobra,
Histomonas meleagridis, 8, 16, 59, 64, 98, 118
Holepyrus hawaiiensis, 42
Homo sapiens, 106, 117
Horoplopleura oenomydis, 86, 114
Horse, parasites of, 75, 104
Host List of parasites, 95
Hydatid, 37
Hydatigera taeniaeformis, 13, 45, 79, 81, 84, 96, 108, 113
Hymenolepis diminuta, 13, 82, 84, 113
Hymenolepis nana, 7, 13, 21, 36, 82, 84, 106, 113
Hymenolepis nana, 7, 13, 21, 36, 82, 84, 106, 113
Hymenolepididae, 13
Hymenolepidinae, 13
Hymenolepis diminuta, 82, 84, 113
Hymenolepis nana, 7, 13, 21, 36, 82, 84, 106, 113
Hymenophorus, 79, 108
Hyponema balteatus, 60, 98
Laeplus echininus, 42, 86, 107, 114
Laelaps hawaiiensis (see Laelaps nuttalli)
Laelaps nuttalli, 42, 86, 107, 114
Lagopoeus docophoroides, 69, 111
Lairdoetus geometricus, 42
Lairoductus naeivus, 42
Lauroiinae, 10
Leptopyjx segnis, 42, 78, 79, 85, 107, 108, 114
Lice (refer to Host List)
Linneut, parasites of, 70, 106
Linognathus africanus, 75, 103
Linognathus setosus, 75, 101
Liothrix lutea, 103
Liparurus caponis, 69, 100, 103, 110
Liparurus maculosus, 69, 110
Lithorhaphis, 79, 108
Litargus balleatus, 60, 98
Littorina pintado, 25, 41, 67, 118
Lizard, 46
Lophorhys californica varicola, 111
Lucilia sericata, 55, 98
Luciola cruciata, 53
Luciola lateralis, 53
Myrsemus viridis, 85
Machaerilaenus hawaiiensis, 69, 95
Macrobrachium, 31
Malaria, 6, 27, 57
Man, parasites of, 27, 106
Mastigamobidae, 8
Megninia, 70, 118
Megninia columbae, 69, 70, 110, 111
Menacanthus spinosus, 70, 109
Menacanthus stramineus, 69, 100
Menopon, 69, 102
Menopon fulcomaculatum, 69, 116, 111
Menopon gallinae, 69, 100, 102, 103, 111, 118
Menopon phaeostomum, 69, 103, 109
Metagonilninae, 14
Metastrongylinae, 12
Metastrongylus elongatus, 12, 87, 91, 116
Microsporida, 9
Microsporida, 9
Microtetrarines, 13, 65, 108, 112
Mites (refer to Host List)
Mollinesia formosus, 39
Mongoose, parasites of, 77, 108
Moniezia benedeni, 13, 47, 49, 97
Moniezia expansa, 13, 50, 86, 115
Moniliiformis moniliformis, 80
Monkey (see Simian primate)
Montesauria, 70, 109
Mosquitoes, 42, 74
Mouse, parasites of, 78, 108
Mus communis, 38, 39, 104, 107, 114
Munia nisoria, 114
Mys musculus, 96, 108
Mus maculipennis, 76, 99, 105
Mynah, parasites of, 62, 65, 70, 108
Myobia musculi, 79, 108, 113
Myocoptes musculus, 79, 108
Myrsidea, 116
Myrsidea cyrtostigma, 69, 70, 93, 105
Myrsidea incerta, 70, 93
Myrsidea invadens, 69, 70, 102, 109
Nasaepoa cinerea, 82, 113
Nemathelminthes, 9
Nematoloe, 9, 15
Nematodirinae, 11
Nematodirus spathiger, 11, 86, 115
Neoberrya, 70, 116
Night heron (see Heron)
Nightingale, Pekin, parasites of, 57, 109
Nipponstrongylus brasilienis, 11, 80, 113
Noddy tern (see Tern)
Nosophyllus fasciatus, 42, 43, 78, 79, 85, 107, 108, 113, 114
Notocerus cati, 46, 78, 80, 96, 108
Notocerus cati cuniculi, 112
Numida meleagris galeata, 103
Nystalorax nystalorax, 39, 104
Oesophagostominae, 11
Oesophagostomum dentatum, 11, 87, 89, 91, 116
Oesophagostomum radiatum, 11, 47, 49, 97
Oestrus ovis, 42, 47, 107, 115
Oepae beckianum, 84
Oepae clavulina, 84
Oepae goodalli, 84
Oepae javanicum, 24, 84, 112
Oepae mauritianum, 84
Oepae oparinum, 84
Oepae opella, 84
Opisthorchis sinensis, 39
Orchestia platensis, 60, 65, 98, 99
Oribatid mites, 49, 97, 105, 115
Oribatula, 86
Ornithostrongylinae, 11
Ornithostrongylus quaecliratnus, 11, 64, 118
Oryctolagus cunicularis, 112
Ostertagia circumcincta, 11, 75, 103
Ostertagia ostertagi, 11, 49, 97
Otohias megumi, 22, 56, 57, 87, 98, 115
Ovis aries, 115, 117
Owl, Hawaiian, parasites of, 70, 109
Oxya chinensis, 60, 62, 98, 99, 115
Oxydema tissiforme, 60, 98
Oxylipeurus polytrapezius, 69, 118
Oxyspirura mansoni, 12, 62, 99, 102, 108, 110, 112, 115
Oxyspirurinae, 12
Oxyuridae, 10
Oxyuridea, 9
Oxyurinae, 10
Oxyuris equi, 10, 75, 105

Pacific golden plover (see Plover)
Palorus ratzeburgi, 60, 98
Paragonimus westermani, 25, 41
Parasitic meningoecephalitis, 31
Parasites of Pukeke, 114
Passer domesticus, 115
Pavo cristatus, 109
Peafowl, parasites of, 109
Pediculoides ventricosus (see Pyemotes boylei)
Pediculus humanus capitis, 42, 107
Pediculus humanus humanus, 42, 107
Pekin nightingale (see Nightingale)
Peloribates, 86
Pentatrichomonas hominis, 8, 106
Periplaneta americana, 82, 113
Phagocila longus, 14, 45, 74, 96, 101
Phasianus, 110
Phasianus colchicus torquatus, 60, 110
Pheasant, Chinese ring-necked, parasites of, 60, 62, 63, 69, 70, 110
Pheasant, parasites of, 110
Pheidole, 65, 100
Pherecinita, 92, 116
Philophthalmidae, 14
Philophthalminae, 14
Philophthalmus grallii, 14, 25, 41, 68, 69, 100
Philophthalmus lacrymosus, 41
Philopterus macgregori, 69, 95
Philopterus subflavescens, 70, 106, 114
Phthirus pubis, 42, 107
Physaloptera muris-brasiilienis, 12, 80, 81, 82, 113
Physaloptera praepatialis, 12, 43, 44, 96
Physalopteridae, 12
Physalopteriniae, 12
Physocephalus sexalatus, 12, 91, 116
Pig, wild, 91
Pigeon, parasites of, 57, 58, 64, 65, 69, 110
Pigeon-fly, 16, 57, 70
Pla ampullacea, 31, 35
Pinworm, 27, 28
Plagiorchiidae, 14
Plagiorchiinae, 14
Plagiorchis muris, 14, 41, 85, 114
Plasmodiidae, 9
Plasmodium vaughani, 9, 57, 109
Platyhelminthes, 13
Platynosomum fiasosum, 14, 44, 45, 96
Plover, Pacific golden, parasites of, 70, 111
Pseudosuccinea columella, 24, 53, 97
Psoroptes equi, 80, 112
Psoroptes equi ovis, 87, 115
Pterolichus, 70, 102
Pterolichus obtusus, 70, 100
Pteronyssus, 70, 108, 118
Pulex irritans, 42, 85, 101, 107, 113, 114
Pycnoscelus surinamensis, 62, 99
Pyemotes boylei, 42, 107
Quadraceps birostris, 70, 111, 117
Quadraceps oraria, 70, 100
Quadraceps separata, 70, 118
Quail:
California valley, parasites of, 69, 70, 111
Japan, parasites of, 69, 60, 70, 119

Rabbit, parasites of, 69, 79, 112
Rafordia aflinis, 79, 108
Rafordia ensifera, 86, 114
Raillietina, 65, 111
Raillietina cesticillus, 13, 65, 99
Raillietina tetragona, 13, 65, 100
Rahaldia advena, 70, 100
Raphignathus, 69, 110
Rat, parasites of, 80, 112
Rattus exulans, 80, 112
Rattus norvegicus, 80, 96, 112
Rattus rattus, 80, 96, 112
Rhabdiasidea, 9
Rhipicephalus sanguineus, 22, 42, 75, 101, 107
Rhizolustigida, 8
Rhizopodea, 8
Ricebird, parasites of, 70, 114
Roundworms, life cycle of, 17
Ruddy turnstone (see Turnstone)
Rumen fluke, 50, 97

Saemundssonia conicus, 70, 111, 117
Saemundssonia snyderi, 70, 117
Sarcoptes scabiei suis, 94, 117
Scheloribates, 76
Scheloribates laevigatus, 49
Schistosoma japonicum, 23
Schistosoma mansoni, 23
Schistosomatidae, 14
Schistosomatinae, 14
Schistosomi, 6
Sciolonyza dorsata, 53
Sciolonyza flies, 53
Scolopendra subspinipes, 42
Sepsidion macroplus, 53
Sheep, parasites of, 86, 115
Simian primate, 33
Sitophilus oryzae, 60, 98
Solenops geminata, 42
Sparrow, English, parasites of, 62, 69, 70, 115
Spiruridae, 12
Spirurida, 12
Spirurinae, 12
Stagnicola emarginata angu'ata, 85
Stellantchamus falcatus, 14, 25, 37, 38, 40, 43, 74, 85, 96, 101, 107, 114
Stenomelania neowombi, 25, 38, 39, 41, 69, 100, 104, 107, 113
Stephanofilaria stilesi, 13, 48, 49, 57, 97
Stephanofofitariidae, 13
Stephanuridae, 14
Stephanurus dentatus, 11, 87, 90, 91, 116
Sterna luna, 117
Stomoxys calcitrans, 42, 57, 65, 76, 98, 99, 105, 107
Streptopelia chinensis, 65, 102
Strongylidae, 9, 10, 11
Strongyliace, 10
Strongyliinae, 11
Strongyloides papillosus, 9, 49, 97
Strongyloides ransomi, 9, 23, 87, 117
Strongyloides rattii, 9, 80, 113
Strongyloides stercoralis, 9, 23, 28, 106
Strongyloides westerni, 25
Strongyloidea, 9
Strongylus equinus, 11, 75, 77, 105
Subulina octona, 24, 43, 44, 66, 82, 95, 96, 100, 112
Subulura brumpti, 10, 63, 99, 110
Subuluridae, 10
Sus scrofa domestica, 106, 116, 117
Swine, parasites of, 87, 91, 116
Syphacia obvelata, 10, 80, 113
Syphaciainae, 10

Taenia hydatigena, 13, 74, 86, 92, 93, 101, 115, 117
Taenia saginata, 13, 36, 37, 49, 97, 106
Taenia solium, 7, 13, 30, 37, 106, 117
Taeniidae, 13
Tapeworms, life cycle of, 20
Tattler, wandering, parasites of, 70, 117
Telosporca, 8
Tenebrio molitor, 82, 85, 113
Tenebroides nana, 60, 82, 98, 113
Tern:
gray-backed, parasites of, 70, 117	noddy, parasites of, 70, 117
Tetrameris, 65
Tetranorum, 65, 100
Tetranorum carpitum, 65
Thelaziaidae, 12
Thiara granifera, 25, 38, 41, 69, 100, 104, 107, 113
Thrips, 42
Ticks (refer to Host List)
Toxascaris leonina, 9, 71, 101
Toxocara canis, 7, 9, 23, 29, 35, 48, 71, 73, 96, 10, 106
Toxocarinae, 9
Toxocarosis, 29
Toxoplasmatae, 9
Trematoda, 14, 15
Tritoma rubrofasciata, 42, 79, 80, 107, 108, 112
Tribolium castaneum, 60, 63, 82, 98, 99, 113
Tribolium confusum, 85
Tribolium ferrugineum, 113
Trichinella spiralis, 7, 10, 28, 77, 80, 82, 87
106, 108, 113, 117
Trichinellidae, 10
Trichinosis:
human, 28, 92
rodent, 80, 92
swine, 92
Trichodectes canis, 75, 101
Trichodectes latus, 74, 101
Trichomonadida, 10
Trichomonadidae, 10
Trichomonas gallinacea, 8, 58, 111
Trichomonas hominis, 27
Trichomoniasis, 58
Trichosomoides crassicauda, 10, 80, 81, 113
Trichosomoididae, 10
Trichostrongylidae, 11
Trichostrongylinae, 11
Trichostrongylus, 28
Trichostrongylus axei, 11, 49, 75, 97, 105
Trichostrongylus colubriformis, 11, 27, 75,
86, 103, 106, 115
Trichuriasis, 7
Trichuridae, 10
Trichuridea, 10
Trichurinae, 10
Trichuris asiatica, 10, 75, 97, 103
Trichuris trichiura, 10, 27, 28, 87, 89, 106,
117
Trichuris vulpis, 10, 71, 101
Triodontophorus brevicauda, 11, 75, 105
Triodontophorus serratus, 11, 75, 105
Tropisuridae, 13
Tropisurus, 13, 115
Tropisaurus americanus, 13, 64, 99
Trouessartia, 70, 109, 118
Trypanosoma conorhini, 8, 15, 78, 80, 108,
112
Trypanosoma lewisi, 8, 15, 16, 80, 112
Trypanosomatidae, 8
Turkey, parasites of, 58, 59, 64, 69, 118
Turnstone, ruddy, parasites of, 41, 118
Typhaea stercorea, 60, 98
Typhus, 45

Uchida, 69, 110

Verminous arthritis, 76
Veronicella alte, 26, 31, 84, 112
Vespa occidentalis, 42
Vesliaria coccinea, 105
Vianinae, 11
Visceral larva migrans, 29

Wandering tattler (see Tattler)
White-eye, Japanese, parasites of, 70, 118
Wild pig, parasites of, 91
Wuchereria bancrofti, 28

Xenopsylla cheopis, 42, 43, 78, 79, 85, 86,
107, 108, 113, 114
Xenopsylla vexabilis hawaiiensis, 42, 43,
78, 79, 80, 86, 107, 108, 112, 114
Xiphophorus helleri, 39, 104, 114
Xoloptes, 69, 111

Zoomastigophorea, 8
Zosterops, 70
Zosterops palpebratus japonicus, 118